

Calretinin Immunohistochemistery: An Aid in the Diagnosis of Hirschsprung's Disease

Mehran Hiradfar¹, Nourieh Sharifi², Mohammad Khajedaluee³, Nona Zabolinejad*², Shirin Taraz Jamshidi²

Abstract

Objective(s)

Definite diagnosis of Hirschsprung's disease (HD) is based on histopathological study, but there are limitations associated with standard histology and histochemistry in this regard. The aim of this study was to investigate calretinin immunostaining patterns in both ganglionic and aganglionic HD intestinal specimens and to compare them with control specimens.

Materials and Methods

Specimens included 30 patients with histopathologic diagnosis of HD and 20 patients that underwent colectomy for other reasons (as control group). Eighty paraffin wax blocks of full thickness intestinal specimens (30 blocks of ganglionic segments, 30 blocks of aganglionic segments and 20 blocks of control group) were studied. Calretinin immunoreactivity and pattern of staining for ganglion cells (nuclear and cytoplasmic) and also nerve fibers in different layers of bowel were evaluated in IHC stained slides.

Results

There were positive immunostaining of nerve fibers in the lamina propria, submucosa and muscularis propria in control and patient group. There were also nuclear and cytoplasmic staining of ganglion cells in submucosa and muscularis propria in all specimens of both control group (100%) and ganglionic segments (100%). Calretinin immunoexpression of nerve fibers in muscularis propria of the aganglionic segments was negative in all but two cases (6.7%). This method had sensitivity of 93.3% and specificity of 100% for diagnosis of HD in full thickness specimens of intestinal wall. The positive predictive value was 100% and negative predictive value was 93.8%.

Conclusion

Calretinin immunohistochemistry can be used on suction rectal biopsies as a reliable and adjunctive method to diagnose HD.

Keywords: Calretinin, Hirschsprung Disease, Immunohistochemistry

¹Department of Pediatric Surgery, Mashhad University of Medical Sciences, Mashhad, Iran

²Department of Pathology, Mashhad University of Medical Sciences, Mashhad, Iran

³Department of Community Medicine, Mashhad University of Medical Sciences, Mashhad, Iran

^{*}Corresponding author: Tel: +98-511-7269021; Fax: +98-511-7277470; email:zabolinejadn@mums.ac.ir

Introduction

aganglionic Hirschsprung's disease (HD, megacolon) is a common cause of congenital intestinal pseudo-obstruction occurring in 1 out of 5000 live births. It is characterized by the absence of intramural ganglion cells and the presence of excessive numbers of cholinergic nerve fibers (1, 2). Although anorectal manometry or radiologic studies are used to establish the diagnosis of HD, the definite diagnosis is made by histologic demonstration of agangliosis in both the myenteric (Auerbach) and submucous (Meissner) plexuses (3-5). Identification of ganglion cells in the myenteric plexus is easier, but needs a full-thickness rectal biopsy that requires a surgical procedure under general anesthesia and also is associated with some morbidity and technical difficulties. Rectal suction biopsy is increasingly becoming the procedure of choice for obtaining specimens for the initial diagnosis of HD (6). However, there are limitations to this technique such as morphologic immaturity of ganglion cells particularly in neonates and infants and also the need for evaluation of many sections (more than 50) before a biopsy can be interpreted as negative for ganglion cells which is a time consuming process (2). For these reasons a number of ancillary methods have been introduced to facilitate the diagnosis. The most widely applied technique is acetylcholinesterase histochemistry (AChE) (7-10). AChE stain shows an increase in coarse cholinergic nerve fibers in the muscularis mucosae, sometimes extending into the lamina propria. This technique requires frozen tissue samples and, there are false-negative results mostly due to the young age of patients (2, 8).

The aim of our study was to investigate calretinin immunostaining patterns in both ganglionic and aganglionic HD intestinal specimens and to compare them with control specimens. This study was done for the first time in Iran.

Materials and Methods

This is a retrospective study on specimens from patients with histopathologic diagnosis of HD

that underwent radical operation in Dr Sheikh Children Hospital, Mashhad University of Medical Sciences between 2008 and 2010.

For each patient one full thickness paraffin block from ganglionic and another block from aganglionic segment were retrieved.

In addition, 20 colon specimens (in which the diagnosis of HD was excluded based on H&E identification of ganglion cells) matched for age and sex, were used as control group.

In total, 80 paraffin blocks, previously fixed in 10% buffered formaline, were examined and processed. These comprised 30 blocks from the aganglionic zone, 30 from the ganglionic zone and 20 blocks from control group.

