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Carvacrol and *Zataria multiflora* influenced the PPARy agonist effects on systemic inflammation and oxidative stress induced by inhaled paraquat in rat

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ARTICLEINFO	A B S T R A C T		
<i>Article type:</i> Original article	Objective(s) : The effects of PPAR- γ agonist alone and in combination with carvacrol and Zataria multiflora on inhaled paraquat (PQ) induced-systemic inflammation and oxidative stress were		
<i>Article history:</i> Received: Jan 19, 2020 Accepted: Mar 11, 2020	examined. <i>Materials and Methods:</i> Control group exposed to normal saline aerosol, one group exposed to 54 mg/m ³ PQ aerosol and four groups exposed to PQ aerosol and treated with 5 mg/kg/day pioglitazone, pioglitazone + 200 mg/kg/day Z. <i>multiflora</i> extract, pioglitazone + 20 mg/kg/day carvacrol, and 0.03 mg /kg/day dexamethasone for 16 days after the end of exposure to PQ were		
<i>Keywords:</i> Carvacrol	studied. Exposure to normal saline or PQ was performed every other days for 30 min (8 times). Different variables were measured after the end of treatment period.		
Oxidative stress Paraquat PPAR-γ agonist Systemic inflammation Zataria multiflora	Results: PQ exposure significantly increased serum levels of NO ₂ , MDA and IL-6 but dexreased CAT and IFN- γ levels and IFN- γ /IL-6 ratio compared to control group (<i>P</i> <0.01 to <i>P</i> <0.001). Treatment with pioglitazone only improved serum level of MDA (<i>P</i> <0.01). Treatment with combination of pioglitazone and carvacrol as well as treatment with dexamethasone improved all measured variables compared to PQ exposed group (<i>P</i> <0.05 to <i>P</i> <0.001). The effects of pioglitazone + Z. <i>multiflura</i> and pioglitazone + carvacrol on almost all measured variables were significantly higher than pioglitazone alone (<i>P</i> <0.05 to <i>P</i> <0.001).		
	Conclusion: The effects of combination therapy of pioglitazone with Z. <i>multiflora</i> or carvacrol on inhaled paraquat (PQ) induced-oxidative stress and systemic inflammation were higher than the effects of pioglitazone alone. These results suggested that the effects of the extract and carvacrol may mediated through PPAR-y receptors.		

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Introduction

Although agrochemicals such as herbicides, fertilizers and pesticides are an important factor in increasing the products yield in agriculture, negative effects of these compounds on human health is undeniable (1). Paraquat (PQ) is a nonselective herbicide which inducing inflammation and oxidative stress in various organs (2). This herbicide is highly used for control of more than 100 different crops including corn, rice, soybeans, vegetables, orchards, and many other crops in about 100 countries in the global agricultural field, especially in Asian countries (3).

Paraquat is used as herbicide in agriculture in nearly 130 countries, especially in third world countries and the rate of death from pesticide poisoning is generally, 250,000 to 370,000 each year. The disability-adjusted life years (DALY) in the northeast of Colombia by PQ poisoning was around 53.4 years per 100,000 inhabitants in 2019 but in 2015 it was higher (26,900 DALYs per 100,000), (4). In Iran, the incidence of acute PQ poisoning from 2004 to 2013 was 30,485 (3049 per year) which was increased from 1808 in 2004 to 4283 in 2013 and years of life lost (YLL) was 20,709 in this

period of time (5).

It was shown that PQ is highly toxic for human and most animals (6). Small dose of PQ (30 mg/kg) is lethal in adult human (7) due to generation of intracellular reactive oxygen species, redox reactions and lipid peroxidation of cellular membranes (8). Systemic inflammation induced by inhaled PQ is related to actively absorbance of its particles which causes leukocytosis, pulmonary hypertension, heart enlargement, acute renal damage, edema and increasing amylase, glucose and creatinine (9). It has also been shown that exposure to PQ can lead to an increase in inflammatory factors such as interleukins and TNF- α (9). PQ poisoning and its lethal effects makes it become a severe public health challenge. Therefore, preventing systemic absorption and the effect of PQ is one of the important issues in human health. The current treatment of acute PQ poisoning is the prescribing activated charcoal. Moreover, some anti-oxidants- such as acetylcysteine and salicylate, antiinflammatory and immunosuppressive agents are also used as potential treatments for PQ poisoning (8).

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There is no effective therapeutic drugs has yet been introduced to prevent PQ-induced systemic

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toxicity (3, 10). However, the available treatment for PQ poisoning includes using immunosuppressive drugs such as dexamethasone, cyclophosphamide and methylprednisolone. Acetylcysteine and salicylate as anti-oxidant agents are also of therapeutic values by reducing free radical scavenging, anti-inflammatory and NF-kB inhibitory actions. However, the therapeutic efficacy of these drugs are very weak and despite the mentioned treatments, there is a high rate of mortality due to acute PQ poisoning (11).

