

RESEARCH PAPER

In-vitro Anti-cancer and Antioxidant Activity of Biogenically Synthesized Co₃O₄@NiO Core-shell Nanostructures

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ABSTRACT

The anticancer and antioxidant activities of *Datura stramonium* plant leaf extract mediated Co₃O₄ and NiO nanoparticles, as well as Co₃O₄@NiO core-shell nanostructures were investigated. The Co₃O₄@NiO CSNs were synthesized with core-shell concentration ratios of 2:1, 1:1, and 2:3. The Co₃O₄@NiO CSNs and their parent nanoparticles were characterized by using advanced techniques. The band gap energy of Co₃O₄ (11) and NiO (11) NPs were found to be 3.06 eV and 3.45 eV, whereas, the band gap energies of the Co₃O₄ core and NiO shell were 3.68 and 4.52 eV, respectively. Similarly, the average crystalline size of Co₃O₄ (11) and NiO (11) NPs, as well as Co₃O₄@NiO (11) CSNs were found to be 18, 12, and 13 nm, respectively. The spherical shape of Co₃O₄, NiO NPs, and the spherical rod of Co₃O₄@NiO CSNs were investigated from SEM images. The particle sizes of cubic Co₃O₄, NiO NPs, and Co₃O₄@NiO CSNs obtained from TEM images were 15.55, 8.184, and 16.99 nm, respectively. The polycrystalline nature of CN was revealed from SAED analysis. The percentage inhibition of Co₃O₄@NiO CSNs against MCF-7 cancer cells at 100, 200, 400, and 800 µg/mL were found to be 52.41, 68.21, 74.11, and 81.38%, respectively. The IC₅₀ of Co₃O₄@NiO CSNs and epirubicin were found to be 21.25 and 18.01 µg/mL, respectively. The findings show that green synthesized Co₃O₄@NiO CSNs possess potential anticancer activities nearly comparable with standard drugs with less toxicity against normal cells.

Keywords: Breast cancer, Anticancer; Antioxidant; Cytotoxicity

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INTRODUCTION

Cancer and oxidative stress have recently been identified as the primary global health issues. Cancer, a non-communicable illness, is the second leading cause of mortality worldwide, behind cardiovascular disease. A cancer cell invades a healthy part of the body, changing biochemical and cellular functioning and eventually causing death [1, 2, 3, 4]. Breast cancer is mainly developed due to environmental and genetic factors. The report shows that about 27% of breast cancer is caused by genetic factors. Breast cancer is the most often diagnosed cancers that develop in

the lining of the milk duct. Breast cancer became the most frequent class of fatal cancer in women throughout the world a few decades ago [5, 6]. The report showed the preferred prevalence rate for breast cancer increased from 16.7 to 33.6 per 100,000 women between 2000 and 2009. In 2014, the report showed about 232,670 new cases were detected and 40,000 women died in the USA. According to the most recent statistics from global research, the number of breast cancer patients in India will increase from 115,000 to 200,000 by 2030 [7]. Additionally, WHO reported that breast cancer caused 685,000 deaths and 2.26 million new cases in women globally in 2020 [8].

Several treatments have been developed to treat cancer- causing tumors, the most prevalent being surgery, radiation, immunotherapy,

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biological, and chemotherapy [9, 10, 11]. However, these therapeutic approaches have severe side effects on normal cells, with limited effectiveness, toxicity, and non-selective towards cancer cells [12, 13, 14]. In breast cancer treatment, the type of chemotherapy used is typically determined by the tumor's unique receptor status. The primary receptors evaluated while choosing the proper chemotherapy treatment are hormone receptors (estrogen receptor (ER) and progesterone receptor (PR)), human epidermal growth factor receptor 2 (HER2), and triple-negative breast cancer (TNBC). Chemotherapy is frequently used in conjunction with targeted treatments of ER, PR, and HER2-positive breast cancer. However, chemotherapy remains the major systemic treatment option for triple-negative breast tumors, which have no targeted treatments. TNBC is a particularly difficult kind of breast cancer because it lacks the presence of the hormone receptor and HER2. This implies that TNBC does not react to targeted therapy. Chemotherapy intervenes in cell cycle control, suppresses proliferation, and has a cytotoxic impact, which causes intolerable side effects. Furthermore, this medication is ineffective against a wide variety of malignant tumors [15, 16, 17].

As a result, the researchers are interested in designing anticancer drugs that are nontoxic, selective, and effective. Cognizant of the promising effects of nanoparticles such as ZnO [18], Co_3O_4 [19], Fe_3O_4 [20] we explored the underlying mechanism through which iron oxide nanoparticles induce toxicity in human breast cancer cells (MCF-7, CuO [21], NiO [22], Si@NiO [4] researchers are interested in moving their attention from conventional medical techniques to the utilization of nanostructured materials for applying their atom-like behavior in anticancer therapy. The most notable of them are core-shell nanostructures, due to their important uses in drug administration, biomarkers, targeted therapy, and molecular imaging with minimal side effects in breast cancer treatment [23]. For example, ZnO-CuO nanoparticles inhibit breast cancer cell line (MCF-7) growth more than conventional drug doxorubicin at the same concentration [24].

In addition to cancer, the global population faces oxidative stress, which is caused by the overproduction of reactive oxygen species (ROS) in cells. In normal cellular metabolism, ROS such as superoxide radical (O_2^-), hydroxyl radical (OH^\cdot), and hydrogen peroxide (H_2O_2) are formed as

a byproduct that has a significant role in cell signaling and homeostasis [25, 26] Environmental stressors such as high temperatures, drought, salt, chemo-toxicity, radiation, and microbial attack cause an excess of reactive oxygen species to be formed [25] despite extraordinary progress. So, new cancer treatment modalities are needed. Tumor-treating fields (TTFs).

To avoid oxidative damage, endogenous antioxidants such as superoxide dismutase, catalase, and peroxiredoxins scavenge ROS produced during normal cellular metabolism and because of external factors [26]. However, excessive ROS formation creates oxidative stress, which is an imbalance between ROS and antioxidants and causes chronic illnesses such as diabetes, cancer, cardiovascular, and neurological disorders [19, 27]. To address this health problem, researchers have taken the initiative to develop various potent antioxidants to prevent oxidative damage. Exogenous delivery of antioxidants holds promise to alleviate oxidative stress to regain the redox balance. Amongst, there exist reports of the developed metal oxide and their combination with their potent antioxidants. Among the commonly studied, metal oxide nanoparticles such as ZnO [28], CuO [19], Co_3O_4 [29], Fe_3O_4 [30], NiO [31] are reported to be efficient in scavenging reactive oxygen species. Although several studies have been conducted and encouraging findings have been obtained, the problem of treating cancer and oxidative stress has not yet been entirely solved. As a result, the struggle to develop the most effective and strong medications to address the aforementioned human health concerns continues. In recent developments, metal oxide core-shell nanostructures are favored in biomedical applications over monometallic oxide and metal oxide nanostructures due to their superior attributes including low toxicity, high solubility, thermal and chemical stability, minimal defects, biocompatibility, and enhanced permeability to specific target cells as highlighted in recent literature reports [32]. Metal oxide core-shell nanoparticles (MOCSNs) such as NiO@ZnO [33, 34], Co_3O_4 @NiO [35], Fe_3O_4 @Ag [32, 36] Co_3O_4 @ZnO [37], CuO@ZnO [38], Co_3O_4 @ SiO_2 [39], and TiO_2 @ Co_3O_4 [32] have been synthesized by using different methods and for several applications. These metal oxide core-shell nanostructures are synthesized by chemical methods such as sol-gel, hydrothermal,

sono-coprecipitation, solvothermal, emulsion polymerization, microemulsion polymerization, ultrasonic microwave, electrochemical, and electroless deposition [40, 41]. Chemical methods are often expensive and potentially hazardous to the environment and living organisms. Thus, it has become imperative to design green synthesis methods to quest for safe, eco-friendly, cost-effective, quick, easily accessible, highly stable, and biocompatible synthesis [42, 43]. Owing to their wide availability and ease of preparation, plant-based phytochemicals caught the center of focus in green synthesis.

Datura stramonium (*D. stramonium*) is commonly called Jimson weed and it has a wide range of secondary metabolites with strong pharmacological effects. Previous research has reported on the secondary metabolites of *D. stramonium*, including alkaloids, flavonoids, glycosides, terpenoids, tannins, and phenolic compounds [44, 45]. Apart from its therapeutic use, substances like scopolamine and hyoscyamine are recognized for their psychoactivator properties [46]. Additionally, *D. stramonium* plant leaf extract was utilized in the green production of several nanoparticles because of the reducing and stabilizing qualities of its phytochemical ingredients.

Co₃O₄ and NiO NPs are presently recognized as the next generation of anticancer and antioxidant agents due to their bioavailability, dispersibility, selectivity, low host toxicity, permeability, and high effectiveness. Moreover, the synergetic effect that might arise from the combination of these metal oxides also derives interest in designing core-shell type nanostructures. In addition, core-shell nanostructure based therapies are being investigated and designed for a variety of breast cancer subtypes, with the most promising outcomes shown in the treatment of triple-negative breast cancer (TNBC) [15].

In this work, the Co₃O₄@NiO (CN) CSNs with a Co₃O₄ core encapsulated by NiO shell were synthesized using *D. stramonium* leaf extract as a reducing and capping agent that gave distinct qualities and capabilities that are not found in the individual materials. In addition, the anticancer and antioxidant efficacy of *D. stramonium* leaf extract mediated CN CSNs has not yet been reported. Finally, the anticancer effect against MCF-7 (Michigan Cancer Foundation-7) breast cancer cell line and the antioxidant activity of the

green synthesized Co₃O₄@NiO CSNs were studied.

MATERIALS AND METHODS

Chemicals and reagents

In this study, chemicals such as ethanol (99.9%, LabTech Chemicals), cobalt acetate hexahydrate (Co(CH₃CO₂)₂·6H₂O) (Sigma Aldrich), nickel acetate dihydrate (Ni(CH₃CO₂)₂·2H₂O) (Sigma Aldrich), distilled water, 2,2-diphenyl-1-picrylhydrazyl (DPPH) (Sigma Aldrich), ascorbic acid (Sigma Aldrich), Epirubicin (Sigma Aldrich), sodium hydroxide (NaOH) (Sigma Aldrich), and dimethyl sulfoxide (DMSO Sigma-Aldrich) have been used for the synthesis of Co₃O₄, and NiO NPs as well as Co₃O₄@NiO CN CSNs and evaluation of their biological activities. The chemicals and reagents were of analytical grade and used without further purification.

Datura stramonium leaf extract preparation

The collected *D. Stramonium* plant leaf was washed using tap water. After washing, the leaf was rinsed with distilled water to remove any contaminant substances from the surface of the leaf. The cleaned plant leaves were dried under the shed for 15 days and then ground using a grinding machine. During the extraction, the powder of *D. stramonium* leaves was mixed with 500 mL of distilled water in a 1000 mL conical flask. The mixture was boiled at 40 °C using a hotplate and stirred for 90 min. The extract was cooled to room temperature and filtered using Whatman Number 1 filter paper. Then afterward, the filtrate was collected and stored in the refrigerator at 4 °C.

Synthesis of cobalt oxide and nickel oxide nanoparticles

The cobalt oxide nanoparticles (Co₃O₄ NPs) were synthesized in a 1000 mL beaker in 1:2, 1:1, and 2:1 ratios of 0.5 M cobalt acetate hexahydrate and *D. stramonium* leaf extract, respectively, as shown in Fig. 1. For the synthesis of cobalt nanoparticles, the extract of the *D. stramonium* was added dropwise to 0.5 M aqueous solution of cobalt acetate hexahydrate followed by constant stirring for 4 h. Then, 0.1 M of NaOH was added and stirred for an additional 30 min to adjust the pH of the solution and enhance the formation of precipitate. The resulting precipitate was centrifuged at 4,000 rpm for 20 min to obtain Co₃O₄ NP precipitate. The sample was thoroughly washed with ethanol and distilled water to remove

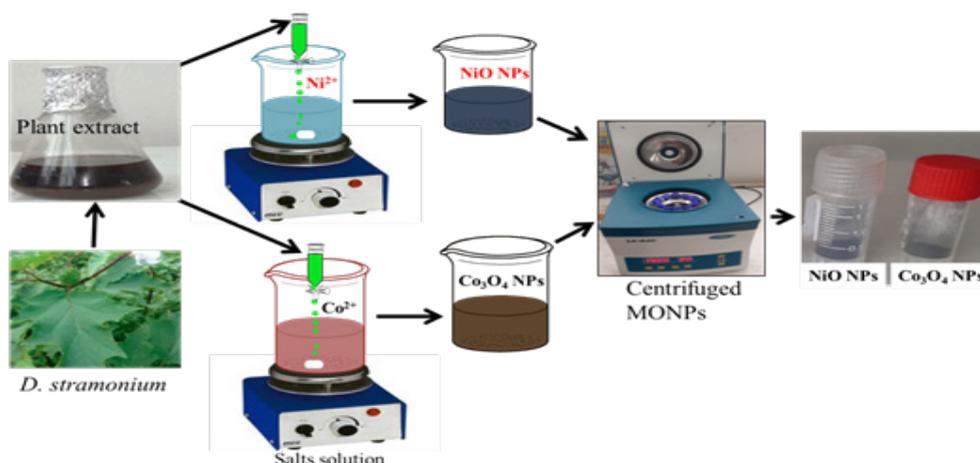


Fig. 1. Schematic procedure for the biogenic synthesis of Co₃O₄ and NiO NPs utilizing *D. stramonium* leaf extract

unwanted substances from the formed precipitate. The synthesized cobalt oxide nanoparticles were collected on the ceramic crucible and dried in an oven at 100 °C overnight and then allowed to cool. The same procedure was repeated for nickel oxide nanoparticles (NiO NPs) synthesis using the extract solution of the same plant leaf. The synthesized nanoparticles were stored safely for further analysis [47]