Immunostaining was performed on paraffin embedded archival tissues following the Avidin -Biotin peroxidase technique. In brief, sections measuring 3-5 µm thick were cut, air dried for 15 min, heat-fixed at 42 °C and then air-dried overnight at room temperature. After deparaffinization with xylene, endogenous peroxidase activity was eliminated by treating the slides with AOH/HO for 30 min at room temperature. Then the slides were incubated with the diluted primary antibody (Clone: DAK- Calret 1, monoclonal mouse anti-human, Dako, Denmark, 1/100 dilution) at 40 °C in a humidified chamber for 60 min. Biotinylated antimouse IgG and avidin-biotin peroxidase complex were added in sequence. The sections were then incubated with DAB for 10 min for visualization of the peroxidase reaction. After being washed in water for few minutes, the sections were counter stained with hematoxylin, dehydrated in alcohol, cleared in xylen and mounted.

Calretinin immunoreactivity and pattern of staining for ganglion cells (nuclear and cytoplasmic) and also nerve fibers in different layers of bowel (lamina propria, muscularis mucosa, submucosa and muscularis propria) were evaluated in IHC stained slides. Epithelium of plural tissue was selected as positive control, and antibody was omitted from staining process negative control. Two pathologists for independently reviewed the hematoxylin and eosin and immunohistochemichal stained slides and agreed on diagnoses by consensus.

Statistical calculations were carried out using SPSS version 11.5. For description of results in each group, methods of descriptive analysis such as central tendency, dispersion and frequency distribution were used as tables.

Results

Patient's age varied from 2 days to 12 years (mean, 17 ± 4 months) and male-to female ratio was 2.75.

Table 1. Prevalence of calretinin expression in nerve fibers of different layers of intestine in patients with HD^* and in control group

	Control group (20)	Patients (30)	
Layers of intestine	n (%)	Ganglionic bowels n (%)	Aganglionic bowels n (%)
Lamina propria	19 (95)	21 (70)	0 (0)
Muscularis mucosa	20 (100)	25 (83)	0 (0)
Submucosa	20 (100)	30 (100)	0 (0)
Muscularis propria	20 (100)	30 (100)	2 (6.7%)

*Hirschsprung disease



Figure 1. Positive calretinin immunostainig of nerve fibers in lamina propria (A), submucosa (B), and muscularis mucosa (C). Nuclear and cytoplasmic calretinin immunostainig of ganglion cells (D)

Layers of intestine	Types of HD				
	Short segment and classic		Long segment, extended and total colonic		
	Ganglionic	aganglionic	Ganglionic	aganglionic	
Lamina propria	n (%) 14 (70%)	n (%) 0 (0)	n (%) 7 (70)	n (%) 0 (0)	
Muscularis mucosa	16 (80%)	0 (0)	9 (90)	0 (0)	
Submucosa	20 (100%)	0 (0)	10 (100)	0 (0)	
Muscularis propria	20 (100%)	2 (6.7%)	10 (100)	0 (0)	

Table 2. Prevalence of calretinin expression in nerve fibers of different layers of intestine in patients with HD considering to extent of disease



Figure 2. Total absence of staining after calretinin immunohistochemistry in the aganlionic segment in lamina propria (A), submucosa (B) and muscularis propria (C)

There were 17 classic cases (56.7%), 7 long segment cases (23.3%), 3 short segment cases (10%), 2 extended cases (6.7%), and 1 case of total colonic aganglionosis (3.3%).

There were positive immunostaining of nerve fibers in the lamina propria, muscularis mucosa, submucosa and muscularis propria in control and patient groups (Figure1 & Table 1). This immunoexpression was present in different types of HD (with different extents of aganglionosis) (Table 2).

There were also nuclear and cytoplasmic staining of ganglion cells in submucosa and muscularis propria in all specimens of both control group (100%) and ganglionic bowels (100%) (Figure 1). The immunostaining of ganglion cells was specific, and no other cells were stained.

Calretinin immunoexpression was negative in all but two cases (6.7%) of aganglionic segments. One case was a 12-month-old boy with classic HD with pattern a of immunostaining in the ganglionic bowel similar control group. We found positive to

immunostaining only in hypertrophic nerve bundles of muscularis propria in the ganglionic segment (Figure 3). Another case was a19month-old girl with short segment HD that showed normal immunostaining in the ganglionic part, and positive immunostaining only in large nerves of muscularis propria of the aganglionic segment. So, this method had sensitivity of 93.3% and specificity of 100% for diagnosis of HD in full thickness specimens of intestinal wall. The positive predictive value was 100% and negative predictive value was 93.8%.