Peroxisome proliferator-activated receptors (PPARs) are nuclear, ligand-activated transcription factors receptor that have effective role in improvement of various diseases such as inflammatory disorders, cancer and lung diseases (12). Up to now, three PPAR subtypes have been identified including: PPARa, PPAR β/δ , and PPAR γ (13). PPAR γ receptor is the main target of thiazolidinediones (TZDs). This drugs are frequently used to combat the hyperglycemia state which can be associated with metabolic syndrome (14). It was shown that some natural or synthetic PPARy ligands such as prostaglandins, eicosanoids, fatty acids, leukotrienes, rosiglitazone, and pioglitazone (15) are capable regulating cellular metabolism, differentiation, apoptosis and inflammation via modulating of different gene products (14, 16).

Zataria multiflora Boiss (Z. multiflora) belongs to the Lamiaceae family. This medicinal herb grows only in the south and the central regions of Iran, Afghanistan and Pakistan (17). Traditionally, this plant have been used as a remedy against the lung inflammation (18). Z. multiflora extract contains carvacrol and thymol, two active ingredients with strong anti-oxidant properties that have the potential to be used in inflammatory and immune deficiency diseases associated with increased oxidative stress (18). Z. multiflora and its constituent carvacrol also showed anti-oxidant and anti-inflamatory effects by scavenging free radicals, as well as the protective effects on malondialdehyde (19).

Although the effects of PPAR γ agonists, *Z. multiflora* extract and carvacrol on the treatment of inflammatory and immune-dysregulation disorders were reported, no available study has evaluated the effects of these compounds on systemic toxicity induced by PQ inhalation. Hence the aims of the present study are exploring the potential effects of PPAR γ agonist alone and in combination with *Z. multiflora* hydroalcoholic extract and carvacrol on the PQ inhalation induced systemic toxicity in the male Wistar rat.

Materials and Methods

Animal and experimental groups

Thirty male Wistar rats (weighing 200-250 g) were

purchased and keep in the animal house, School of Medicine, Mashhad University of Medical Sciences, Iran in standard condition (22±2 °C temperature, 12 hr light/dark cycle and diet and tap drinking water *ad libitum*). In the present study only male rats with weight range of 200-250 g were studied as inclusion criteria, and female rats with low or high weight were considered as exclusion criteria. Experiments were done according to criteria outlined in the Guide for Care and Use of Laboratory Animals (NIH US publication 23-68

of Mashhad University of Medical Sciences for Animal Experiments (code 961202). Rats were randomly assigned into six groups (n=5 in each group) as: 1) Exposed rats to aerosol of normal saline, every other day for 16 days, 8 times, (control group). 2) Exposed rats to aerosol of PQ (Sigma Aldrich Co, China), (20). 3) Exposed rats to aerosol of PQ and treated with 5 mg/kg/day pioglitazone (Samisaz Pharmaceutical Company, Iran), by gavage for 16 days after the end of PQ exposure period (21). 4) Exposed rats to aerosol of PQ and treated with pioglitazone + 200 mg/ kg/day Z. multiflora hydroalcoholic extract by gavage for 16 days after the end of PQ exposure period. 5) Exposed rats to aerosol of PQ and treated with pioglitazone + 20 mg/kg/day carvacrol by gavage for 16 days after the end of PQ exposure period (22). 6) Exposed rats to aerosol of PQ and treated with 0.03 mg/kg/day dexamethasone (Sigma, St. Louis, MO, Germany), by gavage for 16 days after the end of PQ exposure period (23). Carvacrol and pioglitazone were diluted in saline by adding few drop of Tween-80 to make their appropriate concentrations for gavage. In control and PQ exposed groups also the same volume of saline (5 ml) as the extract, carvacrol and pioglitazone was gavaged in the same manner as treated groups.

revised 1985; ttp://oacuod.nih.gov/regs/guide/guidex.

htm). The study was approved by the Ethics Committee

Exposure to paraquat

PQ aerosol was delivered to animal head box dimensions $15 \times 18 \times 30$ cm for eight times in one other day order, totally for 16 days (for 30 min/day). PQ aerosol was produced by a nebulizer (Omron CX3, Japan, particle size 3-5 µm) with an air flow of 8 lit/min. Each time, 4.5 ml of 1.33 mg/ml PQ (Sigma Aldrich Co, China) solution was added to the nebulizer chamber. The solution volume output of the nebulizer was 0.15 ml/ min and its air output was 3.7 l/min. Therefore the PQ dose in the aerosol was 54 mg/m³ (20, 23). Pioglitazone alone or in combination with *Z. multiflora* extract or carvacrol as well as dexamethasone were daily gavaged after the end of PQ exposure for16 days. In control group, saline was used instead of the PQ solution (24), (Figure 1).