Synthesis of Co₃O₄@NiO CSNs

Cobalt acetate hexahydrate (Co(CH₃CO₂)₂·6H₂O) and nickel acetate dihydrate (Ni(CH₃CO₂)₂·2H₂O) precursor salts were utilized to synthesize CN CSNs. The CN CSNs were synthesized by holding the concentration of Co(CH₃CO₂)₂·6H₂O constant (0.5 M) while varying the concentration of

Ni(CH₃CO₂)₂·2H₂O as 0.25, 0.5, and 0.75 denoted as CN21, CN11, and CN23, respectively, employing *D. stramonium* leaf extract as a reducing agent as shown in Fig. 2. Specifically in the synthesis of CN11, the *D. stramonium* leaf extract was gradually added to 0.5 M of Co(CH₃CO₂)₂·6H₂O in a 1:1 volume ratio and stirred for 4 h. To adjust the pH of the mixture and enhance the precipitate formation, 0.1 M of NaOH was added and stirred for 30 min. Then to synthesize the CN CSNs, 0.5 M of Ni(CH₃CO₂)₂·2H₂O about 100 mL was added and followed by the gradual addition of an equivalent volume of plant extract. The mixture was stirred for another 4 h. After the completion of stirring for 4 h, 0.1 M of NaOH was added to adjust the pH to 12 and stirred for 30 min. The precipitate was kept in a refrigerator for 12 h to enrich precipitate

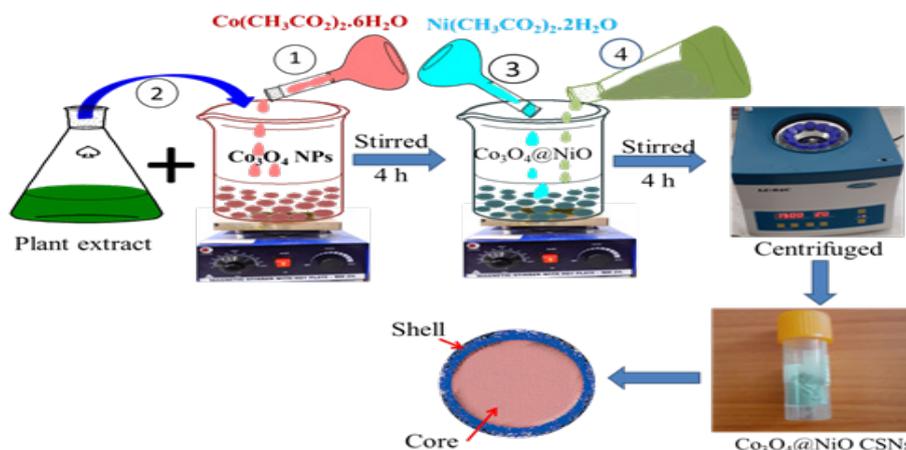


Fig. 2. The schematic route of the biogenic synthesis of CN CSNs

formation. Then, the synthesized $\text{Co}_3\text{O}_4@\text{NiO}$ CSNs were washed three times using distilled water and ethanol. Finally, the precipitate was collected on the ceramic crucible and dried in the oven at 100 °C. The same procedures were repeated for the CN21 and CN23. The dried CN CSNs were calcined at 400 °C and kept for further analysis.

Characterization

Various characterization techniques were used to confirm the synthesis and properties of the metal oxides and the core-shell nanostructures. The thermal study of the Co_3O_4 NPs, NiO NPs, and CN CSNs has been done using the thermal gravimetric-differential thermal analysis (TGA-DAT (DTG-60H, SHI-MADZU Corporation, Japan)) in air flow heating rate of 10 /min starting at 30-800. To determine several functional groups, the green synthesized nanostructures were homogenized with a KBr pellet and analyzed using Fourier transform infrared spectroscopy (FT-IR) Bruker TENSOR27 spectrometer with a scanning range of 4000 - 400 cm^{-1} . The crystal structure and average crystallite size of Co_3O_4 NPs, NiO NPs, CN21, CN11, and CN23 nanoparticles were analyzed by using X-ray Diffraction (XRD-7000, SHIMADZU Corporation, Japan) furnished with a Cu metal used for generating a Cu $\text{K}\alpha$ radiation with $\lambda = 0.15406$ nm and recorded in the range from 10-80. The scanning electron microscopy SEM, model CamScan MV2300) was used to investigate the morphological structure of nanoparticles. Furthermore, the nanostructural properties of the nanoparticles have been elucidated by using a transmission electron microscope (TEM), higher transmission electron microscope (HRTEM), and selective area electron diffraction (SAED) model Philips CM30) functioned in 200 kV.

Anticancer activities

Breast cancer (MCF-7) and peripheral blood mononuclear (PBM) cells were acquired from India's National Center for Cell Sciences (NCCS). The breast cancer cells were cultivated using the standard technique of Dulbecco's Modified Eagle's Medium supplemented with fetal bovine serum. MCF-7 and PBM cells were bathed and grown in a humidified incubator at 37 °C in 5% CO_2 and injected with 10% (v/v) fetal bovine serum (FBS), penicillin (100 IU/mL), and streptomycin (100 g/mL) until they reached confluence. The cell was separated using a cell-dissociating solution

(0.2% trypsin, 0.02% EDTA, and 0.05% glucose in PBS). After verifying the cells' vitality, they were centrifuged. In a 96-well plate, 5×10^4 cells/well were cultured and incubated for 24 h at 37 °C with 5% CO_2 [3].

The cytotoxicity of CN CSNs and Epirubicin on PBM and MCF-7 cells was determined *in vitro* using the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay [5]. The standard cells were trypsinized and adjusted to 5×10^4 cells/ml for cell counting in medium with 10% FBS (v/v). Each of the 96 wells of the μL plate received 100 μL of diluted cell suspension (5×10^4 cells/well). Following incubation, the test solutions in the wells were discarded, and 0.05 mg MTT was applied to each well. The plates were held for 4 h at 37 °C under a 5% CO_2 environment. The consequential formazan was dissolved in 100 μL of DMSO with moderate shaking at 37 °C. The absorbance was measured at 570 nm with a microplate reader. Equation (1) was used to calculate % growth inhibition, and dose-response curves for each cell line were utilized to get IC_{50} values [10]. Cell growth inhibitions were measured in triplicate and reported as mean \pm standard deviation.

$$\% \text{Inhibition} = \frac{\text{OD of control} - \text{OD of sample}}{\text{OD of control}} * 100$$

(1)

The OD of control represents the optical density of the untreated control, whereas the OD of the sample represents the optical density of the treated sample, which is treated. The percentage inhibition was quantified by determining the difference between optical density between the control and the sample.

Antioxidant activities

The 2,2-diphenyl-1-picrylhydrazyl (DPPH) free radical was used to test the radical scavenging capacity of biogenically produced Co_3O_4 and NiO NPs as well as CN CSNs. The activity was assessed using the modified approach described in the earlier work report [27]. In DMSO, 3 mL of 100 μM DPPH was added to 1000 μL of methanolic Co_3O_4 and NiO NPs, as well as CN CSNs at concentrations of 50, 100, 200, 300, and 500 $\mu\text{g}/\text{mL}$. After sonication, the mixture was maintained in a dark chamber for 30 min before being incubated at 37 ± 2 °C for the same amount of time. The UV-vis absorbance (UV-Vis) of the same concentration a

positive control (ascorbic acid), Co₃O₄, NiO NPs as well as CN CSNs mixed with 0.1 mM DPPH was measured at λ_{max} of 517 nm. All the studies were carried out in triplicate, and the average absorbance for each sample was taken into consideration. Finally, the % scavenging capacity of nanoparticles was calculated using equation (2) [48].

$$\% \text{Radical scavenging activity} = \frac{\text{Absorbance of control} - \text{Absorbance of sample}}{\text{Absorbance of control}} \times 100 \quad (2)$$

Statistical data analysis

The experimental data were evaluated by using a statistical method known as the analysis of variance (ANOVA) function of the statistical package for social science (SPSS) version 20 and provided as a for triplicate experiments. Advanced software such as ImageJ (153-win imagej.exe), GatanMicroscopy Suite (GMS 64 bit), Origin software (Originpro 64 bit), and Microsoft Excel, 2013.

RESULTS AND DISCUSSIONS

As reported in our previous work, the phytochemical constituents of *D. stramonium* plant leaf, such as alkaloids, flavonoids, tannins, saponins, phenols, phytosterols, glycosides, terpenoids, and anthraquinones, were qualitatively screened. The phytochemicals containing hydroxyl and carboxy group were thought to be employed to decrease the size of the metal ion and cap the

synthesized nanoparticles [49].

The CN CSNs were synthesized in two steps utilizing the *D. Stramonium* leaf extract from Co(CH₃CO₂)₂·6H₂O which was used as a core and Ni(CH₃CO₂)₂·2H₂O precursor salts as a shell in the nanostructures. The precursor salt of the core was mixed with the shell in (2:1), (1:1), and (2:3) concentration ratios. The synthesized Co₃O₄ core was coated with a NiO shell. The Ni²⁺ ions were electrostatically bound to the functional group of phytochemicals found on the surface of the core and possess reduction to some extent. The complete reduction of the shell was carried out after further addition of plant extract. The synthesized CSNs were capped and stabilized by these bioactive molecules as shown in Fig. 3. In the green synthesis of MONPs, the mechanistic electron interaction occurs in different stages. Under the controlled temperature and pH, the phytochemicals from the extract donate electrons to reduce metal ions to metal nanoparticles. Functional groups found in phytochemicals, including hydroxyl (-OH), carboxyl (-COOH), amino (-NH₂), and others, can adsorb onto the surface of nanoparticles, giving stability and influencing the size and form of the particles. The metal ions are saturated to form hydroxyl complexes and grow up to form crystallite with oxygen after supersaturation as depicted in Fig. 3. The phytochemicals of *D. Stramonium* were used as a capping agent to stabilize the metal nanoparticles by preventing agglomeration [50, 51].

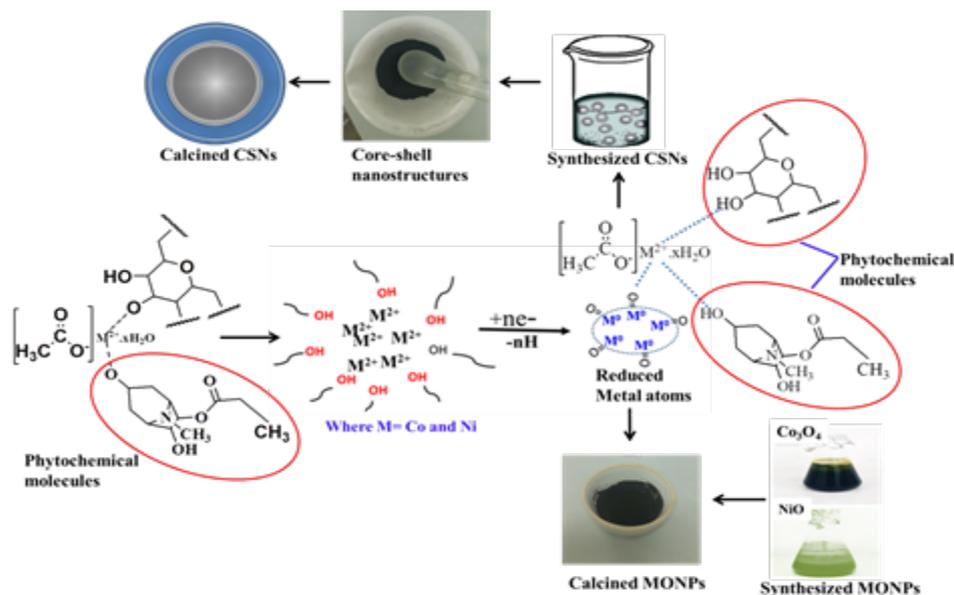


Fig. 3. Mechanism of green synthesis of metal oxide nanoparticles and core-shell nanostructures

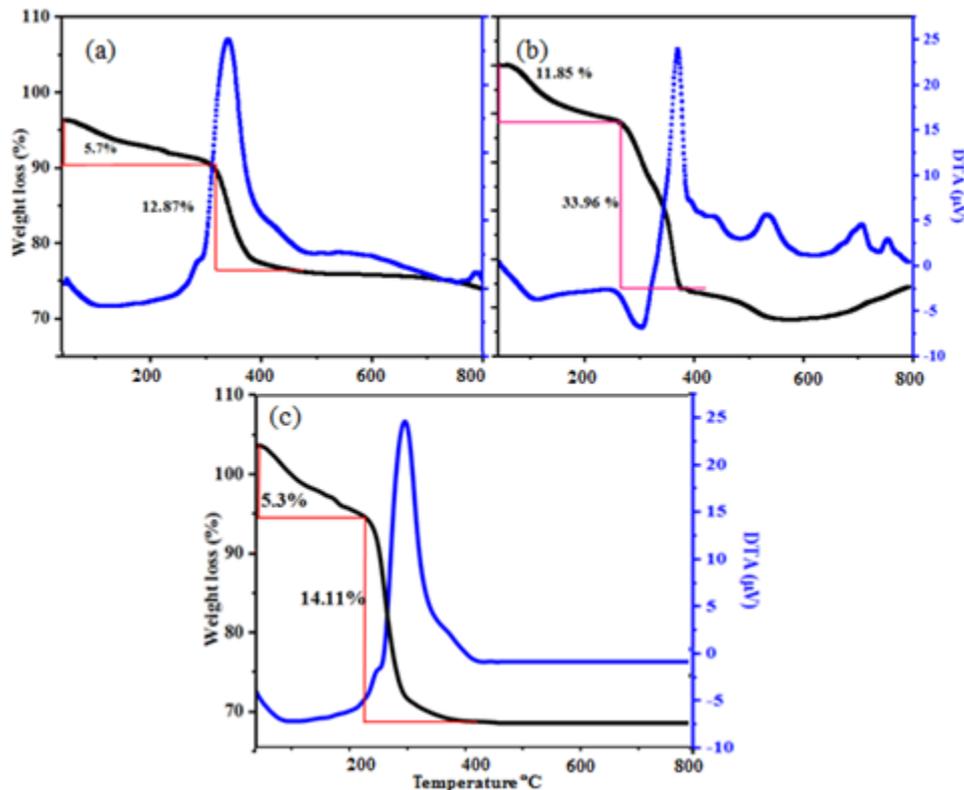


Fig. 4. The graph of TGA/DTA curve (a) Co₃O₄ NPs, (b) NiO NPs, and (c) Co₃O₄@NiO core-shell nanostructures.

