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Efficacy of insulin targeted gene therapy for type 1 diabetes mellitus: A systematic review and meta-analysis of rodent studies

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ABSTRACT

Diabetes mellitus (DM) is a major worldwide public health challenge, for which gene therapy offers a potential therapeutic approach. To date, no systematic review or meta-analysis has been published in this area, so we examined all relevant published studies on rodents to elucidate the overall effects of gene therapy on bodyweight, intraperitoneal glucose tolerance test (IPGTT), fasting blood glucose, and insulin in animals with type 1 DM. The Cochrane Library, PubMed, Embase, ISI Web of Science, SCOPUS, and Google Scholar were systematically searched for potentially relevant studies. Mean \pm standard deviation (SD) was pooled using a random-effects model. After the primary search, out of 528 studies identified, 16 studies were in concordance with predefined criteria and selected for the final assessment. Of these, 12 studies used viral manipulation, and 4 employed non-viral vectors for gene delivery. The meta-analysis showed gene therapy with a viral vector decreased mean IPGTT (-12.69 mmol/l, P<0.001), fasting blood glucose (-13.51 mmol/l, P<0.001), insulin (398.28 pmol/l, P<0.001), and bodyweight (24.22 g, P<0.001), whereas non-viral vectors reduced fasting glucose (-29.95 mmol/l, P<0.001) and elevated insulin (114.92 pmol/l, P<0.001). Gene therapy has favorable effects on alleviating type 1 DM related factors in diabetic rodents.

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Introduction

Diabetes mellitus (DM) is one of the most important public health challenges worldwide (1,2), of which type 1 diabetes mellitus (T1DM) (3,4) is characterized by hyperglycemia caused by autoimmune destruction of pancreatic β -cells, the main site of insulin production and secretion (4). The prevalence of diabetes was estimated to be 422 million in 2014 (1) and expected to reach 522 million in 2030 (5). The total cost of diabetes and pre-diabetes in the US is \$322 billion per annum (6), and health care costs for people with diabetes are 2.3 times higher than those without diabetes (7). Most of the global burden of this disorder is due to morbidity and mortality that arises from complications of the disease (1,8-13).

The most commonly utilized treatment for T1DM is insulin infusion, which requires close monitoring of blood glucose during insulin therapy, which is then needed daily and reduces patient compliance (14). At the same time, the optimal blood glucose is rarely achieved and patients remain at risk from experiencing regular periods of hypo or hyperglycemia (15, 16). This type of adverse blood glucose places the patient at risk

of hypoglycemic coma or hyperglycemia complications, such as retinopathy, nephropathy, neuropathy, and cardiovascular disease (16, 17). Consequently, a treatment option that is more able to maintain normoglycaemia without adverse complications and greater compliance remains highly desirable (18, 19). Insulin gene therapy is one alternative and represents a novel therapeutic approach to achieving regulated insulin production and delivery (20). Recently, numerous studies have reported the effects of insulin gene therapy on T1DM, which showed some beneficial outcomes in some (21, 22), but not all cases (23). The aim of this paper was to conduct a systematic review and meta-analysis to estimate the effects of insulin gene therapy on T1DM related factors, including bodyweight, intraperitoneal glucose tolerance test (IPGTT), fasting blood glucose, and insulin in diabetic rodents.

Materials and Methods

Search strategy

A comprehensive search was conducted in medical databases including Cochrane reviews, Medline/PubMed, EMBASE, ISI Web of Science, SCOPUS, and



