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Systematic enrichment analysis of microRNA expression profiling studies in endometriosis

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ARTICLE INFO	ABSTRACT
<i>Article type:</i> Review article	<i>Objective(s):</i> The purpose of this study was to conduct a meta-analysis on human microRNAs (miRNAs) expression data of endometriosis tissue profiles versus those of normal controls and to identify novel
<i>Article history:</i> Received: Jul 13, 2014 Accepted: Jun 8, 2015	 putative diagnostic markers. Materials and Methods: PubMed, Embase, Web of Science, Ovid Medline were used to search for endometriosis miRNA expression profiling studies of endometriosis. The miRNAs expression data were extracted, and study quality of each article was assessed. The frequently reported miRNAs with
<i>Keywords:</i> Endometriosis MicroRNAs Pathway analysis Profiling Target prediction	 consistent regulation were screened out by a meta-profiling algorithm. The putative targets of consistently expressed miRNAs were predicted by using four target prediction tools (TargetScan, PicTar, miRanda, miRDB), and gene ontology pathway enrichment analysis (KEGG and Panther pathways) of the miRNA targets were carried out with GeneCodis web tool. <i>Results:</i> A total of 194 related literatures were retrieved in four databases. One hundred and thirty four differentially expressed miRNAs were found in the 12 microRNA expression profiling studies that compared endometriosis tissues with normal tissues, with 28 miRNAs reported in at least two studies, and 9882 candidate genes retrieved for 13 consistently expressed miRNAs. Kyoto encyclopedia of genes and genomes (KEGG) and Panther pathways enrichment analysis showed that endometriosis related differently expressed miRNA targets were mainly enriched in cancer, endocytosis, Wnt signalling pathway, and angiogenesis. It showed that these differently expressed miRNAs and gene are potential biomarkers of endometriosis. <i>Conclusion:</i> miRNAs appear to be potent regulators of gene expression in endometriosis and its associated reproductive disorders, raising the prospect of using miRNAs as biomarkers and therapeutic agent in endometriosis.

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Introduction

MicroRNAs (miRNAs), a class of short noncoding RNA molecules, are proposed as promising biomarkers for early cancer detection and accurate prognosis, as well as targets for efficient treatment. Some miRNAs are post-transcriptional regulators of gene expression and implicated in central biological processes such as cell proliferation, differentiation and apoptosis (1, 2). Endometriosis is a common disease of reproductiveage women, which has complex pathogenesis. The lesions are extensive, highly invasive and recu-rrent, presenting malignant clinical behavior (3, 4). With the development of antisense technology and gene therapy, miRNA is expected to become a new strategy in endometriosis diagnosis and treatment (5, 6). It has been proved presently that miRNAs reside widely in eukaryotes, and are the largest gene family, participating and regulating various important life processes including cell differentiation, proliferation and apoptosis (7). As a method and result of the epigenetic modifications, latest research indicates that miRNA may play important role in the occurrence, development and prognosis of endometriosis, thus providing novel approach for the diagnosis and therapy of endometriosis. MiRNAs are key regulatory elements that control many genes expression and play crucial roles in many biological processes (8). Meta-analysis of mass miRNA expression profiling may uncover potential regulatory mechanisms by which microRNAs result in endometriosis.

Materials and Methods

Search strategies and study selection

PubMed (http://www.ncbi.nlm.nih.gov/pubmed), Embase (http://www.elsevier.com/online-tools/embase), Web of Science (http://thomsonreuters. com/thomson-reuters-web-of-science/), Ovid Medline (http://gateway.ovid.com) were used to search for endometriosis miRNA expression profiling studies published from January 2003 to October 2014 (last accessed on 20 October 2014), by means of the key words: endometriosis AND microRNA.