Thermal analysis

The thermal stability of biogenically synthesized Co₃O₄, NiO, and CN CSNs was studied by using the thermogravimetric/differential thermal (TGA/DTA) analysis technique in an atmosphere through the speed of 10 at a temperature range of 30-800 as shown in Fig. 4. In TGA analysis, the curve for Co₃O₄, NiO, and CN CSNs showed weight loss at two stages. The first weight loss of about 5.7%, 11.85%, and 5.3% in the range of 30-260, 30-260, and 30-220 were found to be water for Co₃O₄, NiO, and CN CSNs, respectively. The loss observed in the second stage was due to the decomposition of some organic molecules from the plant extract utilized during the synthesis of nanoparticles. The percent loss for these organic molecules was found to be 12.87%, 33.96%, and 14.14%, for Co₃O₄, NiO, and CN CSNs within the temperature ranges of 270-350, 270-380, and 230-400, respectively.

Additionally, the exothermic and endothermic energy were also investigated by using differential thermal characterization techniques. The loss of weight and change in energy of synthesized samples indicated that the bioactive molecules involved in the synthesizing of nanoparticles

were removed before calcination temperature. As shown in Fig. 4, the biologically synthesized Co₃O₄ NPs, NiO NPs, and CN CSNs became thermally stable at above 350, 380, and 400, respectively. These thermal points indicated that the calcination temperature of the synthesized sample was 350, 380, and 400 for Co₃O₄ NPs, NiO NPs, and CN CSNs, respectively.

UV-Vis analysis

The green synthesized Co₃O₄NPs, NiO NPs, and CN CSNs were analyzed by using UV-visible spectroscopy in the range of 200-800 nm wavelengths. As shown in Fig. 5, the absorption band of metal oxide nanoparticles synthesized in (1:1), (1:2), and (2:1) volume ratios and core-shell nanostructures denoted as CN21, CN11, and CN23 concentration ratios were recorded. The green synthesized nanoparticles absorbed light, which was recorded and utilized to calculate band gap energy. According to the generated data, the absorbance values for these nanoparticles differed mostly related to their size. Metal oxide nanoparticles absorb light of a given wavelength, causing electrons to move between the highest occupied and lowest empty orbitals.

As shown in Fig. 5(a), the absorption spectra of Co_3O_4 (11), Co_3O_4 (12), and Co_3O_4 (21) NPs developed bands at two distinct wavelengths. The absorption bands found at 230 and 276 nm were associated with Co_3O_4 (11) NPs, whereas 288 and 338 nm corresponded to Co_3O_4 (12), and 368 and 417 nm to Co_3O_4 (21) NPs, respectively. The absorption bands observed at shorter and longer wavelengths in each spectrum are caused by the charge transfer from $\text{O}^{2-} \rightarrow \text{Co}^{2+}$ and $\text{O}^{2-} \rightarrow \text{Co}^{3+}$, respectively [52]. Furthermore, the absorption spectra of NiO (11), NiO (12), and NiO (21) NPs were detected at 264, 296, and 310 nm, as shown in Fig. 5(b). All the absorption bands that occurred at 264, 296, and 310 nm were due to the charge transfer from O^{2-} to Ni^{2+} [53] magnetic hyperthermia constitutes a complementary way to cancer treatment. This article reports a promising aspect of magnetic hyperthermia addressing superparamagnetic and highly Fe/Au core-shell nanoparticles. Those nanoparticles were prepared using a wet chemical approach at room temperature. We found that the as-synthesized core shells assembled with spherical morphology, including face-centered-cubic Fe cores coated and Au shells. The high-resolution

transmission microscope images (HRTEM). Among the Co_3O_4 and NiO NPs synthesized, the 2:1 ratios show λ_{max} shift to the longer wavelength (red-shift). In contrast to the others, the energy level of Co_3O_4 (21) and NiO (21) NPs associated with their electronic states were changed and resulted in a plasmonic resonance shift. Similarly, the CN CSNs reflected the UV-visible light at two different wavelengths. As depicted in Fig. 5(c), the Co_3O_4 -core and NiO-shell absorb the UV light at (254 nm, 322 nm), (240 nm, 306 nm), and (26 nm, 330 nm) pair of wavelengths that corresponds to electron transition between $\text{O}^{2-} \rightarrow \text{Ni}^{2+}$ within CN21, CN11, and CN23, respectively.

The absorbance data was utilized to derive the optical band gap energy of MONPs. The band gap energy was calculated using the Tauc relation, as stated in equation (3) [54] green synthesis of nickel oxide nanoparticles using phytochemicals from three different sources was employed to synthesize nickel oxide nanoparticles (NiOx NPs).

$$F(r)hv = A(hv - E_g)^2 \quad (3)$$

Where $F(r)$ is the Kubelka-Munk function, $h\nu$ is photon energy, and E_g is band gap energy. The

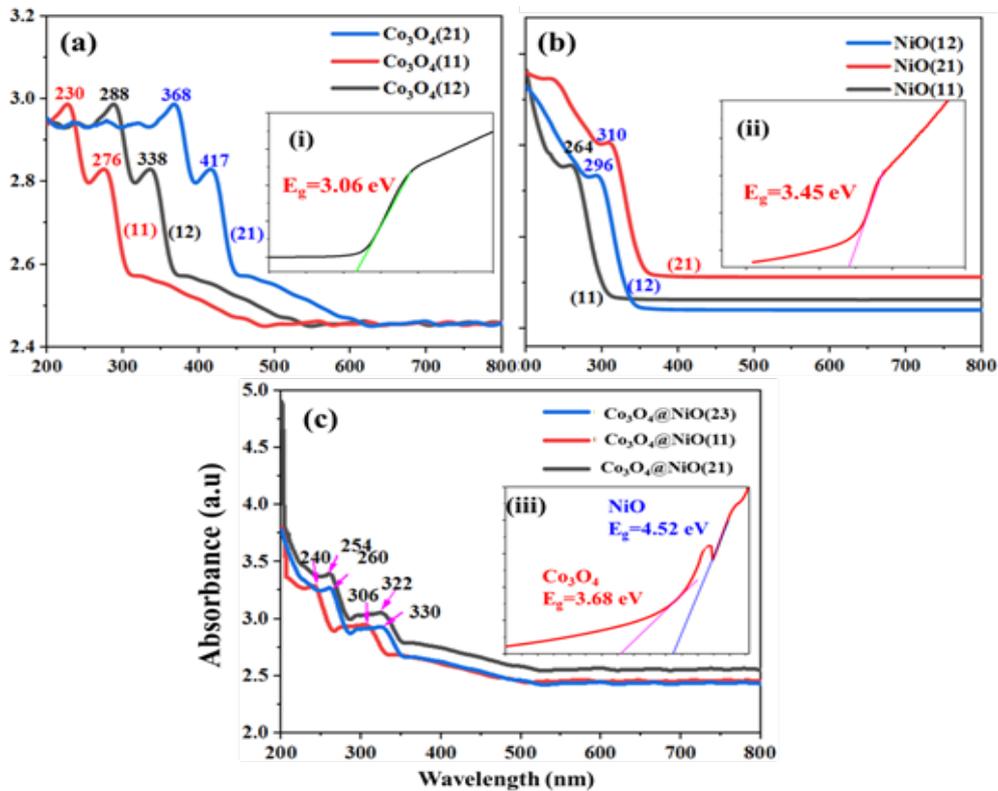


Fig. 5. UV-visible optical absorption spectra with their corresponding band gap energy of (a-i) Co_3O_4 NPs, (b-ii) NiO, and (c-iii) CN CSNs.

band gap energy values of all Co_3O_4 and NiO NPs and CN CSNs were calculated using extrapolation techniques. As shown in Figure 5(a-i and b-ii), the band gap of Co_3O_4 and NiO NPs was found to be 3.06 and 3.45 eV, respectively. In contrast to their corresponding bulk, the band gap value indicates a blue shift [55, 56] diffuse reflectance spectroscopy (DRS). The blue shift is because of the large band gap energy between the valance and conduction band in the nanoparticles. In general, the band gap energy of Co_3O_4 and NiO NPs was larger than their corresponding parent bulk materials [57].

The optical band gap energy of the green synthesized CN CSNs was also studied as shown in Fig. 5(c-iii). The Co_3O_4 core and NiO shell exhibit their characteristic band gap energy. According to Fig. 5(c-iii), the computed band gap energy values of 3.68 and 4.52 eV were believed to be associated with the Co_3O_4 core and NiO shell. The large band gap energy of NiO shell nanoparticles is in good agreement with UV-Vis data which has an inverse relation with absorption.

X-ray diffraction (XRD) analysis

The average crystallite size, the nature of crystal structure, and composition analysis of

biogenically synthesized Co_3O_4 , NiO NPs, and CN CSNs were examined using an XRD pattern. The diffraction pattern of Co_3O_4 NPs synthesized within three volume ratios such that 1:2, 1:1, and 2:1 were obtained from XRD analysis. The analysis was carried out using the X-ray photon (CuK α radiation) which is the energy released during the electron of k-shell return to its original state from the excited state at a characteristic wavelength of 0.154 nm and with the scanning rate of 0.16 s⁻¹ within the range of 2 θ from 10-80°. As depicted in Fig. 6(a), the diffraction peaks of Co_3O_4 NPs were observed for Bragg reflection with 2 θ values of 19.02°, 31.22°, 36.82°, 38.48°, 44.8°, 55.64°, 59.32°, 65.22°, and 77.26° corresponding to the set of lattice planes with Miller indices (hkl) values of (111), (220), (311), (222), (400), (422), (511), (440), and (622), respectively. Similarly, the diffraction peaks of green synthesized NiO NPs shown in Fig. 6(b) were obtained at 2 θ values of 37.42°, 43.42°, 63.06°, 75.56, and 79.58° matching with the miller indices of (111), (200), (220), (311), and (222) planes, respectively. The obtained result was complemented with the previous work and properly matched the standard database JCPDS card no. 00-042-1467 of Co_3O_4 NPs and 01-073-

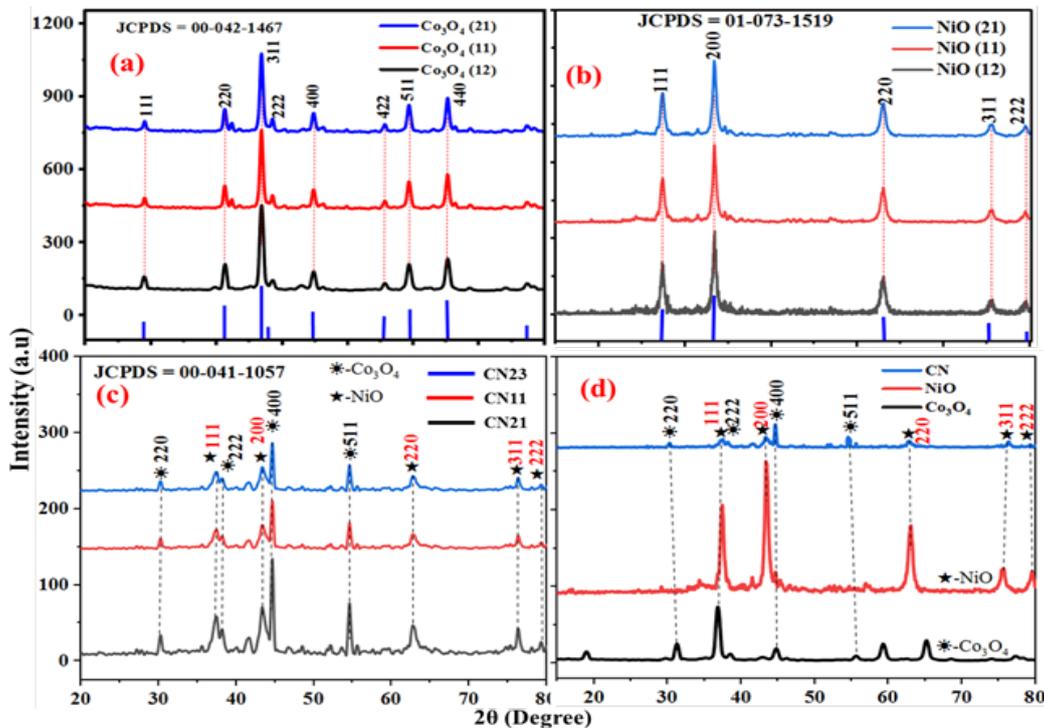


Fig. 6. The XRD pattern of (a) Co_3O_4 NPs, (b) NiO NPs, (c) CN CSNs, and (d) CN CSNs with parent nanoparticles

1519 of NiO NPs. The formation of core-shell nanostructure comprised of Co₃O₄ core and NiO shell NPs was also confirmed by using XRD analysis. As depicted in Fig. 6(c), the diffraction peak from the planes having the miller indices of (220), (111), (222), (200), (511), (220), (311), and (222) at 2θ values of 30.28, 37.52, 38.2, 43.32, 54.54, 62.72, 76.38, and 79.34°, respectively were obtained. The combination of plane lattice and diffraction patterns of CN CSNs indicated the formation of core and shell which agreed with the previous work [52]. Furthermore, the average crystalline sizes of biogenically synthesized Co₃O₄, NiO NPs, and CN CSNs were calculated using Debye-Scherrer's formula presented in equation (4) [58] green nanotechnology has gained attraction in the synthesis of metallic nanoparticles due to their cost-effectiveness, simple preparation steps, and environmentally-friendly. In the present study, copper oxide nanoparticles (CuO NPs

$$D = \frac{k\lambda}{\beta \cos\theta}$$

(4)

Where, D stands for average crystallite size (nm), k is the Scherrer constant ($= 0.89$ eV), is the wavelength of the X-ray photon ($\text{CuK}\alpha = 0.15406$ nm), and β is the full-width at half-maximum (FWHM) of the diffraction peak appeared due to Bragg's reflection in radian, and θ is half of the angle between transmission and diffraction. The calculated average crystalline size of Co₃O₄ NPs synthesized within volume ratios of (1:2), (1:1), and (2:1) were found to be 23, 18, and 21 nm, whereas, NiO NPs were 17, 12, and 15 nm, respectively.