Table 1. Gene therapy for type 1 diabetes mellitus using non-viral vectors

Reference No.	Country	Study design	Target tissue	Model	Gene delivery method	Route of delivery	Sample size					Result					Adjusted variable												
24	Malaysia	sia Case	k-cell & l	Male CD		Orally	Treat group=	Fasting Blood glucose P<0.05 mmol/l mean± SD																					
		control	cell	1 mice	GIP/hins/pBu d		10	days	1	3	5	7	9	11	13	15	 Age and sex adjusted 												
			intestine		а	the nanoparticles	Control diabetic= 10	Treat group		24.3±	21.09±		18.99±1.3	18.85±		19.23±0.97	- adjusted												
						every 2 days	Control normal= 10	Control DM	1.52 25.27±	1.55 26.19± 0.76	3.2 25.26±0.4 2	1.32 4 26.06± 0.58	26.51±0.4 5	1.09 25.45± 0.57	2.17 26.54±0.	6 26.82±0.48													
								Control normal	0.75 6.36±0.69						5.99±0.8	7 5.77±0.98													
										Н	uman inst	ılin pmol/l	mean± SD				_												
									eat group wit group with G					18.217 17.916			=												
									Control	DM				1.048															
25	China	Case control	k-cell intestine	Male wister	chitosan pCMV/ hIns	coloclysis	Treat group = 10 Control diabetic= 10				,	vels mmol/l	,	mean±			Age and sex — adjusted												
		control	incestine	rats	plasmid			Days	0	1		2	3	4		5													
								Treat group				7.1±0.7	6.0± 0.25	5.07±		8.6± 0.34													
								Diabete control chitosan	22.7±1.5 23.5± 1.5	23± 15.7:		23.5±1.9 14± 1.0	22.4±2.3 12.3± 1.2	26±		25.7±2.0 13.9±0.8													
								Cintosan	23.3± 1.3	13.7.	. 1.1	141 1.0	12.3± 1.2	12.5	±1.0	13.910.0													
									1	Plasma ins	ulin level	s pmol/l (P<0.01) mean± 5	ean± SD															
							Days 0	1		2	3		1	5	-														
								0 1				206.2± 9	222.9± 9.7			208.3± 6.2													
								control				97.2± 6.2 52.8± 7.6	92.3± 6.2 159.7± 6.9			84.7± 5.55 156.2± 7.6													
26	China	Case control	k-cell	Male	chitosan	coloclysis	Treat group = 10 Control diabetic= 10									130.21 7.0	Age and sex												
			intestine	wistar rats	pCMV/ hIns			Days	0	1	2	3	4	5	6	7	- adjusted												
					plasmid			Treat group	21.6± 1.0	11.1±		-																	
								Diabete control	21.6± 1.0	22± 1.	31 22.5±	±1.0 25± 1.2	2 23.2± 1.2	2 26±	1.0 26.25	±1.2 27.1± 1.0													
																					chitosan	21.6± 1.0 Pl	15± 1 lasma insu		1.0 21.7±0. pmol/l (/			0.9 15.25	±1.2 15.7± 0.7
								Days	0	1	2	3	4	5	6	7	•												
												Treat group	98.8± 9.5			±1 224.3±1	1 227.1±12. 7		-	±9 194.4±8.3									
								Diabetic control	98.8± 9.5	97.2 ± 7	.63 104.1	±9 93±7.5	97.9± 8.3	88.2±	8.3 83.3±	10. 84.7± 8.3	:												
								chitosan	98.8± 9.5	149.3±	8.3 156.2 6	±7. 152.7±7	'. 149.3±8.3	3 159.7±		±7. 180.5±9													
27	USA	Case		Mice	Linearized	Fertilized	Treat group =									Age and sex													
		control	intestine		GIP/Ins Fragment	mouse embryos	10 Control	Treat group 39 ± 16.9 pmol/l								adjusted													
							diabetic= 5	Control DM 0								•													
27	USA	Case control	k-cell intestine	Mice	Linearized GIP/Ins	Fertilized mouse	Treat group = 5 Control		Fasting blood glucose mmol/l mean±SD Treat group 9.52 ± 1.16								_ Age and sex adjusted												
					Fragment	embryos	diabetic= 3		Control	DM				26 ± 2.6	63														

Google Scholar up to July 2019 using the following medical subject headings (MeSH) and non-MESH keywords relevant to ((" Gene Therapies "[tiab] OR " DNA Therapy "[tiab] OR " Genetic Therapies "[tiab])) AND (("Genetic Vectors")) AND (("diabetes mellitus"[tiab])) AND (("insulin"[tiab])) regardless of language. The reference lists of related articles were then hand-searched for additional relevant studies. Titles/abstracts were screened for relevant studies by two independent investigators.

Study selection

Inclusion and exclusion criteria

Studies were eligible for inclusion in the current analysis if: (i) insulin gene therapy was assessed in animal studies; (ii) their final outcome was diabetes-related factors; and (iii) mean, standard error (SE), or standard deviations (SD) for the mentioned factors were provided. Studies were excluded if they did not include outcome measurements for diabetic control groups or only reported the mean average outcome during the treatment. Two investigators extracted data independently, and any discrepancies were resolved by discussion.

Outcomes

Studies evaluating insulin gene therapy effect on

diabetes-related factors were included in the current study, and the outcomes of interest were IPGTT, fasting blood glucose, insulin, and bodyweight.

Data extraction

The data included the first author, year of publication, country where the study was conducted, sample size, gene delivery route, gene delivery method, target tissue, follow-up duration, main outcome, covariates adjusted for in the analysis and mean and SD or SE. Characteristics of each study on insulin gene therapy by viral and non-viral vector are summarized in Tables 1 and 2, respectively.

Statistical methods

Means after treatment and their SD or SE were collated as the measurable effect of insulin gene therapy on diabetes-related factors. Meta-analysis was performed using the random-effects model and presented as forest plots. Evidence for publication bias was sought by performing Egger's test in addition to visual inspection of the funnel plots. The percentage of variability across the pooled estimates attributable to heterogeneity beyond chance was estimated with the I^2 index, and the P-value for heterogeneity ($I^2 > 50\%$ was considered as significant heterogeneity). In the case of significant between-study heterogeneity, sensitivity analyses were



 $\textbf{Table 2.} \ \textbf{Gene therapy for type 1 diabetes mellitus using viral vectors}$