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Eligible studies had to meet the following criteria: (a) they were miRNA expression profiling studies in endometriosis patients; (b) they used tissue samples obtained from surgically resected endometriosis and corresponding eutopic or normal tissues for comparison; (c) use of miRNA microarray methods; (d) publishing a cut-off criteria of differentially expressed miRNAs. Therefore, the miRNA profiling studies using the serum samples of endometriosis patients or endometriosis cells in vitro were excluded. Review articles of miRNA expression profiles in endometriosis were also excluded. As repeated efforts can improve reliability and reduce error, valuable candidate miRNAs in this paper are defined as those validated and consistently reported by at least two studies.

Data abstraction

Two investigators (SW and YK) independently evaluated and extracted the data with the standard protocol and with all the discrepancies resolved by a third investigator (HX). From the full text and corresponding supplement information, the following eligibility items were collected and recorded for each study: author, journal and year of publication, location of study, selection and characteristics of recruited endometriosis patients, platform of miRNA expression profiling, author defined cut-off criteria of statistically differentially expressed miRNAs and the list of up- and downregulated miRNA features, and their corresponding fold change. Each included studies comparing miRNA expression between surgically resected endometriosis tissues and eutopic endometrium or normal tissues provided a list of differentially expressed miRNAs.

MiRNA target prediction and pathway enrichment analysis

Consensus targets were defined as genes predicted by at least two algorithms of four target prediction tools, including TargetScan, PicTar, miRanda, and miRDB. Although all three of these articles described the expression profile of miRNAs in endometriosis, they did not systematically predict the biological process and pathway of the identified miRNAs. Therefore, we used GeneCodis web tool (http://genecodis.dacya.ucm.es/)(9, 10) to predict the biological process (GO process) and to perform pathway enrichment analysis (KEGG and Panther pathways) of all miRNAs that were identified as consistently expressed miRNAs in the eligible references.

Results

Selection and overview of the datasets

A total of 194 related literatures were indexed in PubMed, Embase, Web of Science and Ovid database. According to the inclusion criteria and identification of duplicate publication, only 12 publications seemed to meet all of the inclusion criteria and none of the exclusion criteria (Figure 1). The characteristics of these studies are listed in Table 1 in chronological order of the published time (Table 1).

Determination of the consistently reported miRNAs

A total of 134 differentially expressed miRNAs were obtained in the 12 miRNAs expression profiling studies that compared endometriosis tissues with normal tissues, with 28 miRNAs reported in at least two studies, including 4 consistently reported up-

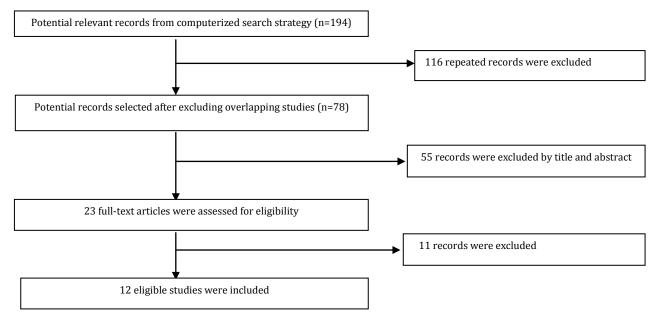


Figure 1. A flow diagram of the literature search and study selection used in this study