Discussion

Diagnosis of HD can be a stressful practice, particularly for pathologists who infrequently encounter the condition. Limitations associated with a H&E-based approach to the diagnosis of HD, with or without AChE histochemistry, urged us to use another reliable diagnostic method such as immunohistochemistry to facilitate diagnosis (9).



Figure 3. False positive immunostaining of nerve fibers in muscularis propria in aganglionic segment

Several immunohistochemical markers have been introduced, but most of them have limitations for use in daily practice (10).

Calretinin is a vitamin D-dependent calcium-binding protein involved in the physiological buffering of excess cytosolic calcium ions; calcium transport and protection against calcium ion overload (11).

McConalgue et al in 1994 showed that calretinin immunoreactivity reveals different neuronal populations in the large intestine of guinea-pig and after that this property has been shown for all ganglion cells and nerve fibers (12). In 2004 Barshack et al investigated calretinin immunorectivity in ten large bowels, full thickness specimens (a total 54 paraffin wax blocks) from patients with classic rectosigmoid HD. Calretinin was not expressed in aganglionic segments of HD and associated nerve fibers, whereas both ganglion cells and nerve fibers were immunopositive in ganglionic HD segments and in normal colons. The transitional showed broad spectrum zone a of histomorphological and immunohistochemical patterns of calretinin expression (13).

In another recent study by Kapur *et al* in 2009 multiple observers independently reviewed calretinin IHC and AChE sections of suction biopsies from 14 HD and 17 controls. There were 2 misdiagnoses and more disagreements in the interpretation of AChE-stained sections, but calretinin IHC yielded no

misdiagnoses or discrepancies. In Kapur's study, similar to ours, both nuclear and cytoplasmic immunoreactivity was present in ganglion cells and also there was positive staining of nerve fibers of lamina propria. and muscularis mucosa submucosa of ganglionic biopsies. They concluded that calretinin IHC can be a superior, alternative to AChE as an adjunctive diagnostic method for evaluating suction rectal biopsies for HD (14). It is interesting to note that Kapur et al found some immunoreactive axons in large nerves of the muscularis mucosa similar to what we found in 2 of our cases in large nerves of the muscularis propria. They stated that immunostaining in large submucosal nerves $(>20 \mu)$ in HD consists of puctate evenly like the spaced axons calretinin immunoreactivity in normal serosal nerves and is distinctly different from confluent granular staining of small-caliber nerves ($< 15 \mu$) in the normal superficial submucosa (14).

In another study in 2009, Guinard-Samuel *et al* evaluated the calretinin immunostaining as a primary diagnostic tool on a large series of suction rectal biopsies .They retrieved 131 biopsies carried out for suspicion of HD in children and infants to compare the accuracy of calretinin immunohistochemistry with the standard method (histology and acetylcholinesterase staining) (15).

their study. In calretinin immunohistochemistry enabled the diagnosis of all HD diagnosed by the standard technique, except for one patient who had a weak positive immunostaining in some nerve fibers (false negative case). It is important to note that 12 additional cases initially considered as suspicious for HD using the standard technique were accurately diagnosed by calretinin immunohistochemistry. They suggested that calretinin might be more accurate than acetylcholinesterase in detecting aganglionosis (15).

So far 3 previous studies have been published to evaluate calretinin IHC in diagnosis of HD (13-15). The number of cases were fewer than ours in two of these studies (9, 13, 14). Only in Guinard-Samuel's study a large series of rectal suction biopsies were used to assess the diagnostic value of calretinin IHC in the diagnosis of HD (15).We evaluated different layers of bowel (lamina propria, muscularis mucosa and muscularis propria) separately, and explored the results in cases with different extents of HD (although the number of cases in each type of HD were limited), which were not addressed in the previous studies.

Guinard-Samuel *et al* compared the accuracy of this technique with AChE staining (15). Although AChE staining has been introduced as an ancillary method in the diagnosis of HD, it has many limitations. Part of these limitations may be related to difficulties in performing and interpreting it even for the pathologists experienced in diagnosing HD (14, 16). Furthermore, presence of false-nagative results due to the young age of patients or long segments of aganglionosis can also limit its application in our daily practice (8, 15). On the other hand immunohistochemistry is based on paraffin sections and its interpretation which is based on negative or positive results is much easier (14, 15).