Reference No.	Country	Study design	Target tissue	Model	Gene delivery method	Route of delivery	Sample size	OR,RR						Adjusted variable			
28	USA	case control	liver	Male CD-1 mice	GIRE)3BP1- 2xfurwith AAV8 capsids	Portal.vein (in liver)	Treat group= 7 Control diabetic= 5 Control normal=	time (60	90 10.72±4.10	120	150 24 8.6± 2.94 6.66	40 300 5±2.6 5±1.32	Age and sex adjusted		
					capsias		5	Control DM 20.1	3 ±5.3 25.1±2.24				! 22.2±1.12 21.1	5 .±2.4 18.3±3.13 6			
22	Germany	case control	liver	rat	CMV/humar furin ins- lentivirus	n portal vein	Treat group= 5 Control DM= 6	Time min Treat group	0 4± 1.2	IPGTT	mmol/l mean 15 30±2	±SD 60 27±8.94	120 23±5.59	180 18±4.47	Age and sex adjusted		
21	Australia	case	liver	mice	HMD/INS-	nortal vein	Treat group= 6	Control DM	19± 7.35		33± 4 nmol/l mea	31±7.35	30±4.90	27±4.90	Age and sex		
		control			FUR lentiviral vector		Control DM= 6 Control normal= 5	Time min Treat Control DM	0 7.8±2.20 20.4±2.3	3 23.5± 4.16 30± 7.84	5 22.7±6.37 28± 8.33	10 16.1±2.20 28.1±7.10		30 7.5±1.71 23.9±5.63	- adjusted		
29	Australia	case control	liver	rat	HMD/INS- FUR lentiviral vector	(IVC) hepation	Treat group= 5 Control DM= 5 Control normal=	Time min	0 7.5± 2.24	IPGTT n 20 16±6.71	30 16±8.94	60 14±6.71	90 11±4.47	120 8±1.12	Age and sex adjusted		
								Control normal	7± 2.24	14±4.47	15±2.24	12±2.24	10±1.12	7±0.67	_		
30	Taiwan	case control	liver	Male C57BL/6J mice		into the liver by a single administrati on	Treat group= 3 Control DM= 3	Time min Treat group Control DM	0 7.49± 0.9	30 16.2 ± 1.4	mmol/l mear 60 19.5± 2.7	90 14.4± 2.2	120 11.1± 1.1 erimental perio		_ Age and sex adjusted		
31	Korea	case control	liver	SpragueD	rAd- SP23137-	tail vein	Treat group= 5 Control DM= 7		Bloo	d glucose 24 l	nr fasting mn		SD		Age and sex		
31		control		awiey rats	rINSfur		CONTROL DIVI- 7	Treat gi Control Treat gi	DM) : 1.5 I ± 2	24 hr fastin 5±0.4 20±0.6	g	aujusteu			
28	USA	case control	liver	Male CD-1 mice	GIRE)3BP1- 2xfurwith AAV8 capsids	portal vein (in liver)	Treat group= 7 Control diabetic= 5 Control normal= 5		ood glucose (P	Befor	average cumu e treat ± 2.19	lative) mea	an± SD mmol/l after treat 2.7 ± 0.79		Age and sex adjusted		
								Control n			± 2.2		6.77 ± 1.05 4± 0.4	;			
32	USA	case control	liver	rat	retroviral vectors pLX/rINS	portal vein	Treat group= 9 Diabeti control = 8	time Treat gi	e	ood glucose le)	mmol/l 24 hr 5± 0.6		Age and sex adjusted			
33	Germany	case	liver	rat	HMD/INS		Treat group= 6	Control	DM	14.9 lucose mean a		nean± SD m	16.7 ± 3		Age and sex		
		control			FUR lentiviral vector	(in liver)	Diabeti control = 6	Treat g	-		before 22.3±3.4 22.3±3.4		aft 15.6 22.3	± 2.1	- adjusted		
31	Korea	case control	liver	SpragueD awley rats	rAd- SP23137- rINSfur	tail vein	Treat group= 5 Control DM= 7	Treat go	roup	d glucose 24 hr fasting mmol/l mean± 0 7.8± 1.5			SD 24h fasting 5±0.4	Age and sex adjusted			
								Treat gr	roup	26.3	3 ± 2		20±0.6		_		
34	Korea	case control	liver	rat	CMV-Ins lentiviral vector		Treat group= 5 Control DM= 5	Fa	Treat group Control DM						Age and sex adjusted		
28	USA	case control	liver l	Male CD-1 mice	GIRE)3BP1 2xfurwith AAV8 capsids	(in liver)	Treat group= 7 Control diabetic= 5 Control normal=	before				P<0.001) m	mol/l mean±SD aft 5.88	Age and sex adjusted			
								Control DM		20.2± 2.2			18 ±				
			17		HMB (P)			Control normal			6.7± 0.16		6.33	- Age 1			
29	Australia	case control	Liver	Rat	FUR lentiviral vector	(IVC) hepation	C Treat group= 5 Control DM= 5 Control normal= 5		0	3 days	se mmol/l r 25 da	iys	50 days	60 days	Age and sex adjusted		
					vector		3	Treat group Control DM	7.3± 2.20 7.4± 2.20	7.5 ±3.13 28± 11	6.3± 4 28.5 ±		6.5±2.23 27±11	6.7 ± 0.67 27.4 ± 4.0			
21	Australia	case control	liver	mice	HMD/INS- FUR lentiviral vector	portal vein	Treat group= 6 Control DM= 6 Control normal=	Time day		0 14.1±7.3	ose mmol/l m	5 7±2.44	10 8 ±2.44	15 7±2.55	Age and sex adjusted		
								Control DM		14.2± 7.3		22 ±4.4	26±5.38	28 ±9.79	=,		
22	Germany	case control	liver	rat	CMV/human furin ins- lentivirus	n portal vein	Treat group= 5 Control DM= 6	Time days Treat group			glucose mmo 5 12.5	l/l		10 12 23	Age and sex adjusted		
33	Germany	case control	liver	rat	HMD/INS- FUR lentiviral		Treat group= 6 Diabeti control =		Blood g	glucose mean after 30 days mmol/l mea Before			n ± SD After	Age and sex adjusted			
					vector			Treat group Control DM		22.3±3.4 22.3±3.4			15± 2 21.9 ±	_			
34	Korea	case control	liver	rat	CMV-Ins lentiviral vector		Treat group= 5 Control DM= 5	Time Treat group	p	Blood gluce Before vi 23.4 ±		ean± SD	10 week after v		Age and sex adjusted		
								Control DM	1	23.4	2.55		28				