First author (reference)	Year	Region	Assay type	Number of samples*	Cut-off criteria	Up- regulated miRNAs	Down- regulated miRNAs
Pan Q (2)	2007	USA	MirVana RNA isolation and enrichment kits	4 pairs+4 N	<i>P</i> <0.05	48	17
Toloubeydokhti T (11)	2008	USA	TaqMan MicroRNA Array	5 pairs+5 N+4 E	5 pairs + 5 N+4 E	1	2
Ohlsson Teague EM (12)	2009	Australia	qRT-PCR	7 pairs	<i>P</i> <0.05 FC>1.5	14	8
Filigheddu N (8)	2010	Italy	TaqMan MicroRNA Array y	13 pairs	<i>P</i> <0.01 FC>2	27	23
Ramon' LA (13)	2011	Spain	TaqMan assay	58 pairs+38 N	<i>P</i> <0.05	3	3
Hawkins SM (14)	2011	USA	TaqMan assay	9 N and 10 E	<i>P</i> <0.01 FC>1.5	10	12
Petracco R (15)	2011	USA	qRT-PCR	50 N+32 E	<i>P</i> <0.05	2	0
Dai L (16)	2012	China	qRT-PCR	12 pairs+12 N	<i>P</i> <0.01	1	0
Liu S (17)	2012	China	TaqMan MicroRNA Array	31 pairs+27 N	<i>P</i> <0.05	0	1
Lin SC (18)	2012	China	qRT-PCR	10 pairs+37 N+17 E	<i>P</i> <0.05	1	0
Shen L (19)	2013	China	qRT-PCR	23 pairs+15 N	<i>P</i> <0.05	0	2
Laudanski P (20)	2013	Poland	TaqMan MicroRNA Array	21E+25N	<i>P</i> <0.05 FC>2	2	13

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Table 1. Twelve microarray-based human endometriosis miRNA expression profiling studies

E= endomeriosis, N=control, pairs= paired eutopic and ectopic endometrial tissues

regulated miRNAs(mir-202, mir-365, mir-1 and mir-150), 9 consistently reported down-regulated miRNAs (mir-23b, mir-200a, mir-200b, mir-200c, mir-

15b, mir-106b, mir-196b, mir-141, mir-375)(Table 2). And 15 inconsistently reported direction miRNA (Table 3).

Table 2. Consistently reported miRNAs (n = 13) in profiling studies (endometriosis tissues versus eutopic tissues or normal tissues)

Direction of expression	miRNA name	Number of studies with same direction	Total number of tissue samples	<i>P</i> -value
ſ	mir-202	2	32	< 0.001
1	mir-365	2	20	< 0.05
1	mir-1	2	20	< 0.05
1	mir-150	2	20	< 0.001
Ļ	mir-23b	3	40	< 0.001
\downarrow	mir-200b	3	39	< 0.05
ţ	mir-200a	3	39	<0.05
\downarrow	mir-200c	2	32	< 0.05
Ļ	mir-141	2	26	< 0.05
\downarrow	mir-375	2	32	≤0.01
Ļ	mir-106b	2	34	< 0.05
\downarrow	mir-196b	2	20	≤0.05
Ļ	mir-15b	2	64	< 0.05

Direction of expression	miRNA name	Number of studies with same direction	Total number of tissue samples	<i>P</i> -value
 ↑	mir-100	3	39	≤0.01
\downarrow	mir-100	1	8	< 0.05
↑	mir-17-5p	1	9	< 0.05
\downarrow	mir-17-5p	3	79	< 0.05
ſ	mir-29c	3	39	< 0.01
\downarrow	mir-29c	1	8	< 0.05
Ť	mir-20a	1	49	< 0.05
\downarrow	mir-20a	3	78	≤0.05
ſ	mir-126	2	20	< 0.05
\downarrow	mir-126	2	39	< 0.001
↑	mir-145	2	20	< 0.05
\downarrow	mir-145	2	29	< 0.05
ſ	mir-125a	2	65	< 0.001
\downarrow	mir-125a	1	8	< 0.05
ſ	mir-99a	2	20	< 0.001
\downarrow	mir-99a	1	8	< 0.05
↑	mir-143	2	20	< 0.001
\downarrow	mir-143	1	8	< 0.05
↑	mir-23a	1	9	0.006
\downarrow	mir-23a	2	31	0.006
↑	mir-222	1	41	< 0.001
\downarrow	mir-222	2	29	< 0.05
↑	mir-199a	1	13	≤0.01
\downarrow	mir-199a	2	20	< 0.05
ſ	mir-125b	1	31	< 0.05
\downarrow	mir-125b	1	8	< 0.05
Ť	mir-21	1	58	< 0.01
\downarrow	mir-21	1	8	< 0.01
ſ	mir-221	1	13	≤0.01
\downarrow	mir-221	1	8	< 0.05