XRD investigation revealed the cubic structure of Co₃O₄ NPs (space group Fd-3m). The length of cubic lattice parameters with $a = b = c$ was determined. The estimated lattice parameter value was $a = b = c = 0.8074012$ nm, matching the theoretical value of 0.8072904 nm length with angles of $\alpha = \beta = \gamma = 90^\circ$. It was also found that NiO has a face-centered cubic structure with the space group 225: Fm-3m. In the crystal structure of NiO NPs, the lattice parameter was computed using equation (4) and found to be $a=b=c=0.42024$ nm with lattice angle ($\alpha = \beta = \gamma = 90^\circ$). The obtained lattice parameter values agreed with the theoretical 0.41268 nm.

Similarly, the average crystallite size of CN21, CN11, and CN23 were found to be 14, 13, and

16 nm, respectively. Reasonably, the calculated average crystallite size of CN11 is smaller than the two ratios due to the optimum concentration and volume ratios of precursor salt to plant extract. In the case of CN23, the amount of bioactive molecules present in the extract was not enough to reduce the size and cap the nanoparticles which led to the agglomeration of the particles and resulted in large particle size. Whereas in CN21 the excess amount of phytochemicals present in the leaf extract is believed to reduce very small size which can undergo nucleation and agglomeration. The XRD pattern of green synthesized CN CSNs was in good agreement with the previous report [57] The peak position of CSNs appeared at the same 2θ of parent materials that conformed well with standard pattern data and previous work. Green-synthesized Co₃O₄@NiO core-shell nanostructures had lower grain sizes than the chemically synthesized nanostructures, which might be advantageous in terms of performance and applications [59]

As shown in Fig. 6(c), the plane of the core material diffracted the X-ray radiation from the planes (220), (222), (400), and (511) at 2θ values of 31.22, 36.82, 38.48, and 55.64°, and the shell with the miller indices (111), (200), (220), (311), and (222) at 2θ of 37.42°, 43.42°, 63.06°, 75.56, and 79.58°, respectively. The Diffraction peaks of the shell material are more in number than that of the core materials relative to the individual nanoparticles. This confirmed that the NiO NPs were formed on the surface of Co₃O₄ NPs as a shell as reported elsewhere in previous reports. As shown in Fig. 6(c), all the diffraction peaks of the biosynthesized CN CSNs within all concentration ratios have fitted the peak of corresponding parent particles [60]. In addition to this, the formation of all CN CSNs correctly matched the standard database JCPDS card no. 00-041-1057.

FT-IR spectral analysis

The functional groups of *D.stramonium* plant leaf extract involved in the synthesis of MONPs and CSNs were reported in our previous work [37]. The functional groups contained in the biogenically produced Co₃O₄, NiO NPs, and Co₃O₄@NiO CSNs were examined using FTIR spectroscopy. The functional group analysis was done between 4000-300 cm⁻¹. The absorption spectra resulting from various vibrational and stretching chemical bonds were used to predict the nature of the

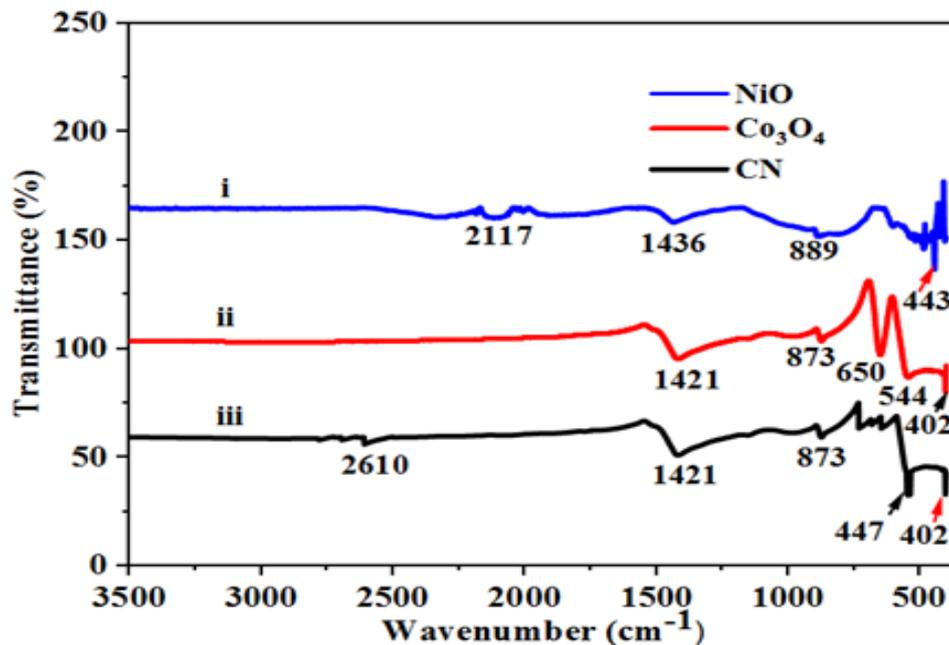


Fig. 7. FTIR spectra of *D. stramonium* leaf extract mediated (i) NiO NPs, (ii) Co₃O₄ NPs, and (iii) CN CSNs.

synthesized nanoparticles. A variety of notable functional groups displayed absorption peaks at distinct wave numbers (cm^{-1}) in green-synthesized Co₃O₄ NPs.

The Co₃O₄ NPs exhibited two sequential stretching vibrations. Co₃O₄ NPs have two mixed valences: Co²⁺ and Co³⁺ ions, which produce O-Co²⁺ and O-Co³⁺, respectively. The tetrahedrally coordinated Co²⁺ and octahedrally coordinated Co³⁺ ions stretched at 544 and 402 cm^{-1} , respectively. Because of nanoparticles' enormous surface area, CO₂ molecules may be absorbed and form intermolecular interactions; causing the two more spectra formation within Co₃O₄ NPs. The adsorbed CO₂ molecules undergo two characteristic bending vibration modes. As indicated in Fig. 7(ii) of Co₃O₄ NPs spectra, CO₂ molecules exhibit in-plane bending vibration at 1421 cm^{-1} and out-of-plane bending vibration at 873 cm^{-1} [56] diffuse reflectance spectroscopy (DRS).

At the higher absorption wave numbers, the absence of peaks indicates that the green synthesized sample was calcined, causing the bioactive molecules to decompose. Similarly, the typical absorption peaks of NiO NPs of C-N stretching, asymmetric carbonate (CO₃²⁻), and symmetric carbonate (CO₃²⁻), were found to be 2117, 1436, and 889 cm^{-1} , respectively. The metal-

oxygen stretching vibration in Ni-O undergoes absorption of infra-red radiation at 443 cm^{-1} as shown in Fig. 7(i). In addition, the absorption peak of Co₃O₄@NiO CSNPs was studied. Fig. 7(iii) shows that the absorption bands for Co₃O₄@NiO CSNPs were at 2610, 1421, 873, 447, and 402 cm^{-1} due to the C=O stretching of carboxylic acid, C-H bending, C=O of symmetric carbonate (CO₃²⁻), Ni-O stretching vibrational, and Co-O vibration, respectively.

SEM-EDAX analysis

The surface topography of the green synthesized Co₃O₄, NiO NPs, and CN CSNs was revealed by using a surface morphology analyzer known as the scanning electron microscope technique. The surface of the Co₃O₄ NPs sample was scanned by an electron beam from a SEM instrument's electron gun, resulting in a nearly spherical and flattened shape, as shown in Fig. 8(a). The flattened surface of Co₃O₄ NPs is believed to be due to the agglomeration of the particles which leads to the enlargement of particles in size. ImageJ software was used to evaluate the particle size distribution of biologically synthesized Co₃O₄ NPs, revealing an average particle size of 63.4 nm in the 20-140 nm range as depicted in Fig. 8(d).

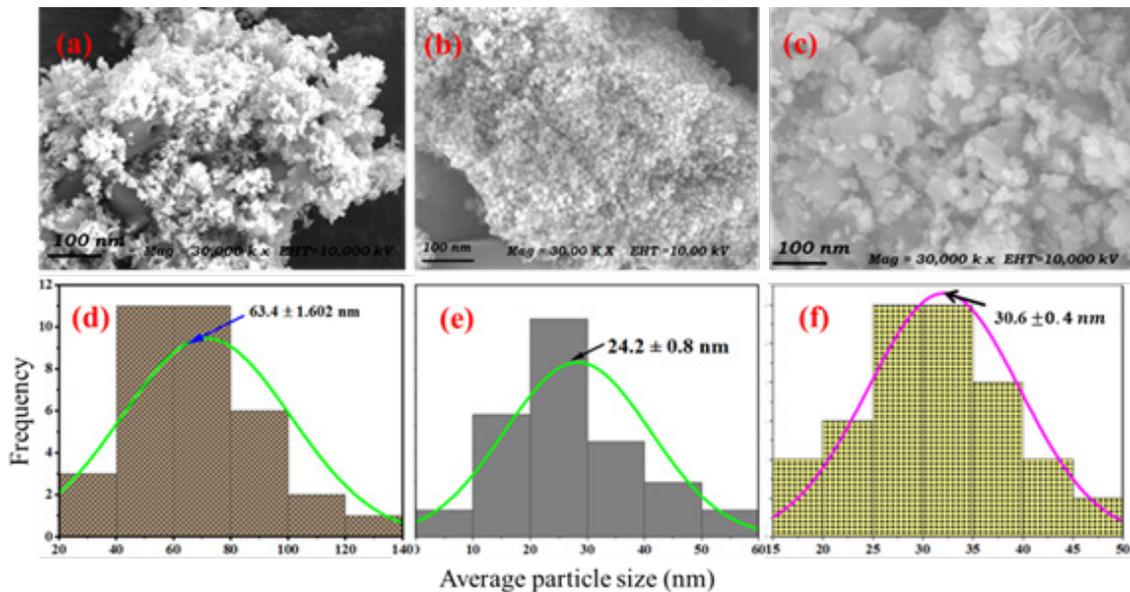


Fig. 8. The SEM images-particle size distribution (a/d) Co₃O₄, (b/e) NiO NPs, and (c/f) CN CSNs

Similarly, the morphological feature of NiO NPs was studied. Fig. 8(b) shows that the NiO NPs had a typical spherical form with a rough surface. Fig. 8(e) shows that NiO NPs have particle sizes ranging from 5 to 60 nm, with an average particle size of 24.2 nm. Similarly, SEM scans revealed the spherical and rod form of biologically synthesized CN CSNs, as illustrated in Fig. 8(c) [61] large specific surface area, simple preparation process, and convenient functionalization. In this study, a core-shell mesoporous material MCM-41@SiO₂ was synthesized and functionalized by CoCl₂, and subsequently KSCN, to prepare an ammonia adsorbent. The adsorbent was proved to possess high surface area, good sphericity, uniform size, good dispersibility, and high adsorption capability following DLS, SEM, TEM, and a static adsorption study. Moreover, the successful functionalization and thermal stability were confirmed by FT-IR, XPS, and TGA. The material was then used to fabricate a glass tube sensor for the rapid naked-eye detection of ammonia gas. The sensor showed good performance in terms of sensing speed, selectivity, accuracy, and reusability. Within 5 s, NH₃ could be detected by the discoloration, and a wide NH₃ concentration from 20 to 1000 ppm could be detected. It also showed a good

linear relationship between discoloration length and ammonia concentration. A reversible color change from blue to yellow indicated the presence of NH₃, which was attributed to the formation and disassembly of ammonia-containing complex [Co(NH₃). The average particle size of CN CSNs has been found that 30.6 nm. In contrast to the average crystalline size obtained from XRD analysis, the particle size of Co₃O₄, NiO NPs, and CN CSNs calculated for SEM images was greater in small nanoscale. The larger average size of particles calculated for the SEM image indicated that particles were agglomerated to some extent.

The elemental composition of Co₃O₄ and NiO NPs was also investigated using energy dispersion X-ray spectroscopy (EDAX). As shown in Fig. 9(b), the 100% elemental composition of Co₃O₄ NPs confirms the existence of solely Co and O, ensuring that the synthesized nanoparticles are free from impurities. The elemental percentages of Co and O in Co₃O₄ NPs were determined to be 35 and 65%, respectively. The high-intensity peak suggests that green synthesized Co₃O₄ NPs have a higher percentage of oxygen. The elemental percentage of Ni and O in NiO NPs was found to be 66.3 and 33.7 %, respectively.