Continued Table 2

21	Australia	case		mice		portal vein	Treat group= 6		Insu	in pmol/l after	glucose admi	nistration me	an± SD		Age and sex					
		control			FUR lentiviral		Control DM= 6 Control normal=	Time min) 3	5	10	15	30	40 40	adjusted					
					vector		5		12.2 1800±	17 1570 ±14.	6 870±12.	810±12.2	730± 12.2	570±12. 570±12.2						
											2			2						
								Control DM			Not de	etected								
								Control 420± norm	11.1 1800±	15.6 1572± 13	4 720±11. 1	660± 11.1	662± 8.94	550±11. 550±11.1						
30	Taiwan	case	liver	Male	rIns I		Treat group= 3		Human inst	ılin pmol/l (A	fter glucose a	ıdministratio	n) mean±S	SD .	Age and sex					
		control				by a single administrati	Control DM = 3	Time min	0	30	60	90	120	150	adjusted					
					ins rAAV	on		Treat group	31.2 ± 3.4	84.72± 12.		95.14±5.8 8	77.07 ±12.1	61.8 ± 5.5						
								Control DM			Not d	etected (0)								
29	Australia	case	liver	rat			Treat group= 5		Hun	nan Insulin afte	Ü		,		Age and sex					
		control			FUR lentiviral	artery	Control DM= 5 Control normal=	Time min	0	20 30		90		120	adjusted					
					vector		5	Treat group	50	325 225		57		50						
								Control normal	75	350 250		85	5	80						
								Control DM				ot detect								
35	USA	case control	liver	Mouse	CMV/ human ins	Intra peritoneal	-				lin after 3 m	onth pmol/l	2		Age and sex adjusted					
					Lentiviral	injection		time		0 Not detected (0)		3 month			uujusteu					
					vector			Treat group					88.89 Not detected (0)							
								Control DM		Not detect			Not detec	eted (0)	_					
36	Italy	case control	liver	Mouse	L-PKp/ human furir		Treat group= 4 Control DM = 4		Treat group	Human insulin after 3 week pmol/l 896-1333					Age and sex adjusted					
		control	niu oi		ins (HD-	•			Control DM					not detected						
					AdV)			C	ontrol norm	al			342.4 ±55							
34	Korea	case		er rat	CMV-Ins		Treat group= 5	Ir	sulin conce	ntration 3 week	k after virus	injection (P=	0.077) mean	n± SD	Age and sex					
		control			lentiviral vector	arly inject	Control DM= 5	Treat group 18.75 ± 10 pmol/l						 adjusted 						
									Control DM			11.11	1 ± 7.6 pmol	1/1						
31	Korea	case	liver	SpragueD	rAd-		Treat group= 5 Control DM= 7			Serum rat ins	ulin level nm	ol/I mean+S	D .	-	Age and sex					
31		control	ontrol	awley rats	SP23137-			Time minut		0	unn iever pin	30'		120	- adjusted					
					rINSfur			Time minu	ie	70±1		92±5		75±1						
								treat												
21					HMD (INC		m	Control norr	nal	101± 3		100± 12		223 ± 10						
21	Australia	case control		mice	HMD/INS- FUR	portal vein	Treat group= 6	days	0	Body w	eight gram n	nean± SD 15	90	120 150	Age and sex adjusted					
					lentiviral		Control DM= 6 Control normal= 5		22.7±3.9	23±1.8	24.5±5.3	24.8±1.4	27.6±0.7		aujusteu					
					vector								27.0±0.7	27.0 11.0 2010.9						
								Control	22.5± 3.86	21.2± 3.8	22± 9.7	21.2±9.2	-							
28	USA	case control		liver				Treat group= 7	-	Body weight gr after 24 hr mean±SD						Age and sex				
					mice	2xfurwith AAV8	(in liver)	Control diabetic= 5	Time	I	Before			ter 24 hr		adjusted				
					capsids		diabetic= 5	Treat group	10	0± 10.5			90±7.9							
								Control DM	1	00± 6.7			85±4.8							
32	USA	case	liver	rat	retroviral	portal vein	Treat group= 16			Body weigh	t gr after 3 da	ys mean±SD			Age and sex					
		control			vectors pLX/rINS	-	Diabeti control = 18	Time	Days		Before	-	After t	reat 3 days	adjusted					
							18	Treat g	roup		173± 10		171	gr± 10.1						
								Contro	ol DM		175±9		13	5 gr± 9						
								Control	normal		177		1	180 gr						
29	Australia	case	ise liver	rat	HMD/INS-	(IVC) hepati	c Treat group= 5			Body we	ight: gram	mean± SD			Adjusted for					
		control			FÚR	artery	Control DM= 5	Time	0	5 days	25 days	50	days	60 days	age and sex					
					lentiviral vector		Control normal= 5	Treat group	300±11	-	389±8.9		±11.18	450±8.94						
								Control DM	292± 13.4	1 332±10	365±17.88	370)±9.39	360±8.94						
34	Korea	case	liver	rat	CMV-Ins		Treat group= 5	-		Body we	eight gram	mean±SD		Ag						
		control			lentiviral vector		Control DM= 5	Time		0		5 days		10 week	adjusted					
					vector			Treat group		121±8.9		115 ± 11 g		135± 33 g						
								Control DM		122± 6.7		116 ± 3.3 g		93± 3.51 g						