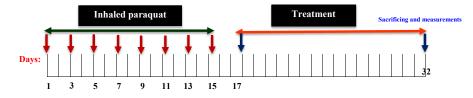


Figure 1. Protocol of exposing animals to inhaled PQ and treatment with pioglitazone or combination of pioglitazone with *Z. multiflora* or carvacrol PQ: Paraquat

Table 1. Studied groups, their exposing to saline or paraquate and treatment protocol

Froups	Exposed to		Treatment with	
	Agent and duration	Dose	Agent and duration	Dose
Control	Salin aerosol for 30/day min, 8 times in every	-	•	-
	other day order over 16 days			
PQ	Paraquat aerosol for 30/day min, 8 times in	54 mg/m ³	-	-
	every other day order over 16 days			
PQ-Pio	Paraquat aerosol (54 mg/m ³) for 30 min, 8 times	54 mg/m ³	Pioglitazone for 16 days after the end	5 mg/kg/day
	in every other day order over 16 days		of PQ exposure	
Pio+Z	Paraquat aerosol for 30 min/day, 8 times in	54 mg/m ³	Pio + Z. multiflora extract for 16 days	Pio + 200 mg/kg/day
	every other day order over 16 days		after the end of PQ exposure	
Pio+Carv	Paraquat aerosol for 30 min/day, 8 times in	54 mg/m ³	Pio + Carvacrol for 16 days after the	Pio + 20 mg/kg/day
	every other day order over 16 days		end of PQ exposure	
PQ-Dexa	Paraquat aerosol for 30 min/day, 8 times in	54 mg/m ³	Dexamethasone for 16 days after the	0.03 mg/kg/day
	every other day order over 16 days		end of PQ exposure	

Plant and extract

The plant was collected (Herbarium No: 35314, FUMH) and the aqueous-ethanolic extract of the plant was prepared as described previously (25). Dried extract was added to water in order to prepare daily fresh extract for gavage. In our previous study, the characteristic of the extract of *Z. multiflora* was identified (26).

Oxidant, and anti-oxidant biomarkers measurement

Serum levels of oxidants including MDA and NO_2 were measured as described in a previous study (27).

The activity of CAT enzyme in serum as an antioxidant marker was measured based on the method described previously (27).

Cytokines measurement

Serum levels of IL-6 and IFN- γ were measured using specific enzyme-linked immunosorbent assay (ELISA) kits (Hangzhou Eastbiopharm, Iran) based on the manufacture technique as previously described (28).

Statistical analysis

The results were presented as mean±SEM and were analyzed using one-way analysis of variance (ANOVA) followed by Tukey's multiple comparison test. Statistically significance were considered when *P*-value was less than 0.05 or lower.

Results

The effects of paraquat exposure

Inhaled PQ (54 mg/m³) caused significant increase in NO₂ and MDA serum levels but decrease CAT activity compare to control group (P<0.01 for NO₂, P<0.001 for MDA and CAT). Exposure of rats to inhaled PQ (54 mg/ m³) caused significant increased IL-6 level but decreased INF- γ level and IFN- γ /IL-6 ratio compared to control group (P<0.01 for IL-6 and IFN- γ /IL-6 ratio but P<0.001 for IFN- γ), (Figures 2-7).

The effects of pioglitazone

Treatment with pioglitazone (5 mg/kg) significantly decrease serum level of MDA compared to PQ group (P<0.01). However, other measured parameters did not

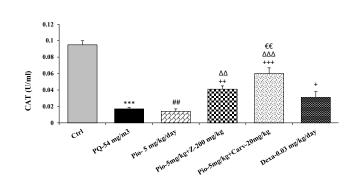


Figure 2. Serum level of catalase (CAT) in different studied groups. The results are expressed as mean±SEM (n=5 in each group). ***P<0.01 compared to the control group. ++P<0.01 and +++P<0.001compared to the PQ group, ##P<0.01 compared to dexamethasone treated group. $\Delta\Delta P$ <0.01 and $\Delta\Delta\Delta P$ <0.001 compared to pioglitazone treated group. $\in \mathbb{E}P$ <0.01 compared to group treated with combination of pioglitazone ant the extract. Comparisons between different groups were made using one-way ANOVA followed by Tukey's multiple comparison test. Ctrl, control group, PQ-54 mg/m3, group exposed to paraquat aerosol with dose of 54 mg/m3, Pio-5 mg/kg/day, Pio-5 + 2-200 mg/kg/day, Pio-5 + Carv-20 mg/kg/day and Dex-0.03mg/kg/ day, gioglitazone, combination or 200 Zataria multiflora and 20 mg/kg/ day and carvacrol respectively or dexamethasone respectively

significantly improved due to pioglitazone treatment compared to PQ group (Figures 2-7).

The effects of combination treatment with pioglitazone and the extract or carvacrol

The combination therapy of pioglitazone and *Z.* multiflura extract improved serum levels of CAT, NO₂, MDA and IL-6 compared to PQ exposed group (P<0.001 for MDA and P<0.01 for other cases), (Figures 2-7). However, treatment with combination of pioglitazone and carvacrol improved all measured variables compared to PQ exposed group (P<0.05 for IFN- γ /IL-6 ratio and P<0.001 for other cases), (Figures 2-7).