Table 3. Inconsistently reported miRNAs (n=15) in profiling studies (endometriosis tissues versus normal tissues)

Pathway analysis of miRNA targets

By using four target prediction tools (TargetScan, PicTar, miRanda, and miRDB), 9882 candidate genes were predicted for 13 inconsistently reported miRNAs. And all the predicted candidate genes were analyzed by pathways enriched analysis. KEGG and Panther pathways enrichment analysis showed that 9882 endometriosis-related miRNA targets were mainly cancer-related pathways, endocytosis, Wnt signalling pathway, and angiogenesis (Table 4).

Discussion

In this paper, we adopt a meta-analysis to screen endometriosis-related miRNAs from miRNA expression profile data of independent profiling studies, and to obtain conservative target predictions for all miRNAs lists of interest using four up-to-date prediction algorithms (TargetScan, PicTar, PicTar, miRanda and miRDB). And pathway enrichment analysis using different target prediction algorithms were concordant for all of the consistently expressed miRNAs. We found out a meta-signature of four upregulated and nine down-regulated miRNAs. Although the selected method for miRNA expression metaanalysis relates to analysis of the primary expression parameters, such method was often not possible due to the unavailability of the primary data. A great number of miRNAs known in the present and different technical platforms adopted in certain study make the appropriate synthesis of primary data very complex. Moreover, the relatively small sample sizes of microarray data may have resulted in inconformity of

GO processes	Process	FDR	Target number
GO:0006355	Regulation of transcription, DNA-dependent	5.55032e-182	963
GO:0007165	signal transduction	signal transduction 2.72922e-125	
GO:0007275	Multicellular organismal development	7.66 e -100	557
GO:0045944	Positive regulation transcription of from RNA polymerase II promoter	4.43532e-86	378
GO:0007399	Nervous system development	3.20348e-70	280
GO:0007155	Cell adhesion	8.37777e-68	342
GO:0045893	Positive regulation of transcription, DNA dependent	3.78756e -66	301
GO:0007268	Transmission synaptic	6.22556e-57	250
GO:0055085	Transmembrane transport	9.4472e-56	353
GO:0000122	Negative regulation transcription of from RNA polymerase II promoter	1.0109e-55	263
KEGG pathways	Pathway	FDR	Target number
KEGG:5200	Pathways in cancer	3.54381e-53	218
KEGG:4144	Endocytosis	5.04266e-44	144
KEGG:4360	Axon guidance	5.91358e-35	101
KEGG:4810	Regulation of actin cytoskeleton	2.65172e-34	140
KEGG:4010	MAPK signaling pathway	4.03001e-33	162
KEGG:4080	Neuroactive ligand receptor	5.56784e-29	158
KEGG:4310	Wnt signaling pathway	5.87106e-29	105
KEGG:4510	Focal adhesion	8.49293e-29	127
KEGG:4060	Cytokine signaling pathway	4.84554 e-27	151
KEGG:4062	Chemokine signaling pathway	1.23502e-26	119
Panther pathways	Pathway	FDR	Target number
P00057	Wnt signalling pathway	3.24683e-35	173
P00005	Angiogenesis	3.83276e-34	113
P00034	Integrin signalling pathway	4.17619e-29	109
P00047	PDGF signalling pathway	6.46839e-25	90
P00031	Inflammation mediated by chemokine cytokine signaling pathway	3.43609e-23	119
P00026	Heterotrimeric G-protein signaling pathway Gi alpha and Gs, and alpha mediated pathway	3.32868e-23	98
P00012	Cadherin signalling pathway	2. 44252e-19	88
P00018	EGF receptor signaling pathway	2.29847e-18	75
P00021	FGF signaling Pathway	7.78382e-18	72
P00004	Alzheimer diseasepresenilin pathway	4.27409e-17	75

Table 4. Top ten of the significant GO processes, KEGG pathways and Panther pathways enriched with miRNA targets involved in endometriosis

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biological consequences. A meta-analysis method could eliminate these disadvantages and directly compare original data extracted from different technical platforms (21).