performed excluding individual studies to obtain an understanding of the reasons for any differences. Also, where there was a high likelihood of differences beyond chance, subgroup analysis, based on the gene delivery method and follow-up duration, was performed.

Publication bias was assessed statistically by Begg's test. *P*<0.05 was considered statistically significant. Statistical analyses were conducted using the statistical software package Stata (ver. 11.2).

Results

Search results and characteristics of included studies

The literature search on the subject of gene therapy and diabetes-related factors yielded 654 articles, of which 33 were reviewed as full texts. Of these, 16 studies met the inclusion criteria. The flow diagram summarizes the results of the study selection process for this meta-analysis (Figure 1). Out of 16 studies published 5 (23,27-28, 32, 35) were conducted in the USA, and 2 (22, 23), 2 (21, 29), 2 (31, 34), 2 (25, 26), 1 (36), 1 (30), 1(24) studies were published respectively

in Germany, Australia, Korea, China, Italy, Taiwan, and Malaysia. Six (21, 22, 28-31), 13 (21-29, 31-34), 5 (21, 28, 29, 32, 34), and 11 (21, 23-26, 29-31, 34-36) studies assessed the effects of insulin gene therapy on IPGGT, FBS, bodyweight, and blood insulin, respectively. The sample size ranged from 3 animals to 18 with follow-up ranging from 2 min to 3 months. Target tissue was based on the liver in all of the viral vector studies and K-cell in non-viral vector studies. The gene delivery method was a viral vector in 11 studies (21, 22, 28, 32, 33, 36), which used the portal vein as the route of delivery. Others (29-31, 34, 35) used the hepatic artery and tail vein. Four studies used a non-viral vector as a gene delivery method, of which 2 (25, 26) used coloclysis as the route of delivery. The remaining used an oral route (24) or the pronuclei of fertilized mouse embryos (27).

Findings from the systematic review:

Some studies that were initially included were subsequently excluded and were reported in a systematic study.

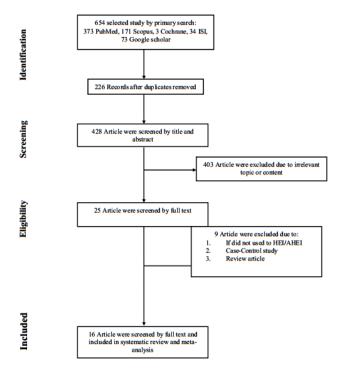


Figure 1. The flow diagram of study selection

Hsu *et al.* reported effect of insulin gene therapy on IPGTT; the IPGTT difference before glucose administration and 150 min after glucose administration in treat group was 1.21 mmol/l but in control diabetic group it was 10.9 mmol/l throughout the experimental period. Therefore, glucose was significantly decreased in the treat group. Similarly, the insulin level in the treat group was significantly increased (30.6±2.1 pmol/l) compared to the control group (30).