The elemental composition of Co₃O₄@NiO

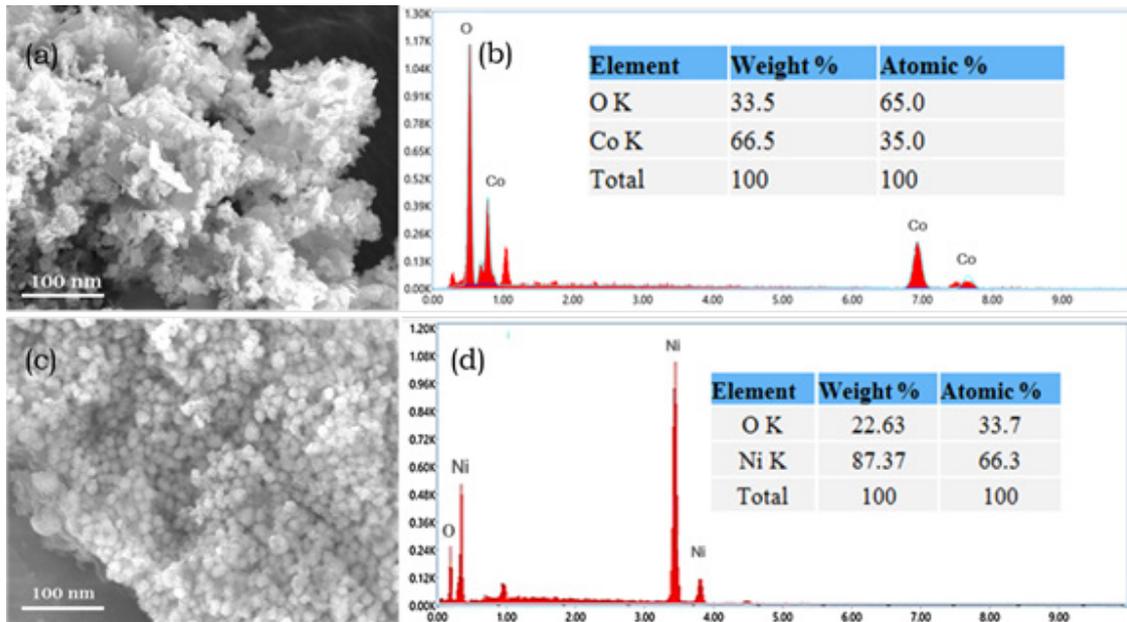


Fig. 9. The SEM image-EDAX spectra of (a/b) Co₃O₄ NPs and (c/d) NiO NPs

CN CSNs was also identified and quantified using energy-dispersive X-ray (EDAX) techniques as depicted in Fig. 10. The distinctive peaks seen on the EDX graph demonstrate the successful synthesis of CN CSNs with desired elemental composition and no impurities. As shown in the graph of Fig. 10(e), all peaks represented the element belonging to the CN CSNs. The map sum spectrum of the sample also ensured that the

green synthesized CN CSNs contained only O, Co, and Ni elements with atomic percentages of 60.48, 15.03, and 24.49%, respectively. Additionally, as depicted in Fig. 10(b) obtained from EDAX, the Co element with blue color is surrounded by the Ni element with green color which implies that Co₃O₄ NPs are found in the core and NiO NPs are on the shell which confirms the formation of Co₃O₄@NiO core-shell nanostructure and agreed with previous

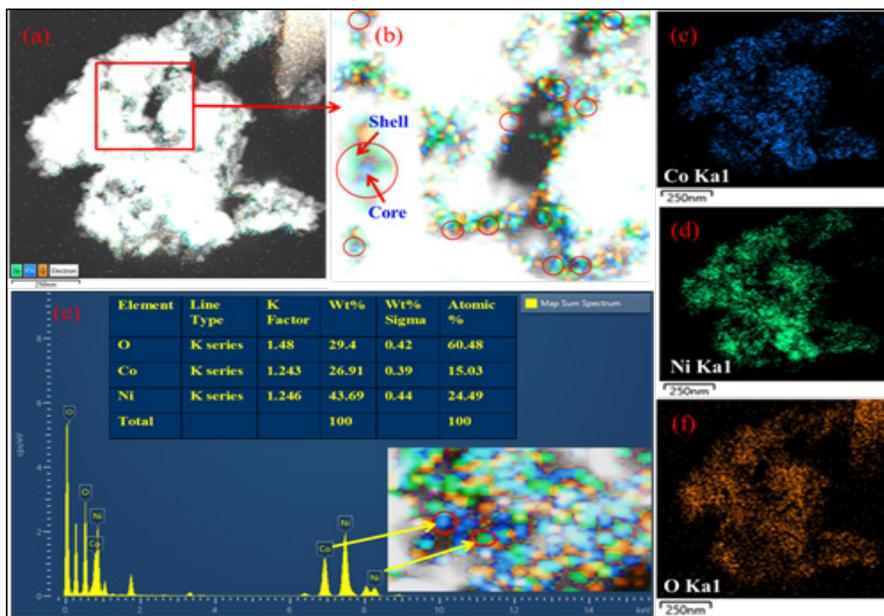


Fig. 10. The elemental composition of Co₃O₄@NiO core-shell nanostructure generated from EDX.

work [62].

TEM/HR-TEM and SAED analysis

The biologically synthesized Co_3O_4 and NiO NPs, as well as the CN CSNs, were further analyzed using TEM/HR-TEM and SAED methods. The TEM and HR-TEM images revealed precise information about the size, shape, and crystallinity of the MONPs and CSNs, while the SAED patterns confirmed the crystalline nature of the synthesized material. Fig. 11(a) shows that the *D. stramonium* plant leaf extract mediated Co_3O_4 NPs have cubic and rod shapes. Fig. 11(b) reveals the spherical and oval shapes of NiO NPs and the particle shapes are not evenly distributed, which is due in part to particle aggregation.

In addition to the mean particle size obtained from SEM Images and average crystalline size from XRD data, the mean particle size of Co_3O_4 and NiO NPs were also calculated from TEM images as shown in Fig. 11(c/d).

In the case of Co_3O_4 NPs, the particle size distribution was found to be between the 10-25 nm range, with a mean particle size of 15.55 nm as determined by the diameter of the particles

measured using ImageJ software, as shown in Fig. 11(c).

Similarly, the average particle size of NiO NPs synthesized from plant leaf extract was evaluated. Fig. 11(d) shows that the particle sizes of NiO NPs were generally between 4-14 nm. The bioactive compounds stabilized the NiO NPs by establishing a protective layer and preventing aggregation, resulting in tiny size. Fig. 11(d) shows that the mean particle size of NiO NPs is 8.184 nm, with an uncertainty of ± 1.64 nm. ImageJ software yielded an uncertainty of ± 1.64 nm, indicating that NiO NPs varied in size from 6.54 to 9.82 nm.

The additional information related to the detailed morphological features of CN CSNs was investigated using TEM. The TEM images of CN CSNs presented in Fig. 12(a) showed a cubic and spherical shape. The shape of CN CSNs investigated from the TEM image is found to be in good agreement with the XRD data. As shown in Fig. 12(a) the darker area in the CN CSNs image is believed to be Co_3O_4 NPs which is a core and the lighter area shows NiO NPs which is a shell of synthesized particles. This finding is because of the difference in the electron sprinkling between

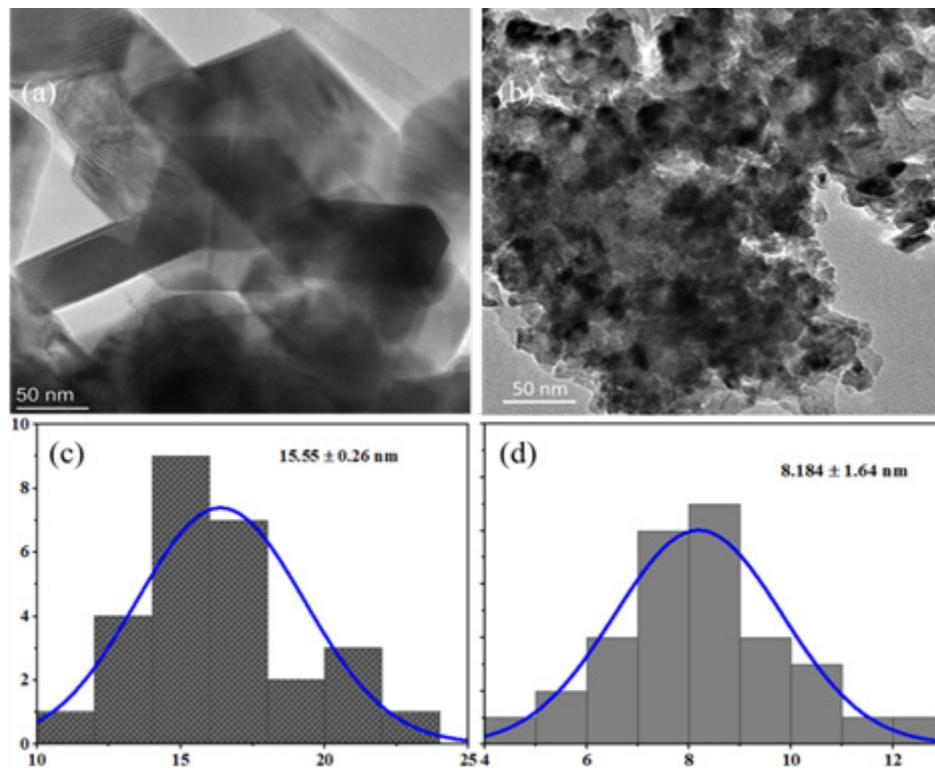


Fig. 11. The TEM image of (a) Co_3O_4 NPs, (b) NiO NPs, and Particle size distribution of (c) Co_3O_4 NPs and (d) NiO NPs

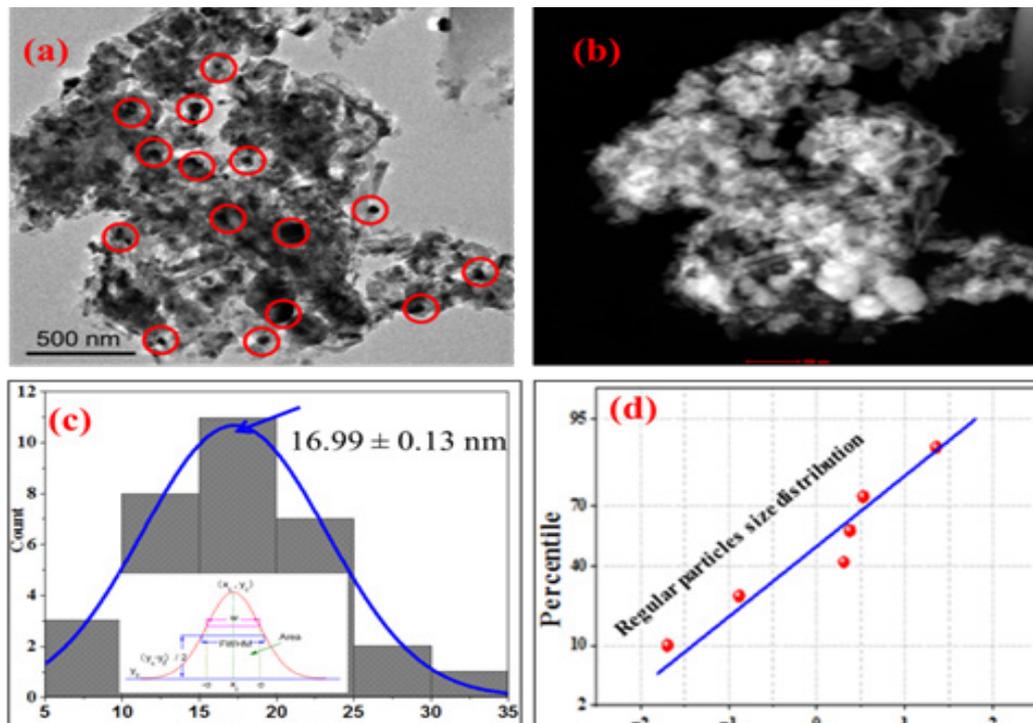


Fig. 12. The CN CSNs analyzed (a) TEM image, (b) STEM image, (c) particle size distribution and (d) regular particle size distributions.

the core and shell particles. Similarly, the scanning transmission electron microscope (STEM) image in Fig. 12(b) indicates the formation of CN CSNs in which the white region shows the high electron density of core and shell materials. The distribution of CN CSNs across the TEM image in terms of their size was also obtained.

The diameter of 60 particles was measured by using Gatan software and the mean particle size was determined from histogram. As depicted in Fig. 12(c) the mean particle size of CN CSNs was found to be 16.99 nm with a standard deviation of 0.13 nm. The range of particle size distribution was within 5-35 nm which implies that the CN CSNs were found to be agglomerated to some extent which might be attributed to the removal of bioactive molecules during calcination. The originally bioactive capped CN CSNs shrunk together after the calcination and resulted in agglomeration.

The grains and lattice fringe alignment of Co_3O_4 NPs was evaluated using HR-TEM as depicted in Fig. 13(b). The structural fingerprints produced by incident electron diffraction after passing through the material were utilized to calculate

the inter-planar distance. The distinctive distance between the fringes was obtained using the IFFT model

The inter-planar distance for Co_3O_4 NPs was determined to be 0.3286 nm, referring to the plane (311). Fig. 13(b) shows the strong visibility and long-range alignment of the structural fingerprinting, showing that the biologically produced Co_3O_4 NPs are single-crystalline for a specific range. The inter-planar distance for Co_3O_4 NPs was determined to be 0.26 nm, referring to the (311) plane.