All the data from the 12 published studies consisted of more than 317 endometriosis and eutopic or normal tissue samples were analyzed directly, and a series of early researches (2, 11, 12, 14) were analyzed with less than ten specimens, which may have unreliable results. Ohlsson Teague *et al* (22) assessed miRNA expression by microarray analysis in seven paired ectopic and eutopic endometrial tissues and

identified 14 up-regulated and eight down-regulated miRNAs in endometriosis tissues. Pan *et al* (2) identified 48 differently expressed miRNAs in a microarray analysis of endometrium of women with and without endometriosis, and mir-23b down-regulation. Compared with nonendometriosis control endometrium, mir-200b expression level decreased obviously in endometriosis (14).

Angiogenesis is the basis of the occurrence and development in endometriosis. Just astumor, implantation metastasis of endometriosis is based on new neurovascular formation and proliferation, invasion

of extracellular matrix, and lesion formation. Recent studies revealed that miRNA may involve in regulation of angiogenesis, and some miRNAs have the distinction of anti-vasoformation, such mir-15b, mir-16, mir-221, and mir-222. as Interestingly, the results suggest that mir-200 family members (mir-200a, mir-200b, mir-200c, mir-141, which were known to be cancer-related miRNAs down-regulated, while expression of some others (mir-21 (2, 13), mir-199a (2, 13, 16)) were inconsistently reported in the selected studies. Some miRNAs which have been specifically investigated in several studies (such as miR-17-5p, mir-23a, mir-23b (19, 23); mir-20a(18); mir-126(17); mir-135a, mir-135b (15)) were found among both up-regulated and down-regulated. All mirRNA-200 family members (miR-200a, miR-200b, miR-200c, miR-141, miR-429) were found down-regulated, and reached the statistical significance in our analysis, of course, mir-429 was only reported in just one study. Currently, our analysis is limited to comparison of endometriosis and eutopic or normal control tissue. The miR-200 family has been reported to be a fundamental regulator of epithelial-mesenchymal transition, thus enhancing their roles in cancer progression. As a founding member in miR-200 family, miR-200b attracts much focus in carcinogenesis in recent years. Down-regulation of miR-200b has been detected in several malignancies (24-26) and in endometriosis (27). The set of miRNAs with significantly decreased expression levels include all members of the miR-200 family known to be involved in the epithelial to mesenchymal transition process (28).

From the clinical viewpoint, it would be meaningful if the discovery of targets correlated with patient diagnosis, therapy and prognosis. These metasignature miRNAs and gene-to-behavior pathway affected by them are potential candidates as diagnostic and therapeutic agents in endometriosis. Our analysis also concentrated on the challenges connected with the development of miRNA-based tests and emphasizes to the importance of strict inspection of the results before conducting clinical trials. Perez-Iratxeta et al (29) applied a combination of data mining and gene ontology to develop a scoring system for discovering disease-associated genes based on text descriptions of genetically inherited diseases and functional annotations of genes. The scoring showed that the chance of validating potential gene is high for some diseases. Mohammadi et al (30) used microarray data mining and gene ontology to identify disease-causing genes, and predicted marker genes with high accuracy. It indicates that the above method of comparing data is derived from different organisms in studies of disease and human health. Here we showed some familiar and novel pathways for endometriosis, including the pathways in cancer, endocytosis and axon guidance. New visions were also provided into Wnt signaling, angiogenesis, and integrin signaling pathway. These pathways that regulate stem cell transformation indicate the role of miRNAs in endometriosis cell deregulation and development. Therefore, some of these miRNAs may be selected as diagnostic index or acted as therapeutic agent for endometriosis. MiRNAs appear to be potent regulators of gene expression in endometriosis and its associated reproductive disorders, raising the prospect of using miRNAs as biomarkers and therapeutic agent in endometriosis.