Another study showed that insulin gene therapy affected IPGTT and that the difference in blood glucose before and after insulin gene therapy in the treat group was 9.54±1.2 mmol/l (15.04±1.6 mmol/l to 5.49±0.4 mmol/l), whereas in the normal control group it was 5.55±0.6 mmol/l throughout the experimental period. Therefore, glucose was significantly decreased without any apparent significant differences in insulin (23). However, Rasouli et al. reported insulin gene therapy by GIP/Ins/pBud increased insulin in comparison to controls (1.048 pmol/l) (24). Similarly, Cheung used GIP/Ins fragments and injected them into pro-nuclei of fertilized mouse embryos. In the transgenic mice mean blood glucose and human insulin was 9.52±1.16 mmol/l and 39±16.9 pmol/l, respectively, which showed decreased blood glucose and raised human insulin (27).

Findings from the meta-analysis on insulin gene therapy and IPGGT

Gene therapy by viral vector

Five studies were identified (21, 22, 28-30), including 28 datasets that met the inclusion criteria based on their mean IPGGT after insulin gene therapy by viral vector that was reduced on average by -12.69 mmol/l (P<0.001) (Figure 2). Publication bias was observed (P=0.007) after using the trim-and-fill method to adjust for funnel plot asymmetry, although these results

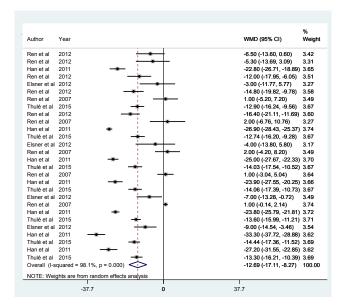


Figure 2. Mean of IPGGT after insulin gene therapy by viral vectors

were unchanged. Between-study heterogeneity was also found (I^2 =98.1, P<0.001). The sensitivity analysis revealed that the exclusion of any single study did not alter the overall effect. For each study, assessment follow-up duration after treatment was classified as \leq 30 min (ID=1), \geq 60 and \leq 90 min (ID=2), and \geq 120 and ≤ 300 min (ID=3). Such subgroup analysis (Figure 3) showed no heterogeneity between studies, although the summary mean for IPGGT after ≥ 120 and ≤ 300 min was lower (-15.46 mmol/l, P<0.001) than achieved at \leq 30 min (-11.12 mmol/l, *P*=0.001) and \geq 60 and \leq 90 (-11.25 mmol/l, P=0.006). The gene delivery method (AAV) (ID=1), r Adeno (ID=2) and lentiviral (ID=3), modified the IPGGT response that was greatest with the r Adeno method, (-25.87 mmol/l, P<0.001) compared to AAV (-13.60 mmol/l, P<0.001) and the lentivirus (-5.17

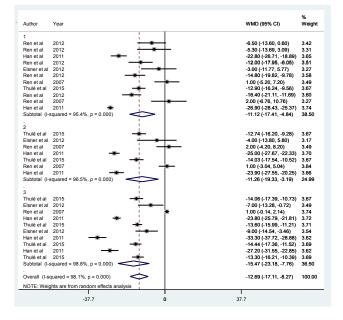


Figure 3. Subgroup analysis by follow-up duration for the mean of IPGGT after insulin gene therapy by viral vectors

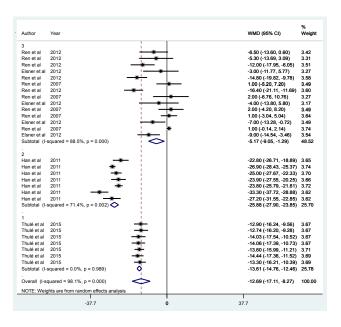


Figure 4. Subgroup analysis by gene delivery methods for the mean of IPGGT after insulin gene therapy by viral vectors

mmol/l, P<0.009) (Figure 4).

Findings from the meta-analysis on insulin gene therapy and FBS:

Gene therapy by viral vector

Eight studies with 15 datasets describing the effects of insulin gene therapy by viral vectors gave a mean reduction in FBS (-13.51 mmol/l) (21, 22, 28, 29, 31-34) (Figure 5). No evidence of publication bias was found (P=0.86). The I² value indicates 96.3% of the variability was accounted for across the pooled estimates. Sensitivity analysis showed the exclusion of each study from the analysis did not change the overall effect. Subgroup analysis by follow-up duration confirmed a prominent effect (P<0.001) of insulin gene therapy on FBS at \leq 5 days (ID=1), \geq 10 and \leq 30 days (ID=2), and \geq 50 and \leq 70 days (ID=3) (Figure 6). Insufficient studies in AAV, retroviral, and viral subgroups meant analysis according to gene delivery method was not possible.