Nowadays, full thickness specimens are being replaced by suction or mucosal rectal biopsies in the diagnosis of HD. Interpretation is more difficult in these specimens. By facilitating the diagnosis, IHC staining may encourage more and more pathologists and surgeons to use suction biopsies instead of using the more invasive diagnostic technique of using full thickness specimens.

Conclusion

As there were no false- negative or false-positive results based on calretinin immunostaining in submucosa in our study, it can be used on suction rectal biopsies as a reliable and adjunctive method to diagnose HD.

Acknowledgment

The authors would like to thank Mashhad University of Medical Sciences, Mashhad, Iran, for their financial support (thesis No. 2244). The authors declare that they have no conflict of interests.

References

- 1. Haricharan RN, Georgeson KE. Hirschsprung disease. Semin Pediatr Surg 2008; 17:266-275.
- 2. Roberts DJ, Goldstein AM, Greame-Cook F, Dahms BB. Gastrointestinal tract and pancreas. In: Gilbert-Barness E, Kapur RP, Oligny LL, editors. Potter's Pathology of the fetus, infant and child. 2nd ed. Philadelphia: Mosby; 2007. p. 1176-1178.
- 3. Martucciello G, Pini Prato A, Puri P, Holschneider AM, Meier-Ruge W, Jasonni V, *et al.* Controversies concerning diagnostic guidelines for anomalies of the enteric nervous system: a report from the fourth International Symposium on Hirschsprung's disease and related neurocristopathies. J Pediatr Surg 2005; 40:1527-1531.
- 4. Noviello C, Cobellis G, Romano M, Amici G, Martino A. Diagnosis of Hirschsprung's Disease: an age-related approach in children below or above one year. Colorectal Dis 2010; 12:1044-1048.
- 5. De Lorijn F, Kremer LC, Reitsma JB, Benninga MA. Diagnostic tests in Hirschsprung disease: a systematic review. J Pediatr Gastroenterol Nutr 2006; 42:496-505.
- 6. Rahman N, Chouhan J, Gould S, Joseph V, Grant H, Hitchcock R, *et al.* Rectal biopsy for Hirschsprung's disease--are we performing too many? Eur J Pediatr Surg 2010; 20:95-97.
- 7. Meier-Ruge WA, Bruder E. Current concepts of enzyme histochemistry in modern pathology. Pathobiology 2008; 75:233-243.
- 8. Nakao M, Suita S, Taguchi T, Hirose R, Shima Y. Fourteen-year experience of acetylcholinesterase staining for rectal mucosal biopsy in neonatal Hirschsprung's disease. J Pediatr Surg 2001; 36:1357-1363.
- 9. Kapur RP. Practical pathology and genetics of Hirschsprung's disease. Semin Pediatr Surg 2009; 18:212-223.
- 10. Kapur RP. Can we stop looking? Immunohistochemistry and the diagnosis of Hirschsprung disease. Am J Clin Pathol 2006; 126:9-12.
- 11. Baimbridge KG, Celio MR, Rogers JH. Calcium-binding proteins in the nervous system. Trends Neurosci 1992; 15:303-308.
- 12. McConalogue K, Low AM, Williamson S, Bornstein JC, Furness JB. Calretinin-immunoreactive neurons and their projections in the guinea-pig colon. Cell Tissue Res 1994; 276:359-365.
- 13. Barshack I, Fridman E, Goldberg I, Chowers Y, Kopolovic J. The loss of calretinin expression indicates aganglionosis in Hirschsprung's disease. J Clin Pathol 2004; 57:712-716.

- 14. Kapur RP, Reed RC, Finn LS, Patterson K, Johanson J, Rutledge JC. Calretinin immunohistochemistry versus acetylcholinesterase histochemistry in the evaluation of suction rectal biopsies for Hirschsprung Disease. Pediatr Dev Pathol 2009; 12:6-15.
- 15. Guinard-Samuel V, Bonnard A, De Lagausie P, Philippe-Chomette P, Alberti C, El Ghoneimi A, *et al.* Calretinin immunohistochemistry: a simple and efficient tool to diagnose Hirschsprung disease. Mod Pathol 2009; 22:1379-1384.
- 16. Pacheco MC, Bove KE. Variability of acetylcholinesterase hyperinnervation patterns in distal rectal suction biopsy specimens in Hirschsprung disease. Pediatr Dev Pathol 2008; 11:274-282.