It should be emphasized that there were some limitations in our analysis. In this study, the inclusion of researches was based on different assay types. The total number of tissues from available data was relatively small, and tissues included endometriosis tissue, eutopic endometrium, and normal tissue. We extracted data retrieved from different studies, and. published large prospective researches were unattainable for endometriosis. In addition, there were only 12 eligible studies in our meta-analysis, which may also lead to a bias in the results.

Conclusion

By systematic enrichment analysis, we found that these differently expressed miRNAs and gene are potential biomarkers of endometriosis. Our analysis also highlights the challenges connected with the development of miRNA-based tests and emphasizes the need for rigorous evaluation of the results before proceeding to clinical trials.

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Conflict of interest

The authors declare the manuscript has not been published previously, in any language. This research is not involved any pharmacological agents, devices or medical technology, there is not any interest relating with any funding from or pecuniary interests in companies.

References

1. Majid S, Saini S, Dar AA, Hirata H, Shahryari V, Tanaka Y, *et al*. MicroRNA-205 inhibits Src-mediated oncogenic pathways in renal cancer. Cancer Res 2011; 71:2611-2621.

2. Pan Q, Luo X, Toloubeydokhti T, Chegini N. The expression profile of micro-RNA in endometrium and endometriosis and the influence of ovarian steroids on their expression. Mol Hum Rep 2007; 13:797-806.

3. Jiang QY, Wu RJ. Growth mechanisms of endometriotic cells in implanted places: a review. Gynecol Endocrinol 2012; 28:562-567.

4. Vlahos NF, Economopoulos KP, Fotiou S.

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Endometriosis, *in vitro* fertilisation and the risk of gynaecological malignancies, including ovarian and breast cancer. Best Pract Res Clin Obstet Gynaecol 2010; 24:39-50.

5. Suryawanshi S, Vlad AM, Lin HM, Mantia-Smaldone G, Laskey R, Lee M, *et al.* Plasma microRNAs as novel biomarkers for endometriosis and endometriosis-associated ovarian cancer. Clin Cancer Res 2013; 19:1213-1224.

6. Verit FF, Cetin O. Biomarkers of endometriosis. Fertil Steril 2013; 100:e19.

7. O'Donnell KA, Wentzel EA, Zeller KI, Dang CV, Mendell JT. c-Myc-regulated microRNAs modulate E2F1 expression. Nature 2005; 435:839-843.

8. Chang SH, Hla T. Gene regulation by RNA binding proteins and microRNAs in angiogenesis. Trends Mol Med 2011; 17: 650-658.

9. Tabas-Madrid D, Nogales-Cadenas R, Pascual-Montano A. GeneCodis3: a non-redundant and modular enrichment analysis tool for functional genomics. Nucl Acids Res 2012; 40:W478-483.

10. Nogales-Cadenas R, Carmona-Saez P, Vazquez M, Vicente C, Yang X, Tirado F, *et al.* GeneCodis: interpreting gene lists through enrichment analysis and integration of diverse biological information. Nucleic Acids Res 2009;37:W317-322.

11. Toloubeydokhti T, Pan Q, Luo X, Bukulmez O, Chegini N. The expression and ovarian steroid regulation of endometrial micro-RNAs. Reprod Sci 2008; 15:993-1001.

12. Ohlsson Teague EM, Van der Hoek KH, Van der Hoek MB, Perry N, Wagaarachchi P, Robertson SA, *et al.* MicroRNA-regulated pathways associated with endometriosis. Mol Endocrinol 2009; 23:265-275.

13. Ramon LA, Braza-Boils A, Gilabert-Estelles J, Gilabert J, Espana F, Chirivella M, *et al.* MicroRNAs expression in endometriosis and their relation to angiogenic factors. Hum Reprod 2011; 26:1082-1090. 14. Hawkins SM, Creighton CJ, Han DY, Zariff A, Anderson ML, Gunaratne PH, *et al.* Functional microRNA involved in endometriosis. Mol Endocrinol 2011; 25:821-832.