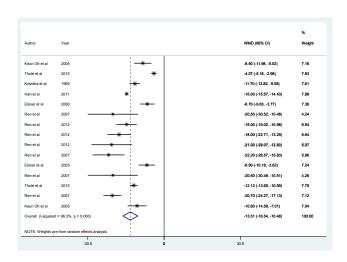


Figure 5. Mean of FBS after insulin gene therapy by viral vectors

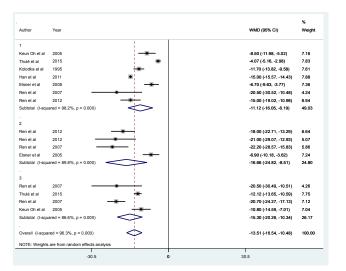


Figure 6. Subgroup analysis by follow-up duration for the mean of FBS after insulin gene therapy by viral vectors

Gene therapy by non-viral vector

Meta-analysis from three studies including 20 datasets that used non-viral vectors (24-26) also showed reduced FBS (-29.95 mmol/l, P<0.001) (Figure 7). Egger's test was significant (P=0.001), but applying trim and fill had no effect on the outcome, as there was between-study heterogeneity (I^2 =98.8, % P=P<0.001). The exclusion of each study from the meta-analysis did not impact the overall sensitivity analysis. According to follow-up duration, studies were categorized into ≤ 1 day (ID=1), ≥ 7 , and ≤ 15 days (ID=2) (Figure 8), and the magnitude of effect increase with time of follow up, i.e., FBS at ≤ 1 days, -16.08 g (P<0.001) compared with ≥ 7 and ≤ 15 days, -112.09 g (P<0.001).

Findings from the meta-analysis on insulin gene therapy and blood insulin

Gene therapy by viral vector

Meta-analysis of 6 studies (21, 31, 32, 34-36) with 20 datasets describing the effects of insulin gene therapy with lentiviral carriers on blood insulin showed a mean

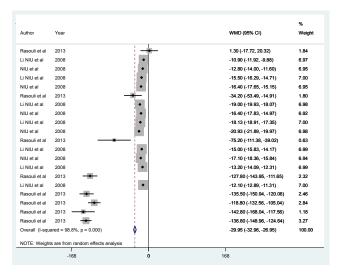


Figure 7. Mean of FBS after insulin gene therapy by non-viral vectors

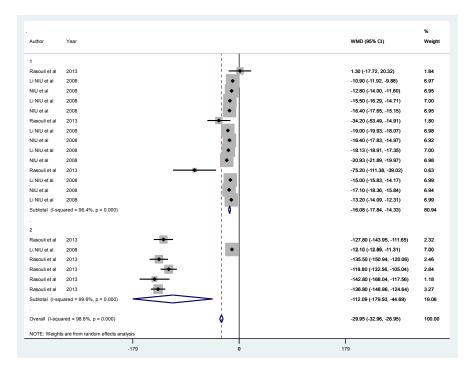


Figure 8. Subgroup analysis by follow-up duration for the mean of FBS after insulin gene therapy by non-viral vectors

increase of 398.3 pmol/l (P<0.001) (Figure 9). There was no publication bias (P=0.06) and overall heterogeneity (I^2 =100%), as well as between-study heterogeneity for the duration of measurement, i.e., ≤ 10 min (ID=1), ≥ 15 and ≤ 30 min (ID=2), ≥ 40 and ≤ 60 min (ID=3), ≥ 90 and ≤ 150 min (ID=4), ≥ 21 , and ≤ 90 days (ID=5) (Figure 10).

Gene therapy by non-viral vector

Random-effects meta-analysis confirmed the effects of insulin gene therapy with non-viral carriers on raised blood insulin by 114.9 pmol/l (P<0.001) (Figure 11). There was no evidence of publication bias (P=0.70), and heterogeneity between studies was high (I²=94.9%),

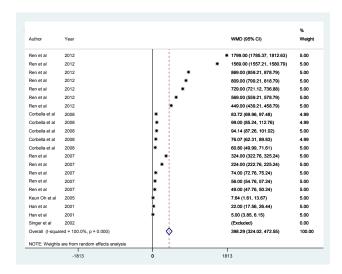


Figure 9. Mean of insulin level after insulin gene therapy by viral vectors

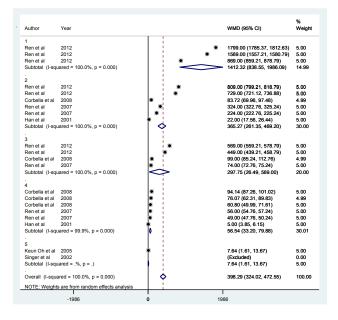


Figure 10. Subgroup analysis by follow-up duration for the mean of insulin level after insulin gene therapy by viral vectors

which was unaffected by one study or follow-up duration (Figure 12).

Findings from the meta-analysis on insulin gene therapy and bodyweight

Gene therapy by viral vector

Gene therapy by viral vectors increased bodyweight (Figure 13) in 5 studies (21, 24, 25, 29, 31) with 11 datasets, which increased by 24.2 g (P<0.001). There was no evidence of publication bias (P=0.45), and between-study heterogeneity was apparent (I²=96.4%,

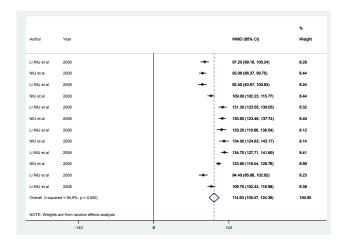


Figure 11. Mean of insulin level after insulin gene therapy by nonviral vectors

P<0.001), with no single study influencing the final effect. Subgroup analysis by follow-up duration showed an increased response with time, i.e., ≤ 5 days (ID=1), 13.3 g, (P<0.01); between ≥ 50 and ≤ 70 days (ID=3), 62.1 g, (P<0.001); but no effect during ≥ 10 and ≤ 25 days (ID=2) (7.09 g P=0.14) (Figure 14). Due to insufficient studies in the AAV and retroviral, subgroup analysis according to gene delivery methods was not performed.