The effects of combination therapy of pioglitazone and *Z. multiflura* extract on serum levels of CAT and INF- γ as well as IFN- γ /IL-6 ratio were higher than



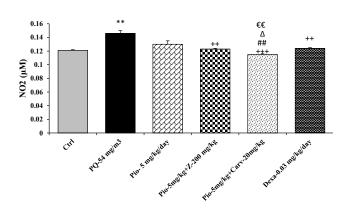


Figure 3. Serum level of nitrite (NO2) in different studied groups. The results are expressed as mean±SEM (n=5 in each group). ***P*<0.01 compared to the control group. ++*P*<0.01 and +++*P*<0.001 compared to the PQ group. ##*P*<0.01 compared to dexamethasone treated group. ΔP <0.05 compared to pioglitazone treated group. €€*P*<0.01 compared to group treated with combination of pioglitazone ant the extract. Comparisons between different groups were made using one-way ANOVA followed by Tukey's multiple comparison test. Ctrl, control group, PQ-54 mg/m3, group exposed to paraquat aerosol with dose of 54 mg/m³. Pio-5 mg/kg/day, Pio-5 + Carv-20 mg/kg/day and Dex-0.03mg/kg/day, groups exposed to PQ-54 mg/m³ and treated with 5 mg/kg/day and carvacrol respectively or dexamethasone respectively

treatment effect of pioglitazone alone (P<0.01 for all cases), (Figures 2, 6 and 7). All measured variables in the treatment group with combination of pioglitazone and carvacrol were significantly higher than pioglitazone alone treated group P<0.05 for NO₂, MDA, IL-6 and P<0.001 for other cases), (Figures 2-7). In addition, the effects of treatment with combination of pioglitazone

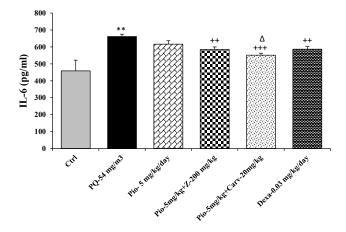


Figure 5. Serum level of interlocking-6 (IL-6) in different studied groups. The results are expressed as mean±SEM (n=5 in each group). ***P*<0.01 compared to the control group. ++*P*<0.01 and +++*P*<0.001 compared to the PQ group. ΔP <0.05 compared to pioglitazone treated group. Comparisons between different groups were made using one-way ANOVA followed by Tukey's multiple comparison test. Ctrl, control group, PQ-54 mg/m³, group exposed to paraquat aerosol with dose of 54 mg/m³, pio-5 mg/kg/day, Pio-5 mg/kg/day, Pio-5 + Carv-20 mg/kg/day and Dex-0.03mg/kg/day, groups exposed to PQ-54 mg/m³ and treated with 5 mg/kg/day pioglitazone, combination or 200 *Zataria multiflora* and 20 mg/kg/day and carvacrol respectively or dexamethasone respectively

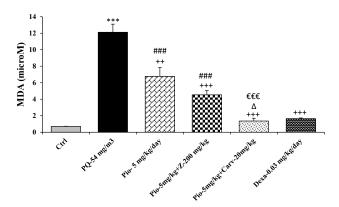


Figure 4. Serum level of malondialdehyde (MDA) in different studied groups. The results are expressed as mean±SEM (n=5 in each group). ****P*<0.001 compared to the control group. ++ *P*<0.01 and +++*P*<0.001 compared to the PQ group. ###*P*<0.001 compared to dexamethasone treated group. ΔP <0.05 compared to pioglitazone treated group. $\notin \in eP$ <0.001 compared to group treated with combination of pioglitazone and the extract. Comparisons between different groups were made using one-way ANOVA followed by Tukey's multiple comparison test. Ctrl, control group, PQ-54 mg/m³, group exposed to paraquat aerosol with dose of 54 mg/m³, Pio-5 mg/kg/day, Pio-5 + Z- 200 mg/kg/day, Pio-5 + Carv-20 mg/kg/day and Dex-0.03mg/kg/day, gioglitazone, combination or 200 *Zataria multiflora* and 20 mg/kg/day day and carvacrol respectively or dexamethasone respectively

and carvacrol on serum levels of all measured variables except IL-6 were higher than the treatment effects of combination with pioglitazone and *Z.multiflura* (P<0.05 to P<0.001), (Figures 2-3, 5 and 6).

The effects of dexamethasone

Dexamethasone treatment caused significant

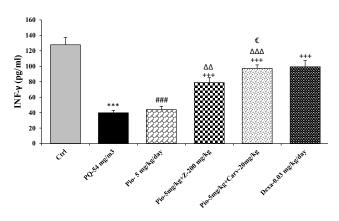


Figure 6. Serum level of Interferon gama (INF- γ) in different studied groups. The results are expressed as mean±SEM (n=5 in each group). ****P*<0.001 compared to the control group. +++*P*<0.001 compared to the PQ group. ###*P*<0.001 compared to dexamethasone treated group. $\Delta\Delta P$ <0.01 and $\Delta\Delta\Delta P$ <0.001 compared to pioglitazone treated group. €*P*<0.05 compared to treated group with combination of pioglitazone ant the extract. Comparisons between different groups were made using one-way ANOVA followed by Tukey's multiple comparison test. Ctrl, control group, PQ-54 mg/m³, group exposed to paraquat aerosol with dose of 54 mg/m³, Pio-5 mg/kg/day, Pio-5 mg/kg/day, Pio-5 + Z-200 mg/kg/day, Pio-5 + Carv-20 mg/kg/day and Dex-0.03mg/kg/ day, groups exposed to PQ-54 mg/m³ and treated with 5 mg/kg/day pioglitazone, combination or 200 *Zataria multiflora* and 20 mg/kg/ day and carvacrol respectively or dexamethasone respectively