15. Petracco R, Grechukhina O, Popkhadze S, Massasa E, Zhou Y, Taylor HS. MicroRNA 135 regulates HOXA10 expression in endometriosis. J Clin Endocrinol Metab 2011; 96:E1925-1933.

16. Dai L, Gu L, Di W. MiR-199a attenuates endometrial stromal cell invasiveness through suppression of the IKKbeta/NF-kappaB pathway and reduced interleukin-8 expression. Mol Hum Reprod 2012; 18:136-145.

17. Liu S, Gao S, Wang XY, Wang DB. Expression of miR-126 and Crk in endometriosis: miR-126 may affect the progression of endometriosis by regulating Crk expression. Arch Gynecol Obstet 2012; 285:1065-1072.

18. Lin SC, Wang CC, Wu MH, Yang SH, Li YH, Tsai SJ.

Hypoxia-induced microRNA-20a expression increases ERK phosphorylation and angiogenic gene expression in endometriotic stromal cells. J Clin Endocrinol Metab 2012; 97:E1515-1523.

19. Shen L, Yang S, Huang W, Xu W, Wang Q, Song Y, *et al.* MicroRNA23a and microRNA23b deregulation derepresses SF-1 and upregulates estrogen signaling in ovarian endometriosis. J Clin Endocrinol Metab 2013; 98:1575-1582.

20. Laudanski P, Charkiewicz R, Kuzmicki M, Szamatowicz J, Charkiewicz A, Niklinski J. MicroRNAs expression profiling of eutopic proliferative endometrium in women with ovarian endometriosis. Reprod Biol Endocrinol 2013; 11:78.

21. Vosa U, Vooder T, Kolde R, Vilo J, Metspalu A, Annilo T. Meta-analysis of microRNA expression in lung cancer. Int J Cancer J Int du Cancer 2013; 132:2884-2893.

22. Teague EM, Print CG, Hull ML. The role of microRNAs in endometriosis and associated reproductive conditions. Hum Reprod Update 2010; 16:142-165.

23. Jia SZ, Yang Y, Lang J, Sun P, Leng J. Plasma miR-17-5p, miR-20a and miR-22 are down-regulated in women with endometriosis. Hum Reprod 2013; 28:322-330.

24. Pacurari M, Addison JB, Bondalapati N, Wan YW, Luo D, Qian Y, *et al.* The microRNA-200 family targets multiple non-small cell lung cancer prognostic markers in H1299 cells and BEAS-2B cells. Int J Oncol 2013; 43:548-560.

25. Yoshino H, Enokida H, Itesako T, Tatarano S, Kinoshita T, Fuse M, *et al.* Epithelial-mesenchymal transition-related microRNA-200s regulate molecular targets and pathways in renal cell carcinoma. J Hum Gen 2013; 58:508-516.

26. He M, Liu Y, Deng X, Qi S, Sun X, Liu G, *et al*. Down-regulation of miR-200b-3p by bw p73 contributes to the androgen-independence of prostate cancer cells. Prostate 2013; 73:1048-1056.

27. Castilla MA, Diaz-Martin J, Sarrio D, Romero-Perez L, Lopez-Garcia MA, Vieites B, *et al.* MicroRNA-200 family modulation in distinct breast cancer phenotypes. PloS One 2012; 7:e47709.

28. Duns G, van den Berg A, van Dijk MC, van Duivenbode I, Giezen C, Kluiver J, *et al.* The entire miR-200 seed family is strongly deregulated in clear cell renal cell cancer compared to the proximal tubular epithelial cells of the kidney. Genes Chromosomes Cancer 2013; 52:165-173.

29. Perez-Iratxeta C, Bork P, Andrade MA. Association of genes to genetically inherited diseases using data mining. Nat Genet 2002; 31:316-319.

30. Mohammadi A, Saraee MH, Salehi M. Identification of disease-causing genes using microarray data mining and Gene Ontology. BMC Med Genom 2011; 4:12.