Discussion

Reducing blood glucose in T1DM is necessary to avoid side effects such as neuropathy, glaucoma, nephropathy, and cardiomyopathy (37-40), for which the most popular treatment is insulin injection, although this is not very practical. It can also cause hyperinsulinemia, which is a risk factor for progressive insulin resistance and cardiovascular damage (41, 42). Therefore insulin gene therapy is currently a focus of future T1DM treatment, with the restoration of a dynamic and more precise method of insulin production (43). The different approaches that can be used to more effectively maintain euglycemia are promoting the survival and proliferation

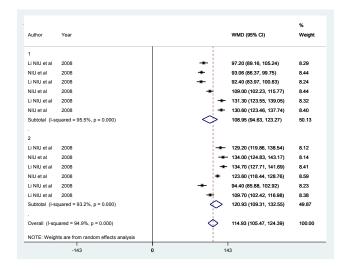


Figure 12. Subgroup analysis by follow-up duration for the mean of insulin level after insulin gene therapy by non-viral vectors

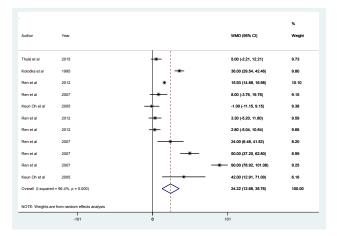


Figure 13. Mean of bodyweight after insulin gene therapy by viral vectors

of islets β cells, preventing their destruction by the immune system, and the employment of non-islets β cells such as hepatocytes, myocytes, fibroblasts, and intestinal and gastric epithelial cells to regulate insulin release (20). Also, gene targeting in T1DM can be achieved using viral or non-viral vectors (43, 44), for which the former is more effective (45). We have performed the first systematic review and meta-analysis to investigate the efficiency of insulin gene therapy for IPGTT, FBS, insulin, and bodyweight.

IPGGT studies on streptozocin (STZ)-induced diabetic mice treated with the insulin gene, delivered intrapancreatically by recombinant Ad (rAD) vector, corrected hyperglycemia and glucose tolerance (31). This response was, however, transient and typically persisted for only 1–3 weeks (20). Studies on STZ-induced diabetic rodents treated with the insulin gene showed that all gene therapies decreased blood glucose and increased insulin. Because lentiviruses and retroviruses are integrative vectors, they can elicit long-term benefits, as shown in rats (29) and mice (21). In both studies, an HMD/INS-FUR construct using INS-

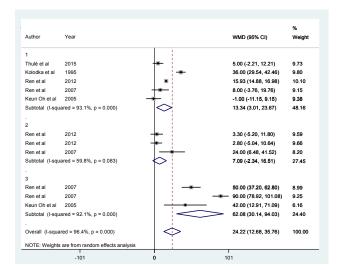


Figure 14. Subgroup analysis by follow-up duration for the mean of weight after insulin gene therapy by viral vectors



FUR was cloned into the site of LV HIV/MSCV (HMD) and injected into the portal vein by intervallic fusion to be delivered into the liver. In STZ-induced diabetic rats, blood glucose was returned to normal for at least 500 days without any adverse response.

One of the other factors associated with insulin gene therapy for T1DM is increased bodyweight, as diabetes improves, as confirmed by our meta-analysis. We also showed significant effects on FBS and blood insulin by non-viral vectors. Until recently, a major limitation of viral insulin therapy has been the lack of mealdependency on insulin secretion in these surrogate cells. K-cells are native endocrine cells that are glucoseresponsive native endocrine cells, located primarily in the stomach, duodenum, jejunum, and gut hormone GIP (46), which normally potentiates postprandial insulin release (27). It has therefore been proposed that K-cells may be suitable targets for T1DM insulin gene therapy (47), although they have a short lifespan of 3-5 days, which necessitates frequent and repeated gene administration. Taking all these studies together, we observed substantial heterogeneity due to animal type, sample volume, the method of determining T1DM and the gene delivery method. Therefore, the random effect model was used to reduce these contrasting effects but could not find the sources of the heterogeneity, which suggests that the efficacy of insulin gene therapy is variable. In spite of these limitations, including publication bias, our study had several strengths, being the first meta-analysis focused on the effects of insulin gene therapy on T1DM related factors.

Conclusion

The meta-analysis findings showed a significant effect for insulin gene therapy and T1DM related factors, including IPGTT, fasting blood glucose, insulin, and bodyweight in diabetic rodents.

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Conflicts of Interest

The authors declare that there are no conflicts of interest.

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