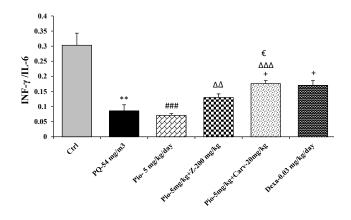


Figure 7. IFN-γ/IL-4 ratio in different studied groups. The results are expressed as mean±SEM (n=5 in each group). ***P*<0.01 compared to the control group. +*P*<0.05 compared to the PQ group. ###*P*<0.001 compared to dexamethasone treated group. ΔΔ *P*<0.01 and ΔΔΔ *P*<0.001 compared to pioglitazone treated group. € *P*<0.05 compared to treated group with combination of pioglitazone ant the extract. Comparisons between different groups were made using one-way ANOVA followed by Tukey's multiple comparison test. Ctrl, control group, PQ-54 mg/m³, group exposed to paraquat aerosol with dose of 54 mg/m3, Pio-5 mg/kg/day, Pio-5 + Z- 200 mg/kg/day, Pio-5 + Carv-20 mg/kg/day and Dex-0.03 mg/kg/day, groups exposed to PQ-54 mg/m³ and treated with 5 mg/kg/day and carvacrol respectively or dexamethasone respectively

improvement in all measured variables compared to PQ exposed group (P<0.05 for CAT and IFN- γ /IL-6 ratio, and P<0.01 for NO₂ and IL-6 and P<0.001 for other cases), (Figures 2-7). The effects of dexamethasone treatment on serum levels of CAT, MDA IFN- γ and IFN- γ /IL-6 ratio were higher than the effects of treatment with pioglitazone (P<0.01 for CAT and P<0.001 for other cases), (Figures 1, 3, 4, 6 and 7). The effect of treatment with dexamethasone on serum level of MDA was also higher than the effect of combination therapy of pioglitazone and *Z. multiflora extract* (P<0.001), (Figure 4). However, the effect of combination therapy of pioglitazone and carvacrol on serum level of NO₂ (P<0.001), (Figure 3).

Discussion

The results indicated that exposure to aerosols of PQ (54 mg/m³) in rat, increased serum level of NO_2 and MDA, but serum CAT activity was decreased. Serum level of IL-6 was also increased but IFN- γ level in the serum, and IFN- γ /IL-6 ratio were decreased in rats exposed to inhaled PQ.

Various experiments and clinical studies have shown, that free radicals generation play a crucial role of PQ-induced injuries and it was shown that the major mechanism of acute and chronic toxicity by paraquat occurred through oxidative stress (29). In addition, significant rise in lipid peroxidation level in the lung tissues was observed with PQ administration in the animals which was accompanied by decreases in SOD and CAT activities (30). Decreased levels of SOD, CAT and increased levels of MDA and NO₂ were showed due to PQ toxicity (31), Positive correlation was also

observed between the enhanced levels of oxidants and inflammatory mediators with PQ administration doses (32) which support the findings of the present study regarding the induction of oxidative stress induced by inhaled PQ.

It was also shown that increased inflammatory cytokines play an important role in paraquat poisoning (33). Increased white blood cell (WBC), neutrophil count (3), and decreased anti-inflammatory cytokines such as IL-10 and IFN- γ , demonstrate inflammatory process caused by paraquat toxicity (2). In addition previous studies demonstrated enhanced genes expression IL-4, TGF- β , IL-17, (34) and TNF- α after PQ challenge (35), which support the results of this study indicating systemic inflammation induced by inhaled PQ.

The current therapy of paraquat poisoning are not effective, which indicated a major need for novel therapies for disorders induced by PQ exposure. In the last decade, researchers have been focused on the use of anti-oxidant and anti-inflammatory compounds for treatment of PQ toxicity. Evidence showed that activation of PPAR-y receptors reduces inflammation and oxidative stress in various tissues, including the kidney, nervous system, and liver (36), Also it has been observed that the PPAR-y agonist pioglitazone improved anti-oxidant capacity, and increases SOD and CAT enzymes production in a kidney ischemia-reperfusion model (37). PPAR- γ agonists inhibit nitric oxid, TNF- α , IL-1 β , IL-6 levels, and MCP-1 from microglia and astrocytes (38). Decreased IL-6 and IL-8 in endometrial stromal cells via a PPAR-y-dependent mechanism by thiazolidinediones was also reported. Therefore there is a positive association between activation of this receptors and suppression of inflammatory factors (39).