Furthermore, the crystallinity of Co_3O_4 NPs was investigated using the SAED image displayed in Fig. 13(d). The parallel pattern is associated with electron diffraction by crystal lattice planes. The alignment and brightness of these parallel organized dots reveal the configuration of atoms within the crystal. Thus, it implies that the atoms in biologically produced Co_3O_4 NPs are arranged consistently and have a single crystalline structure.

The crystal structure and lattice parameters of NiO NPs were examined using high-resolution TEM (HR-TEM). Figs 14(a) and 14(b) show the HR-TEM image and the space between the lattice

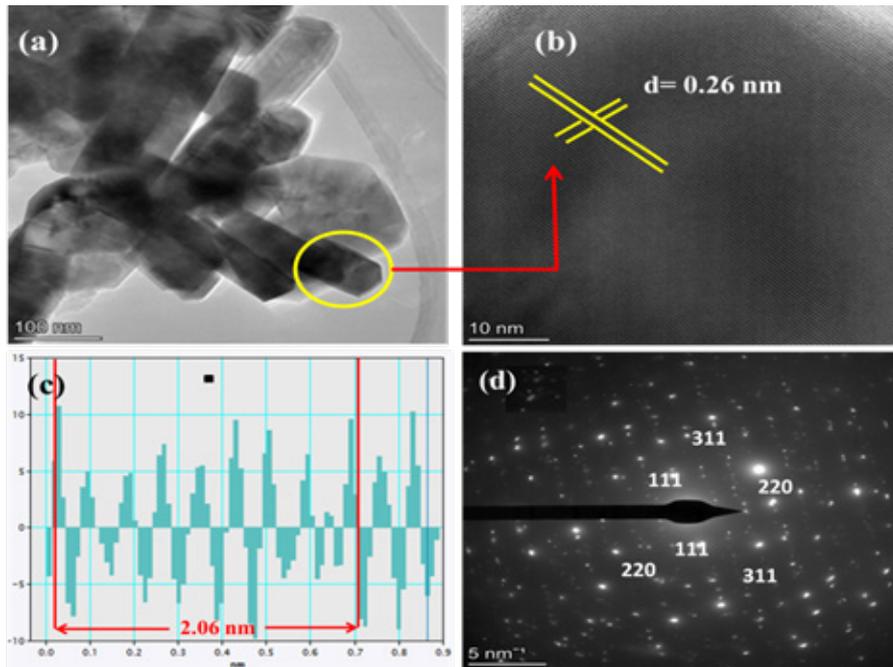


Fig. 13. The morphological images of Co₃O₄ NPs, (a) TEM image, (b) HR-TEM image, (c) inverted Fast Fourier Transform (IFFT), and (d) SAED images

fringes. The d-spacing values shown in Fig. 14(b) were determined using Gatan software. The lattice distance for the respective miller indices (111) was determined to be 0.26 nm. The characteristic (hkl) values of NiO NPs with d-spacing values determined from SAED were in good agreement with the conventional XRD values.

Moreover, the crystal structure of the green

synthesized NiO NPs was determined using SAED images shown in Fig. 14(d). The circular diffraction pattern was generated based on the particle orientation and size. The circular diffraction spot correlates to the crystal structure's lattice points with concentric rings, indicating that the produced NiO NPs are polycrystalline.

The CN CSNs containing Co₃O₄ core and NiO

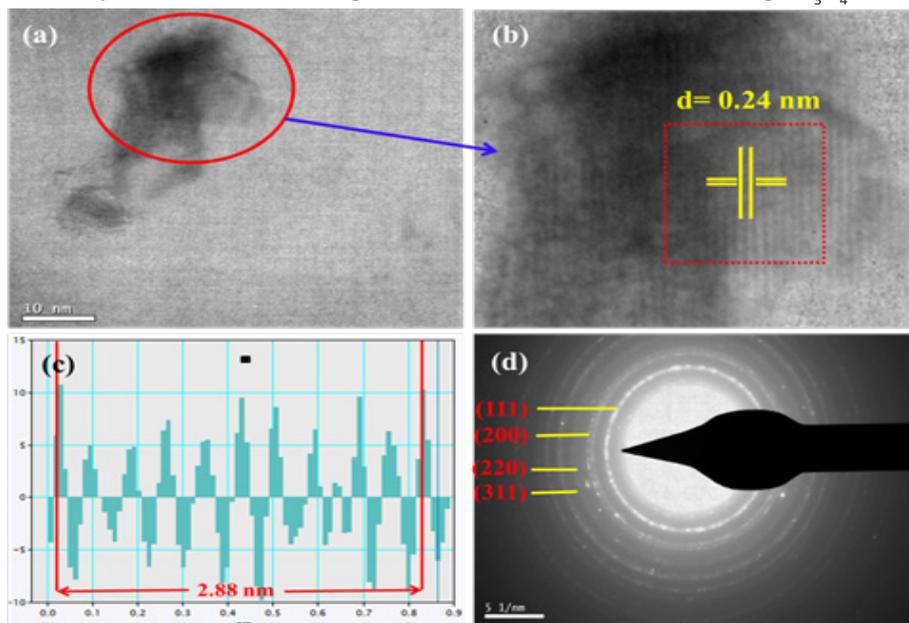


Fig. 14. The morphological images of NiO NPs, (a/b) HR-TEM image, (c) (IFFT), and (d) SAED images

shell were further examined by using HR-TEM. The core-shell structures, interface of core and shell, and the distance between adjacent crystals lattice planes in the nanostructure have been explored. In the analysis, the highly magnified CN CSNs showed the core and shell structure. As shown from the TEM image in Fig. 12(a), the HR-TEM image also showed the dark region that ensures the formation of the core and shell nanostructure. In the area where the NiO shell coats the Co_3O_4 core appeared darker in the HR-TEM image. The darker region is believed to be because of higher electron density in the core, resulting in different electron scattering. Under high-resolution detection with the beam of an electron, the incident electron from the electron gun passes through the core of the CN CSNs sample and experiences strong diffraction because of the high density of the electron. The strong diffraction causes a reduction in the intensity of the electron reaching the detector, resulting in a darker region in the HRTEM image. In contrast, the NiO shell with lower electron density diffracts electrons to some extent whereas most electrons reach the detection and result in brighter regions of the shell

observed in Fig. 15 (a).

On the other hand, the shell material, which has a lower atomic number and electron density, scatters electrons to a lesser extent. As a result, more electrons reach the detector, leading to a brighter appearance in the image. The distance between adjacent crystal lattice planes of the Co_3O_4 core corresponding miller indices (400) was found to be 0.25 nm, which is presented in Fig. 15(b). Similarly, the d-spacing for the NiO shell with miller indices (111), (200), and (220) were 0.25, 0.21, and 0.13 nm respectively. The miller indices obtained from HR-TEM and SAED were found in good agreement with the d-spacing values of standard XRD data.

Moreover, Fig. 15(d) indicated that the SAED image of CN CSNs where miller indices are shown with their corresponding planes. The miller indices were identified from the d-spacing values of the crystal lattice planes of CN CSNs. The calculated lattice space belongs to the miller indices of CN CSNs which are used to confirm the formation of core-shell nanostructure. In addition, the SAED image explored the crystalline structure of green synthesized CN CSNs. As shown in Fig. 15(d), the

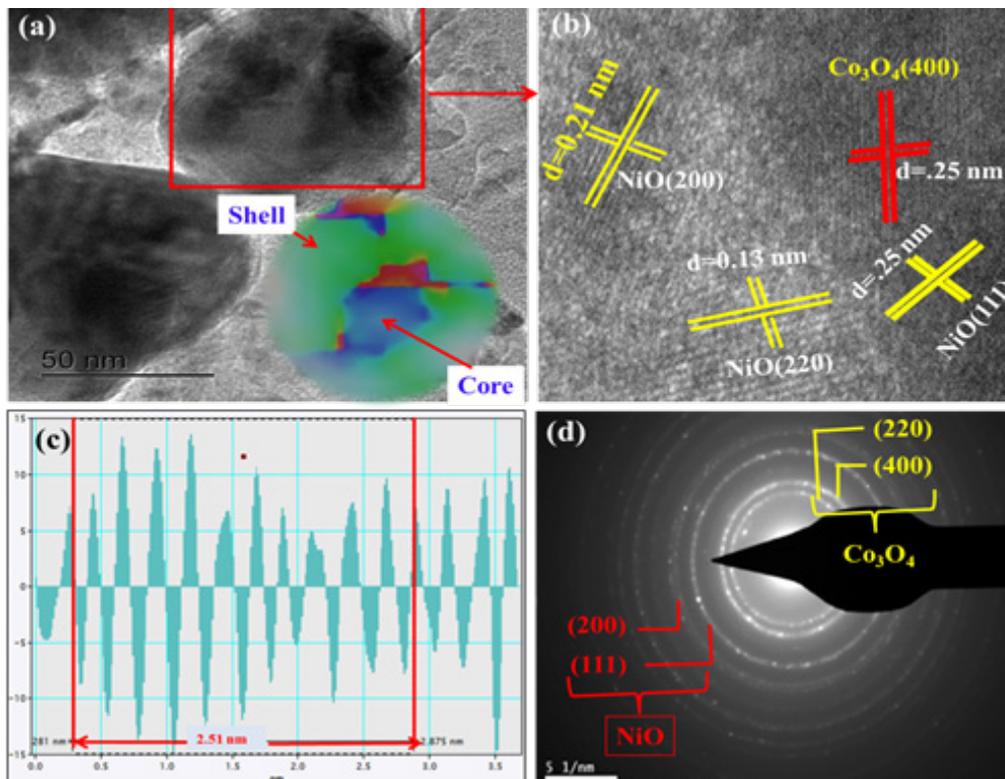


Fig. 15. Biologically synthesized $\text{Co}_3\text{O}_4@NiO$ core-shell (a) HR-TEM, (b) d-spacing obtained from HR-TEM of particles, (c) IFFT, and (d) SAED images.

well-defined ring was revealed that shows the polycrystalline nature of CN CSNs. The nanowire Co₃O₄/NiO with 10 nm thickness of NiO shell was reported in another previous work [35] which requires a combination of high mass-loading and high utilization of electrochemically active material. In this work, we report the fabrication of hierarchical core-shell Co₃O₄@NiO nanowire@nanorod arrays where ultrathin NiO nanorods (~5nm. The fringes with d spacing 0.46 nm correspond to the (111) lattice plane of Co₃O₄, whereas those at 0.24 nm correspond to the (111) lattice plane of NiO. HRTEM data show that Co₃O₄ coexists with NiO [59]. Similarly, the Co₃O₄@NiO CSNs synthesized in the current study had d spacing of 0.36 nm, which could be associated with the (111) lattice plane of the Co₃O₄ core, and 0.25 nm for the NiO shell NPs. The reduced d-spacing of the core and the resulting lattice strain can affect the electrical structure and surface characteristics of nanostructures which could result in increased reactivity and performance in a variety of applications.

Anticancer activities result

The core-shell nanostructures have recently gained attention for their potential anticancer effects due to their biocompatibility, stability, and pharmacokinetics. The two most important conditions for breast cancer therapy are efficacy and selective toxicity. To address these issues, MONPs, the most promising alternative to traditional chemotherapy for breast cancer

treatment, underwent comprehensive testing. Previous studies indicated that ZnO NPs killed 53.3% of MCF-7 cells at a dosage of 100 µg/mL, with an IC₅₀ of 12.7 µg/mL (25). Furthermore, the anticancer properties of Fe₃O₄@ZnO NPs and their parent materials were investigated. Compared to the MONPs, core-shell NPs demonstrated a much stronger cytotoxic effect. This indicates that the properties of the cor-shell nanoparticles have altered, and they outperform single MONPs in all of their activities. Similarly, NiO NP shells were coated onto Co₃O₄ core NPs for breast cancer therapy in this research [63].

The cytotoxicity of CN CSNs, as well as Epirubicin (standard drug), was assessed against breast cancer (MCF-7) cell lines and peripheral blood mononuclear (PBM) cells using an MTT assay. In this work, different exponential concentrations of CN CSNs, as well as Epirubicin of 12.5, 25, 50, 100, 200, 400, and 800 µg/mL have been examined on MCF-7 and PBMC as shown in Fig. 16. In the case of CN CSNs, the in vitro cytotoxicity against MCF-7 cell lines and PBMC healthy cells was examined.

As depicted in Fig. 16, the images of MCF-7 cell lines treated with 100, 200, and 400 µg/mL of CN CSNs and standard drug were taken by using a fluorescence microscope (KFL30LED, India). To determine the cytotoxicity of CN CSNs against the cultured breast cancer cell lines, the micrograph was captured at the wavelength of 520 nm. Consequently, the white (live) and smooth non-white area (dead) cells were observed.