In the present study, treatment with pioglitazone (5 mg/kg), PPAR- γ agonist only decrease serum level of MDA compared to PQ group and did not improved the other measured parameters in PQ exposed rats. These results could be due to inadequate dose of pioglitazone used in the present study. The results of a previous study also showed minor effect of treatment with 200 mg/kg/ day *Z.multiflura* extract in animals exposed to inhaled PQ (23). Treatment with 20 mg/kg/day carvacrol, also showed minor effect on animals exposed to inhaled PQ.

However, treatment with combination of pioglitazone and *Z. multiflura* extract improved serum levels of CAT, $NO_{2'}$ MDA and IL-6 and treatment with combination of pioglitazone and carvacrol improved all measured variables compared to PQ exposed group. In addition, the effects of combination therapy of pioglitazone and *Z.multiflura* extract as well as the effect of pioglitazone and carvacrol were significantly higher than pioglitazone alone treated group. Therefore combination therapy of PPAR- γ agonist with *Z.multiflura* and specially its constituent, carvacrol improved oxidative stress, cytokines and increased IFN- γ /IL-6 ratio or Th1/Th2 balance in animals exposed to inhaled PQ. These results may indicate a synergism effect of the *Z.multiflura* and its constituent, carvacrol with the effect pioglitazone.

Anti-oxidant and anti-inflammatory properties of *Z. multiflora* extract and its constituent carvacrol in animal and human studies were reported (18). The extract and carvacrol showed anti-oxidat and anti-inflammatory effects in a guinea pig model of asthma and COPD

(40). In clinical studies also, *Z. multiflora* extract and carvacrol reduced MDA level as an oxidant marker and increased anti-oxidant levels including thiol, SOD and CAT (41). The plant and its constituent also showed anti-inflammatory effect in animal model of asthma (22). as well as in clinical studies (26, 41, 42). The effect of extract of *Z. multiflora* and carvacrol on enhanced IL-4/IFN- γ ratio as an indicator of Th1/Th2 balance were also shown previously which support the finding of the current study (34, 43, 44).

Therefore, the results may suggest that the effects of the plant and carvacrol could be mediated by their effects on PPAR- γ receptors. However, further studies, examining PPAR- γ antagonists needed to confirm this suggestion. In addition, the effects of treatment with combination of pioglitazone and carvacrol on all measured variables except IL-6 were higher than the treatment effects of combination with pioglitazone and *Z.multiflura* which suggest that the effect of the plant is due to its constituent, carvacrol.

In fact, Hotta *et al.* showed suppressing effect of carvacrol on COX-2 and activation of PPAR (45) which support the findings of the present study regarding the effect of *Z.multiflura* and carvacrol on PPAR- γ receptors.

Farmers are usually exposing to PQ through inhalation. Therefore, the aims of the present study are exploring the potential effects of PPARy agonist alone and in combination with Z. multiflora hydro alcoholic extract and carvacrol on the PQ inhalation induced systemic toxicity in the male Wistar rat by examining oxidant, anti-oxidant markers and cytokine levels in the serum. The results showed significant changes in serum levels of all oxidant, anti-oxidant and cytokine indicating the induction of systemic oxidative stress and inflammation by inhaled PQ which are novel findings. However, the induction of lung inflammation and oxidative stress by inhaled PQ was shown in our previous study (23). Our previous study showed the minimal effect of 200 mg/ kg Z. multiflura extract and 20 mg/kg carvacrol but significant effects of their 800 and 200 mg/kg on rats exposed to paraquat respectively (23). In the present study the low dose of the extract and carvacrol was used to evaluate their synergism effect with pioglithasone.

The results of the present study also showed similar effect of dexamethasone a known anti-inflammatory drug with the effects of the combination of pioglitazone with the extract and carvacrol on oxidative stress and inflammation process induced by inhaled PQ which is a further evidence for the anti-inflammatory and antioxidant effect of the plant and its constituent carvacrol.

Conclusion

Combination therapy of pioglitazone with *Z.* multiflora extract or carvacrol reduced NO_2 and MDA but increased CAT levels as well as reduced IL-6 but increased IFN- γ and Th1/Th2 balance in rats exposure to PQ, similar is to of the effect of dexamethasone. However, pioglitazone alone in the present study and the extract and carvacrol in previous study at studied doses showed minimum effects on changes induced by PQ aerosol. These results suggest that the effects of *Z.* multiflora extract or carvacrol are mediated by their effect on PPAR- γ receptor.

Financial Disclosure

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Conflicts of Interest

The authors declare that there are no conflicts of interest.

References

1. Dhananjayan V, Ravichandran B. Occupational health risk of farmers exposed to pesticides in agricultural activities. Curr Opin Environ Sci Health 2018;4:31-37.

2. Hu X, Liang Y, Zhao H, Zhao M. Effects of AT-RvD1 on paraquatinduced acute renal injury in mice. Int Immunopharmacol 2019;67:231-238.

3. Dinis-Oliveira R, Duarte J, Sanchez-Navarro A, Remiao F, Bastos M, Carvalho F. Paraquat poisonings: mechanisms of lung toxicity, clinical features, and treatment. Crit Rev Toxicol 2008;38:13-71.

4. Buendía JA, Chavarriaga GJR, Zuluaga AF. Burden of paraquat poisoning in the department of Antioquia, Colombia. BMC Pharmacol Toxicol 2019; 20:11.

5. Asadi R, Afshari R. Ten-year disease burden of acute poisonings in Northeast Iran and estimations for national rates. Hum Exp Toxicol 2016; 35:747-59.

6. Kim Sj, Gil HW, Yang JO, Lee EY, Hong SY. The clinical features of acute kidney injury in patients with acute paraquat intoxication. Nephrol Dial Transplant 2008;24:1226-1232.

7. Yoon SC. Clinical outcome of paraquat poisoning. Korean J Intern Med 2009;24:93-94.

8. Gawarammana IB, Buckley NA. Medical management of paraquat ingestion. Br J Clin Pharmacol 2011;72:745-757.

9. Zhao G, Li S, Hong G, Li M, Wu B, Qiu Q, *et al.* The effect of resveratrol on paraquat-induced acute lung injury in mice and its mechanism. Zhongguo Wei Zhong Bing Ji Jiu Yi Xue 2016;28:33-37.

10. Sun L, Yan PB, Zhang Y, Wei LQ, Li GQ. Effect of activated charcoal hemoperfusion on renal function in patients with paraquat poisoning. Exp Ther Med 2018;15:2688-2692.

11. Gawarammana IB, Buckley NA. Medical management of paraquat ingestion. British J Clin Pharmacol 2011;72:745-757. 12. Tyagi S, Gupta P, Saini AS, Kaushal C, Sharma S. The peroxisome proliferator-activated receptor: A family of nuclear receptors role in various diseases. J Adv Pharm Technol Res 2011;2:236-240.

13. Youssef J, Badr MZ. PPARs: history and advances, Peroxisome Proliferator-Activated Receptors (PPARs). Springer 2013; 1-6.

14. Wang L, Waltenberger B, Pferschy-Wenzig E-M, Blunder M, Liu X, Malainer C, *et al.* Natural product agonists of peroxisome proliferator-activated receptor gamma (PPARγ): a review. Biochem Pharmacol 2014;92:73-89.

15. Fujiki Y, Okumoto K, Kinoshita N, Ghaedi K. Lessons from peroxisome-deficient Chinese hamster ovary (CHO) cell mutants. Biochim Biophys Acta 2006;1763:1374-1381.

16. Ohtera A, Miyamae Y, Yoshida K, Maejima K, Akita T, Kakizuka A, *et al*. Identification of a new type of covalent PPARγ agonist using a ligand-linking strategy. ACS Chem Biol 2015;10:2794-2804.

17. Sajed H, Sahebkar A, Iranshahi M. *Zataria multiflora* Boiss.(Shirazi thyme)—an ancient condiment with modern pharmaceutical uses. J Ethnopharmacol 2013;145:686-698.

18. Khazdair MR, Ghorani V, Alavinezhad A, Boskabady MH. Pharmacological effects of *Zataria multiflora* Boiss L. and its constituents focus on their anti-inflammatory, anti-oxidant, and immunomodulatory effects. Fundam Clin Pharmacol 2018;32:26-50.

19. Ahmadipour A, Sharififar F, Nakhaipour F, Samanian M, Karami-Mohajeri S. Hepatoprotective effect of *Zataria multiflora* Boisson cisplatin-induced oxidative stress in male rat. IJMS 2015;8:275.

20. Burleigh-Flayer H, Alarie Y. Concentration-dependent respiratory response of guinea pigs to paraquat aerosol. Arch Toxicol 1987;59:391-396.

21. Malekinejad H, Khoramjouy M, Hobbenaghi R, Amniattalab A. Atorvastatin attenuates the paraquat-induced pulmonary inflammation via PPARγ receptors: A new indication for atorvastatin. Pestic Biochem Phys 2014;114:79-89.

22. Jalali S, Boskabady MH, Rohani AH, Eidi A. The effect of carvacrol on serum cytokines and endothelin levels of ovalbumin sensitized guinea-pigs. Iran J Basic Med Sci 2013;16:615-619.

23. Heydari M, Mokhtari-Zaer A, Amin F, Memarzia A, Saadat S, Hosseini M, *et al*. The effect of *Zataria multiflora* hydroalcoholic extract on memory and lung changes induced by rats that inhaled paraquat. Nutr Neurosci 2019:1-14.

24. Alemán-Laporte J, Bandini LA, Garcia-Gomes MS, Zanatto DA, Fantoni DT, Amador Pereira MA, *et al.* Combination of ketamine and xylazine with opioids and acepromazine in rats: Physiological changes and their analgesic effect analysed by ultrasonic vocalization, Laboratory animals. 2019:0023677219850211.

25. Hakimizadeh E, Askary A, Shamsizadeh A, Rahmani M, Vazirinejad R, Ayoobi F, *et al.* Effect of hydro-alcoholic extract of Artemisia aucheri on castor oil-induced diarrhea in male rat. J Shahrekord Univ Med Sci 2013;15-22.