Fig. 17 and Table 1 indicate that CN CSNs inhibit

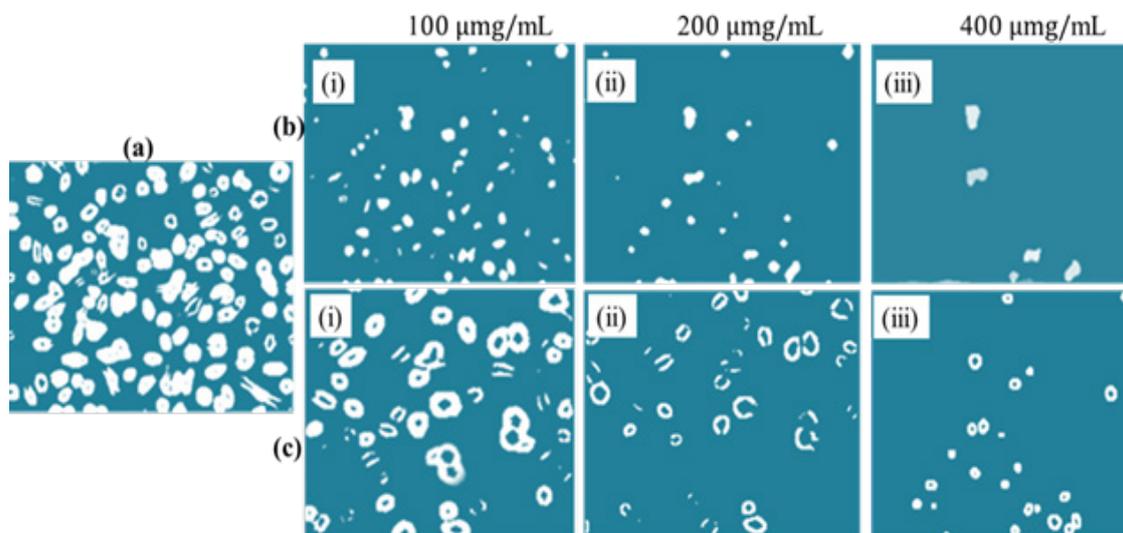


Fig. 16. Cytotoxicity on MCF-7 cell lines: (a) untreated, (b) treated with Epirubicin of 100 µg/mL(i), 200 µg/mL(ii), and 400 µg/mL (iii) and (c) treated with CN CSNs of 100 µg/mL(i), 200 µg/mL(ii), and 400 µg/mL (iii).

cancerous cells at 12.5, 25, 50, 100, 200, 400, and 800 µg/mL by about 22.16, 32.20, 43.62, 52.41, 68.21, 74.11, and 81.38%, whereas Epirubicin drug inhibits 33.42, 46.68, 55.31, 74.53, 81.22, 86.77, and 96.15%, respectively. The CN CSNs and Epirubicin have IC₅₀ values of 73.89 and 15.08 on MCF-7, and 455.86 and 419.40 on PBMC. This suggests that D. stramonium leaf extract-mediated CN CSNs are as effective as conventional drugs in terms of anticancer activity while being less toxic to normal cells.

As shown in Fig. 16, the number of cells decreased as the concentration of CN CSNs against MCF-7 cell lines. The mechanistic biological contact of CN CSNs with MCF-7 may include endocytosis,

in which Co^{2/3+} and Ni²⁺ ions pass through the cell membrane. The tiny size and high surface charge of CN CSNs allow for powerful anticancer action. CN CSNs can produce reactive oxygen species, resulting in oxidative stress and cancer cell death. Elevated ROS levels in the cell can also promote DNA alteration and lesion, which can result in both double and single strand breaks. Table 1 reveals that CN CSNs exhibit substantial toxicity on MCF-7 cell lines, with an IC₅₀ value of 73.89 µg/mL, indicating their potency as an anticancer agent.

Mechanistic Interaction of CSNs with Breast Cancer Cells

The objective of anticancer drugs is to prevent

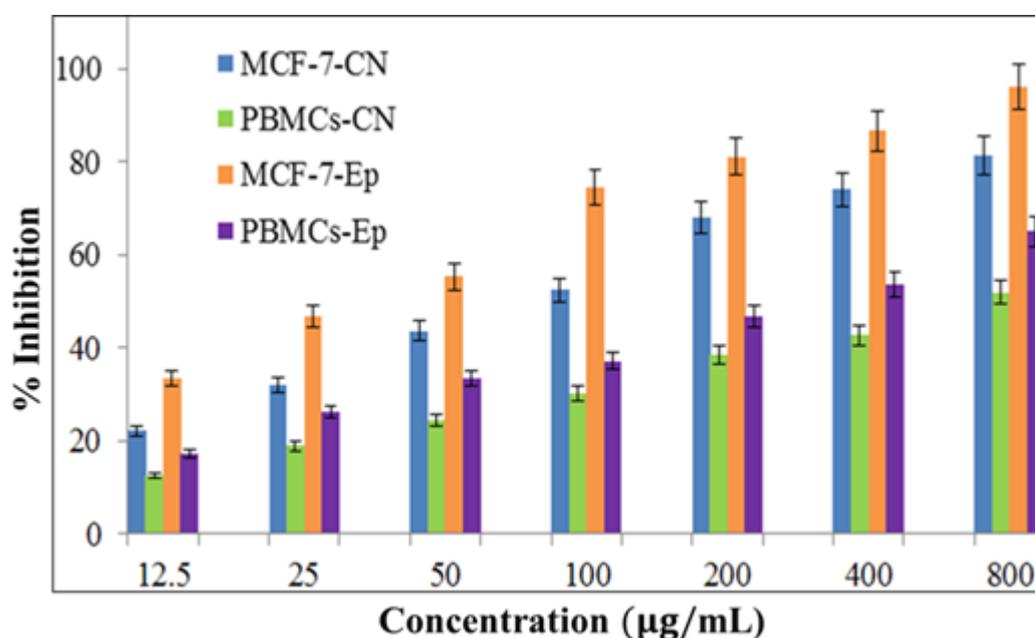


Fig. 17. Cytotoxicity of CN CSNs and Ep against PBM and MCF-7 cell lines

Table 1. The percentage inhibition of CN CSNs and Ep against PBM and MCF-7 cell lines at different concentrations.

Conc. (µg/mL)	Co ₃ O ₄ @NiO (% In)		Epirubicin (% In)	
	MCF-7	PBMCs	MCF-7	PBMCs
DMSO	0.00	0.00	0.00	0.00
12.5	22.16 ± 0.03	12.57 ± 0.02	33.42 ± 0.03	17.13 ± 0.01
25	32.20 ± 0.02	18.88 ± 0.02	46.68 ± 0.01	26.33 ± 0.03
50	43.62 ± 0.00	24.44 ± 0.01	55.31 ± 0.02	33.56 ± 0.02
100	52.41 ± 0.01	30.21 ± 0.01	74.53 ± 0.03	37.23 ± 0.00
200	68.21 ± 0.03	38.54 ± 0.03	81.22 ± 0.01	46.66 ± 0.03
400	74.11 ± 0.03	42.67 ± 0.04	86.77 ± 0.00	53.52 ± 0.02
800	81.38 ± 0.01	51.94 ± 0.02	96.15 ± 0.02	65.18 ± 0.04
IC ₅₀	73.89	455.86	15.08	419.40

tumor cell growth and cause death without a significant negative impact on healthy cells. The specific molecular mechanisms involved in the interaction between nanoparticles and breast cancer cells can vary depending on the type of nanoparticle, its physicochemical properties, the breast cancer cell line, and the experimental conditions [64-65]. The CN CSNs having unique physicochemical features have emerged as a new hope for cancer therapy. By delivering drugs directly to cancer cells, anticancer treatment can be more effective while minimizing adverse effects. The CN CSNs inhibit and induce death by oxidative stress, causing DNA damage, arresting the cell cycle, and inducing cancer cell death by apoptosis as well as nonapoptotic modes of cell death.

In cancer cell proliferation, cyclin-dependent kinase 1/2 (CDK1/2) plays a crucial role in the whole cell cycle. It has become a new target for cancer therapy. CDK1 interacts with cyclin B1 to facilitate the transition from the growth (G2) phase into mitosis, whereas CDK2 activates the growth 1/synthesis (G1/S) and synthesis/growth 2 (S/G2) transitions in the interphase stage of cell cycle. As shown in Fig. 18, the delivered metal ion of CN CSNs believed to inhibit the activity of CDKs

by binding to groups such as imidazole nitrogen of histidine, amide nitrogen atoms of asparagine and glutamine, carboxylate oxygen atoms of aspartic acid and glutamic acid, hydroxyl oxygen atoms of serine, threonine, and tyrosine residues, and thiol group of cysteine amino acids. Similarly, particular elements inside DNA polymerase, such as the nitrogen, oxygen, and sulfur atoms of amino acids, reacted with metal ions found in CN CSNs, inhibiting its action. The other key mechanism of CSNs to induce the death to cancer cells is producing Reactive oxygen species (ROS). Once the CN CSNs are internalized by the cancer cell through the endocytic pathway and dissolved into metal ions, and disrupts the cellular homeostasis. This homeostasis disruption, resulted in the production of excessive ROS within the cancer cells, leading to oxidative stress and mitochondrial dysfunction.

Additionally, the core-shell nanoparticles are recognized and taken up by the breast cancer cells through receptor-mediated endocytosis. Once the CN CSNs are internalized, they become entrapped within the endocytic vesicles, which then fuse with lysosomes to form endolysosomes as shown in Fig. 18. The CN CSNs is released into the cytoplasm once the endolysosomal membrane is disrupted,

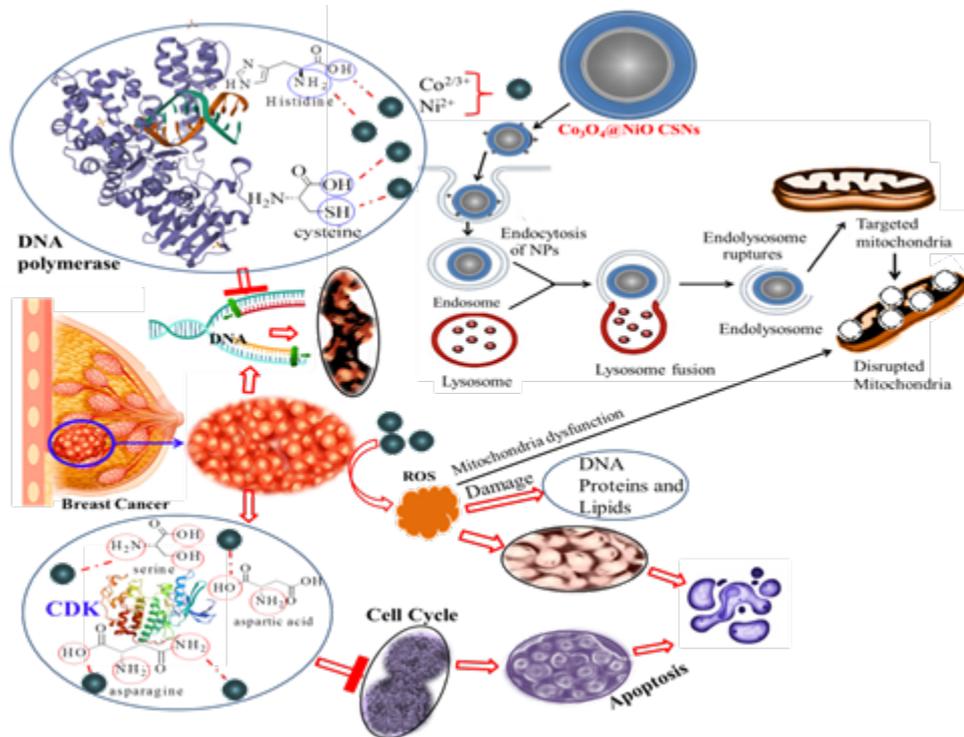


Fig. 18. Mechanistic interaction of CN CSNs with breast cancer cells

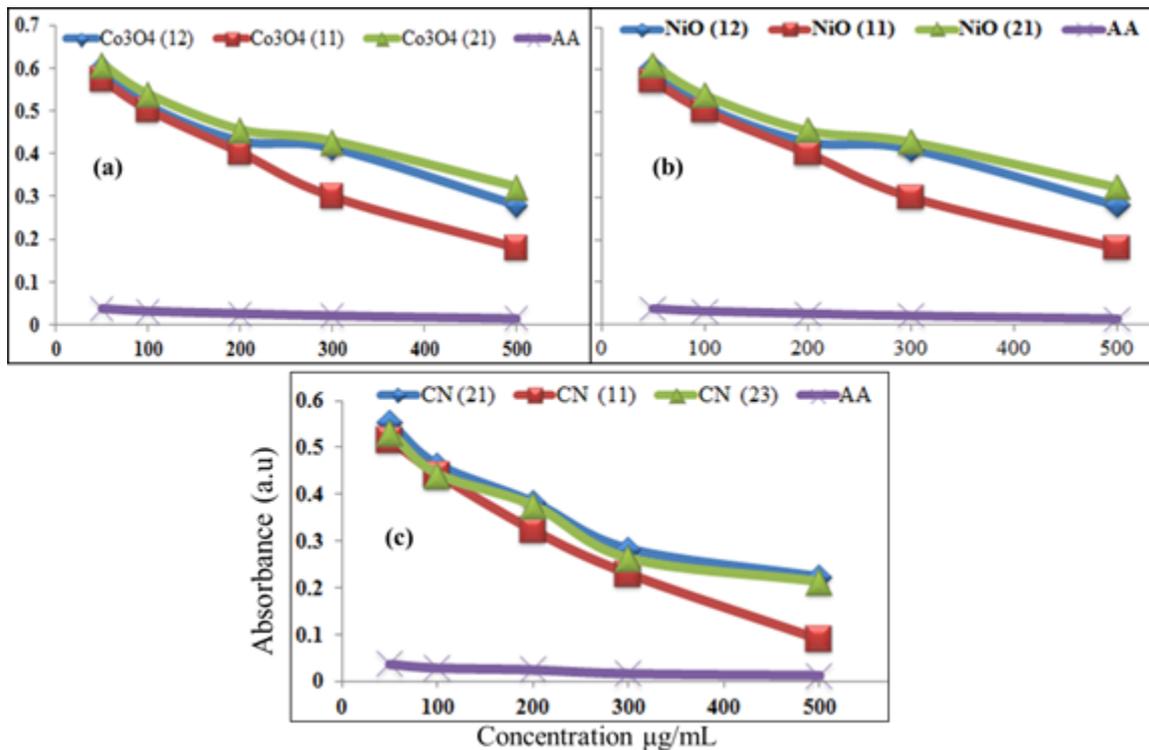


Fig. 19. The absorbance of (a) Co₃O₄ NPs, (b) NiO NPs, and (c) CN CSNs at concentrations of (50, 100, 200, 300, and 500 µg/mL).

and then the drug is targeted to the mitochondria and leads to cell death [66]

Antioxidant activities analysis result

The scavenging activity of *D. stramonium* leaf extract-mediated Co₃O₄ and NiO NPs, as well as CN CSNs, was tested against the stable free radical 2, 2-diphenyl-1-picrylhydrazyl (DPPH) using spectroscopic techniques. The antioxidant activity test of Co₃O₄ and NiO NPs, as well as CN CSNs at 50, 100, 200, 300, and 500 µg/mL concentrations, were determined. The ability of these metal oxide nanoparticles to donate electrons is believed to be the basis for their effect on DPPH. The samples introduced into the solution reduce a violet DPPH to a yellow color depending on concentration. As shown in Fig. 19, the absorbance of the reaction mixture at 517 nm was measured using a spectrophotometer and displayed as a function of MONPs and CSN concentration.