26. Khazdair MR, Ghorani V, Alavinezhad A, Boskabady MH. Effect of *Zataria multiflora* on serum cytokine levels and pulmonary function tests in sulfur mustard-induced lung disorders: A randomized double-blind clinical trial. J Ethnopharmacol 2020;248:112325-112333.

27. Shakeri F, Soukhtanloo M, Boskabady MH. The effect of hydro-ethanolic extract of Curcuma longa rhizome and curcumin on total and differential WBC and serum oxidant, anti-oxidant biomarkers in rat model of asthma. Iran J Basic Med Sci 2017;20:155-165.

28. Saadat S, Beheshti F, Askari VR, Hosseini M, Roshan NM, Boskabady MH. Aminoguanidine affects systemic and lung inflammation induced by lipopolysaccharide in rats. Respir Res 2019;20:96.

29. Facecchia K, Fochesato LA, Ray SD, Stohs SJ, Pandey S. Oxidative toxicity in neurodegenerative diseases: role of mitochondrial dysfunction and therapeutic strategies. J Toxicol 2011;2011:683728-683739.

30. Cheresh P, Kim S-J, Tulasiram S, Kamp DW. Oxidative stress and pulmonary fibrosis. Biochim Biophys Acta 2013;1832:1028-1040.

31. Keeling PL, Smith LL. Relevance of NADPH depletion and mixed disulphide formation in rat lung to the mechanism of cell damage following paraquat administration. Biochem Pharmacol 1982;31:3243-3249.

32. Suntres ZE. Exploring the potential benefit of natural product extracts in paraquat toxicity. Fitoterapia 2018; 131: 160-167.

33. Meng Z, Dong Y, Gao H, Yao D, Gong Y, Meng Q, *et al.* The effects of ω -3 fish oil emulsion-based parenteral nutrition plus combination treatment for acute paraquat poisoning. Glob J Med Res 2019;47:600-614.

34. Kianmehr M, Rezaei A, Hosseini M, Khazdair MR, Rezaee R, Askari VR, *et al.* Immunomodulatory effect of characterized extract of *Zataria multiflora* on Th1, Th2 and Th17 in normal and Th2 polarization state. Food Chem Toxicol 2017;99:119-127.

35. Piguet PF, Collart MA, Grau GE, Sappino AP, Vassalli P. Requirement of tumour necrosis factor for development of silica-induced pulmonary fibrosis. Nature 1990;344:245.

36. Zou C, Hu H, Xi X, Shi Z, Wang G, Huang X. Pioglitazone protects against renal ischemia-reperfusion injury by enhancing anti-oxidant capacity. J Surg Res 2013;184:1092-1095.

37. Al Rouq F, El Eter E. PPAR-γ activator induces neuroprotection in hypercholesterolemic rats subjected to global cerebral ischemia/reperfusion injury: *In vivo* and *in vitro* inhibition of oxidative stress. Exp Gerontol 2014;51:1-7.

38. Storer PD, Xu J, Chavis J, Drew PD. Peroxisome proliferatoractivated receptor-gamma agonists inhibit the activation of microglia and astrocytes: implications for multiple sclerosis. J Neuroimmunol 2005;161:113-122.

39. McKinnon B, Bersinger NA, Mueller MD. Peroxisome proliferating activating receptor gamma–independent attenuation of interleukin 6 and interleukin 8 secretion from primary endometrial stromal cells by thiazolidinediones. Fertil Steril 2012;97:657-664.

40. Khazdair M, Alavinezhad A, Boskabady M. Carvacrol ameliorates haematological parameters, oxidant/anti-oxidant biomarkers and pulmonary function tests in patients with sulphur mustard-induced lung disorders: A randomized double-blind clinical trial. J Clin Pharm Ther 2018;43:664-674. 41. Khazdair MH, Rajabi O, Balali-Moodd M, Beheshtie F, Boskabady MH. The effect of *Zataria multiflora* on pulmonary function tests, hematological and oxidant/anti-oxidant parameters in sulfur mustard exposed veterans, a randomized double-blind clinical trial. Environ Toxicol Pharmacol 2018;58:180-188.

42. Khazdair MR, Boskabady MH. The effect of carvacrol on inflammatory mediators and respiratory symptoms in veterans exposed to sulfur mustard, a randomized, placebocontrolled trial. Respir Med 2019;150:21-29.

43. Kianmehr M, Rezaei A, Boskabady MH. Effect of carvacrol on various cytokines genes expression in splenocytes of asthmatic mice. Iran J Basic Med Sci 2016; 19: 402-410.

44. Boskabady MH, Mehrjardi SS, Rezaee A, Rafatpanah H, Jalali S. The impact of *Zataria multiflora* Boiss extract on *in vitro* and *in vivo* Th1/Th2 cytokine (IFN- γ /IL4) balance. J Ethnopharmacol 2013; 150: 1024-1031.

45. Hotta M1, Nakata R, Katsukawa M, Hori K, Takahashi S, Inoue H. Carvacrol, a component of thyme oil, activates PPARalpha and gamma and suppresses COX-2 expression. J Lipid Res 2010;51:132-139.