The absorbance value reflects the level of DPPH reduction by Co₃O₄ and NiO NPs, as well as CN CSNs are used to assess their antioxidant activity. As depicted in Fig. 19, the absorbance of the reference medication (ascorbic acid), MONPs, and CSNs containing DPPH at 517 nm decreased

as their concentration increased. The drop in absorbance suggests that the MONPs and CSNs are successfully neutralizing the DPPH free radical, validating their antioxidant activity. MONPs and CSNs have increased antioxidant action as their absorbance decreases. Furthermore, because of their significant band gap energy, MONPs with small sizes decrease absorbance quicker than those with large sizes. This ensured that the small-sized particles were powerful antioxidants. Furthermore, because of their high band gap energy, MONPs with small sizes lose absorbance more quickly than those with large sizes. This guaranteed that the small particles were effective antioxidants. Finally, as shown in Figs 19 (a, b, and c), the absorbance order of MONPs synthesized at three volume ratios is (21) > (12) > (11) and CSNs at (21) > (23) > (11) concentration ratios.

The percentage scavenging effect of Co₃O₄ and NiO NPs against DPPH was measured to determine their antioxidant activity. The percentage scavenging indicates the amount to which these MONPs scavenge the DPPH radicals. Fig. 20 (a) and Table 2 demonstrate that Co₃O₄ NPs produced in a (11) volume ratio inhibit 9.230, 22.564, 37.949, 53.846, and 63.308% at 50, 100, 200, 300,

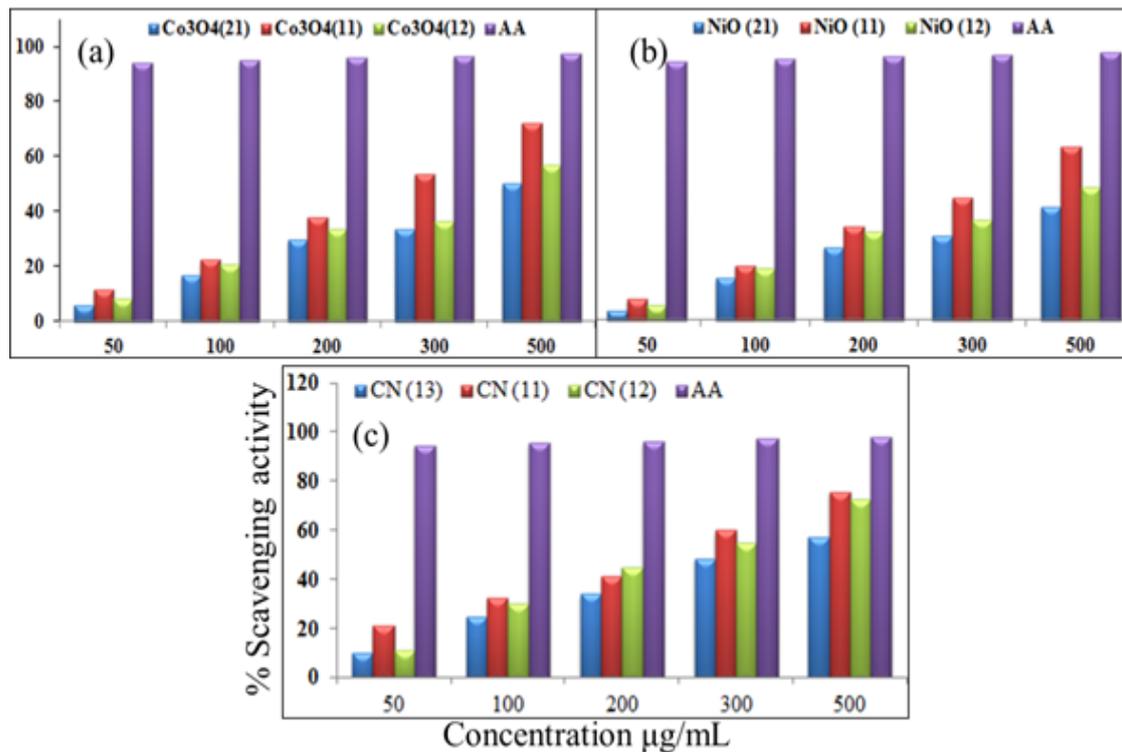


Fig. 20. The antioxidant activity of (a) Co3O4, (b) NiO, and (c) CN CSNs

and 500 µg/mL, respectively. As the concentration of Co3O4 NPs increases, so does their capacity to scavenge DPPH radicals. Additionally, the percentage scavenging of Co₃O₄ NPs synthesized in (12) was also found to be 4.103 (50), 15.385 (100), 28.205 (200), 36.410 (300), and 54.359% (500 µg/mL), whereas (21) scavenge 4.103 (50), 15.385 (100), 28.205 (200), 36.410 (300), and 54.359% (500 µg/mL). Among the three ratios, Co3O4 (11) NPs showed good scavenging potential against the DPPH radical. Similarly, the scavenging activity of NiO NPs was studied and their activity increases with concentration. The scavenging activity of NiO NPs synthesized in three volume ratios was raised in the order of (2:1), (1:2), and (1:1). At 500 µg/mL, Co₃O₄ (11) and NiO (11) NPs demonstrated scavenging activity about 63.308 and 57.0769%, respectively. Co₃O₄ NPs are commonly reported to have stronger antioxidant activity than NiO NPs. This is mostly due to the high redox potential of Co₃O₄ NPs. Co3O4 NPs with high redox potential are more likely to transfer electrons to DPPH, preventing oxidative damage.

Similarly, the antioxidant activity of CN CSNs was investigated as shown in Fig. 20(c) Table 2.

The percentage scavenging activity of CN (11) at 50, 100, 200, 300, and 500 µg/mL was found to be 15.07, 26.35, 38.18, 56.29, and 65.98%. The result of the percentage scavenging activity achieved by CN CSNs was more than that of the parent material. The potent antioxidant activity of CSNs is believed to be due to the synergistic effect, redox potential, and surface area. The shell NiO NPs enhance the overall antioxidant of core-shell nanostructures which is a result due to the synergistic effects [31]. Similarly, the shell materials increase the core's redox potential and surface area, allowing for effective electron transport between the Co3O4 core and the NiO NPs shells, thus enhancing DPPH scavenging capabilities [67].

The IC₅₀ values for DPPH radical scavenging activity of Co₃O₄ (11) NPs, NiO (11) NPs, CN11, and ascorbic acid were found to be 158.11, 165.62, 111.11, and 44.48 µg/mL, respectively. The CN11 with an IC₅₀ of 111.11 µg/mL scavenged DPPH radicals by about 50%. This means the study shows that CN CSNs have a higher scavenging activity than individual Co₃O₄ and NiO NPs.

The mechanistic interaction of MONPs and CSNs with DPPH involves supplying electrons to the

Table 2. Percentage radical scavenging and IC₅₀ of Co₃O₄, NiO NPs, and CN CSNs

Concentration (µg/mL)		50	100	200	300	500	IC ₅₀
NPs	Ratios						(µg/mL)
Co ₃ O ₄	2:1	4.102	13.333	29.744	33.846	47.589	262.21
	1:1	9.230	22.564	37.949	53.846	63.308	158.11
	1:2	4.103	15.385	31.205	36.410	54.359	239.34
NiO	2:1	3.589	15.385	26.666	30.769	41.538	290.01
	1:1	7.692	20.002	34.359	44.615	57.0769	165.62
	1:2	5.641	18.974	32.3077	36.410	48.718	275.62
CN	CN21	10.29	23.01	34.31	48.53	57.52	130.61
	CN11	15.07	26.35	38.18	56.29	65.98	111.11
	CN23	9.27	20.93	31.09	44.91	51.84	144.47
AA		94.154	95.077	96	96.769	97.846	44.84

DPPH radical, to reduce the nitrogen atom as seen in Fig. 21. According to the mechanism outlined by F. Rahman, the unstable purple-colored DPPH• solution was converted into a yellow stable DPPH reduced form during the DPPH• free-radical scavenging process [22, 68].

CONCLUSION

In this work, green synthesis was preferred to conventional techniques as it adopts a more ecologically conscious and sustainable method by embracing concepts of environmental

responsibility. Co₃O₄ and NiO NPs as well as Co₃O₄@NiO CSNs were synthesized by green method utilizing their respective precursor salts and *Datura stramonium* plant leaf extract. The synthesis of the Co₃O₄, NiO NPs and Co₃O₄@NiO CSNs was confirmed by characterization techniques such as FTIR, UV-Vis, XRD, SEM-EDAX, TEM-HRTEM, and SAED. The Co₃O₄ NPs, NiO NPs, and CN CSNs became thermally stable at above 350, 380, and 400, respectively. This suggests that the core-shell architecture improves the thermal stability of the CN CSNs when compared

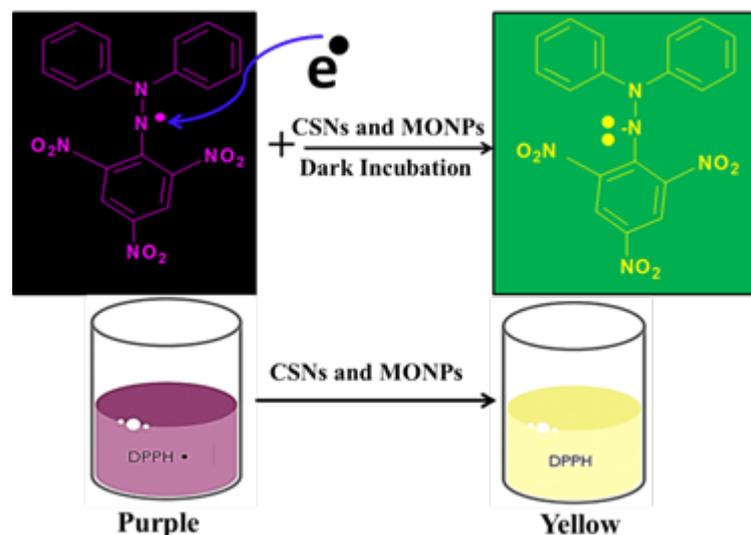


Fig. 21. Mechanistic interaction of electron of MONPs and CSNs against DPPH free radical molecule

to individual Co_3O_4 and NiO nanoparticles. The average crystalline size of CN21, CN11, and CN23 were 14, 13, and 16 nm, respectively. The number of Diffraction peaks in the shell material exceeds that of the core materials concerning the individual nanoparticles. This ensured that the NiO NPs were produced on the surface of Co_3O_4 NPs as a shell, consistent with prior results. The mean particle size of cubic CN CSNs obtained from the TEM image was 16.99 nm. Furthermore, the formation of CN CSNs was confirmed by miller indices obtained from SAED that correspond to the plane of core and shell particles. The cytotoxicity of green synthesized CN CSNs against breast cancer (MCF-7) cell lines at 12.5, 25, 50, 100, 200, 400, and 800 $\mu\text{g}/\text{mL}$ exponential concentrations were 22.16, 32.20, 43.62, 52.41, 68.21, 74.11, and 81.38%, respectively. The results show that the percentage inhibition of CN CSNs on MCF-7 cell lines rises with increasing doses. The IC_{50} values for CN CSNs and Epirubicin on MFC-7 are 73.89 and 15.08, respectively, whereas on PBMC they are 455.86 and 419.40. The result evidenced that *D. stramonium* leaf extract-mediated CN CSNs have comparable anticancer activity against breast cancer (MCF-7) cell lines to the common medicines in use while being less hazardous to normal cells. To fully realize the promising .potentail of the $\text{Co}_3\text{O}_4@/\text{NiO}$ core-shell nanostructure in breast cancer treatment, future study that comprises cellular mechanistic interaction, further precursor optimization, and in-depth biological evaluation, might be needed.

DATA AVAILABILITY

The data used to support the study's findings are accessible from the relevant authors upon request.

ETHICAL APPROVAL

This article does not include any experiments with human participants or animals conducted by the authors.

CONFLICTS OF INTEREST

The authors state that they have no conflicts of interest.

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AUTHORS' CONTRIBUTIONS

Gezahegn Tadesse, Prof. Tegene Desalegn, and Prof. H.C. Ananda Murthy were in charge of conceptualizing, designing the methodology, analyzing the data, and writing the manuscript. Lema Teshome and Addisalem Abebe contributed to the sample characterization process. All authors have read, revised, and provided feedback on the paper.

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